TAM receptors in cardiovascular disease

Lucy McShane (1)^{1,2}, Ira Tabas (1)³, Greg Lemke (1)^{4,5}, Mariola Kurowska-Stolarska⁶*, and Pasquale Maffia (1)^{1,2,7}*

¹Centre for Immunobiology, Institute of Infection, Immunity and Inflammation, College of Medical, Veterinary and Life Sciences, University of Glasgow, Sir Graeme Davies Building, 120 University Place, Glasgow G12 8TA, UK; ²Institute of Cardiovascular and Medical Sciences, College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, Glasgow, UK; ³Departments of Medicine, Physiology, and Cell Biology, Columbia University Irving Medical Center, New York, NY, USA; ⁴Molecular Neurobiology Laboratory, Salk Institute for Biological Studies, La Jolla, CA 92037, USA; ⁵Immunobiology and Microbial Pathogenesis Laboratory, Salk Institute for Biological Studies, La Jolla, CA 92037, USA; ⁶Rheumatoid Arthritis Pathogenesis Centre of Excellence (RACE), Institute of Infection, Immunity and Inflammation, College of Medical, Veterinary and Life Sciences, University of Glasgow, Sir Graeme Davies Building, 120 University Place, Glasgow G12 8TA, UK; and ⁷Department of Pharmacy, University of Naples Federico II, Naples, Italy

Received 4 January 2019; revised 28 February 2019; editorial decision 14 March 2019; accepted 9 April 2019; online publish-ahead-of-print 13 April 2019

This article was handled by a Consulting Editor, Ziad Mallat.

Abstract	The TAM receptors are a distinct family of three receptor tyrosine kinases, namely Tyro3, Axl, and MerTK. Since their discovery in the early 1990s, they have been studied for their ability to influence numerous diseases, including cancer, chronic inflammatory and autoimmune disorders, and cardiovascular diseases. The TAM receptors demonstrate an ability to influence multiple aspects of cardiovascular pathology via their diverse effects on cells of both the vasculature and the immune system. In this review, we will explore the various functions of the TAM receptors and how they influence cardiovascular disease through regulation of vascular remodelling, efferocytosis and inflammation. Based on this information, we will suggest areas in which further research is required and identify potential targets for therapeutic intervention.
Keywords	TAM receptors • Tyro3 • Axl • MerTK • Cardiovascular disease

1. Introduction

Insight from the results of studies using cultured cells, mouse models of cardiovascular disease subjected to genetic engineering or pharmacological intervention, and observational studies in humans have provided strong evidence that the immune responses plays an important role in cardiovascular pathologies.^{1–3} Most importantly, the CANTOS trial (Canakinumab Anti-inflammatory Thrombosis and Outcomes Study) has recently demonstrated the efficacy of targeting interleukin (IL)-1 β for reducing secondary cardiovascular events,⁴ and therefore, much of current research is now focusing on how to limit inflammation to prevent cardiovascular diseases (CVD).

Among the many molecules that influence the immune response, the TAM family of tyrosine kinase receptors have demonstrated a capacity to influence the function of both the vascular and immune system in the steady state and in pathology. Accordingly, there has been much interest in their potential roles in CVD and how they may be viewed as potential therapeutic targets. This family of proteins includes Tyro3, Axl, and MerTK, with the first letters of each giving the family its name.⁵ They remained orphan receptors for the first few years following their

discovery, but by the mid-1990's their ligands were identified as growth arrest-specific 6 (Gas6) and Protein S (Pros1).^{6,7} These ligands bind to the TAM receptors with differential affinity. Gas6 can associate with all three receptors, but with strongest affinity to Axl, then Tyro3, and with lower affinity to MerTK.⁶ Pros1, however, does not bind to Axl at all, and has stronger affinity binding with Tyro3 than MerTK.⁸

2. TAM receptors

2.1 Structure

The basic structure of the TAM receptors includes an extracellular Nterminal region containing two immunoglobulin (Ig)-like domains, followed by two fibronectin type III (FNIII) domains, a hydrophobic domain which traverses the cell membrane, and finally, an intracellular tyrosine kinase C-terminal domain.^{9,10} The Gas6 and Pros1 ligands possess an approximately 50 amino acid stretch which contains gamma carboxylated glutamic acid residues, referred to as the Gla-domain. These residues have a high affinity for calcium, which facilitates binding to phosphatidylserine (PtdSer) molecules found on the surface of platelets and on the

*Corresponding authors. Tel: +44 141 330 7142; E-mail: Pasquale.Maffia@glasgow.ac.uk (P.M.) Tel: +44 141 330 6085; E-mail: Mariola.Kurowska-Stolarska@glasgow.ac.uk (M.K.-S.)
© The Author(s) 2019. Published by Oxford University Press on behalf of the European Society of Cardiology.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

outer leaflet of cell membranes under certain conditions, notably on apoptotic cells.¹¹ The C-terminus of the ligands possess laminin G (globular) domains, which facilitate their interactions with the TAM receptor Ig-like domains.¹² As for most receptor tyrosine kinases (RTKs), activation of the TAM receptors occurs via ligand facilitated dimerization, which mediates autophosphorylation of their tyrosine kinase domain.⁹ This results in coupling of the receptor with proteins involved in signal-ling pathways, which will be discussed in more detail below.

2.2 TAM receptor functions

Remarkably, TAM receptors are not required for embryonic development, which is unusual for one RTK, let alone an entire subset.¹³ However, this property enabled the generation of viable triple TAM knock-out (KO) mice, which propelled studies exploring the functions of the TAM receptor family. When these KO mice reach adulthood, three distinct phenotypes can be observed. The first is male infertility, which occurs due to an inability to clear apoptotic gamete cells in the testes.¹³ The second phenotype is blindness due to retinal epithelial cells not being able to engulf the outer segments of the photoreceptors, which is necessary for the removal of toxic byproducts of phototransduction.¹⁴ The third phenotype is autoimmunity.^{15–18} This stems from the requirement to clear the apoptotic bodies generated during the immune response in order to resolve inflammation.¹⁹ Failure to clear dead cells can cause them to become necrotic, with their accumulation serving as a source of self-antigen.²⁰

Phagocytosis of apoptotic cells (efferocytosis) is a fundamental process for the restoration of immune and tissue homeostasis,²¹ and the TAM receptors play an important role in efferocytosis in adult tissues.^{22,23} In atherosclerosis, efferocytosis is required for clearing of apoptotic cells in lesions, and in advanced atherosclerosis, this process can go awry, leading to post-apoptotic necrosis of lesional cells.^{24–28} This pathological process can lead to large areas of plaque necrosis, which are highly inflammatory and render the plaques susceptible to rupture or erosion. Ruptured or eroded plagues can then promotes occlusive vascular thrombosis, leading to acute coronary syndromes such as myocardial infarction (MI), unstable angina, sudden cardiac death, or stroke.²⁹ In atherosclerosis, efferocytosis is largely orchestrated by professional phagocytes such as macrophages, which are known to express and utilize TAM receptors in this process.^{30–32} In other settings, it is likely that the non-professional phagocytes including cardiac myofibroblasts and epithelial cells also express TAM receptors,^{33,34} particularly Axl which shows high levels of expression in the heart.^{35,36}

In addition to the anti-inflammatory effects of efficient efferocytosis, the TAM receptors are involved in directing the change in the immune response from attack-the-pathogen to repair-and-restore (resolution).^{16,37} This was first indicated by the phenotype of the triple TAM receptor KO mice, which develop a lymphoproliferative disorder and broad-spectrum autoimmunity driven by chronic hyper-activation of antigen presenting cells such as monocytes, dendritic cells (DCs), and macrophages.¹⁵

Expression of TAM receptors is up-regulated in innate immune cells upon their activation in order to prime the system for negative feedback, which is subsequently facilitated by the increased availability of their ligands upon initiation of the adaptive response.¹⁰ Upon ligand-mediated autophosphorylation, TAM receptors can physically associate with the type I interferon receptor (IFNAR)-STAT1 complex, which normally drives the initial amplification of inflammation. However, the association between the R1 subunit of IFNAR and the phosphorylated TAM causes a change in the function of the IFNAR-STAT1 complex to that of an anti-inflammatory signalling molecule, which in turn initiates transcription of the suppressor of cytokine signalling (SOCS)1 and SOCS3 proteins.¹⁶ These proteins ultimately suppress both cytokine receptor and Toll-Like receptor (TLR)3, TLR4, and TLR9 pro-inflammatory signalling pathways.³⁸ This relationship between TAM receptors and ligands is key to maintaining immune homeostasis, particularly between T cells, which produce Pros1 upon activation, and DCs expressing the TAM receptors.³⁹ T cell-DC interactions have been shown to occur within atherosclerotic lesions, which can enhance the pro-inflammatory nature of the plaque.^{40–42} Therefore, deregulation of TAM-mediated suppression and resolution of the inflammatory response can strongly influence cardiovascular immunity.

2.3 Regulation of TAM receptor expression

The expression and activity of the TAM receptors are controlled by various factors at the transcriptional, post-transcriptional, and protein levels. Induction of specific TAM receptor genes can be mediated by various cytokines. For example, transforming growth factor β (TGF- β), and granulocyte-macrophage colony-stimulating-factor, and several proinflammatory stimuli can drive Axl expression, while IL-17A, IL-10, and dexamethasone drive MerTK expression.^{43–46} As Tyro3 has not been closely studied in the context of inflammation or the immune response, it remains unclear which cytokines, if any, specifically up-regulate its expression.

Up-regulation of MerTK expression can also be induced by glucocorticoids and the liver-X-receptors (LXR) α and $\beta.^{47,48}$ TAM receptor regulation by the LXRs is of interest in CVD due to their role in cholesterol biosynthesis and homeostasis, notably cholesterol efflux from macrophages.^{49}

As an example of another level of regulation, microRNA-34a has been demonstrated to suppress protein expression of Axl, and this process can contribute to the autoimmune disorder rheumatoid arthritis.⁵⁰ Similarly, microRNA-7 inhibits Tyro3 expression and is consequently being explored as an RNA-based therapeutic for treating abhorrent Tyro3 overexpression in human hepatocellular carcinoma.⁵¹

Once TAM receptors are fully synthesized, inserted into the plasma membrane, and activated, their extracellular domain can be cleaved by the metalloprotease A Disintegrin And Metalloproteinase (ADAM)-17.^{52,53} This cleavage liberates a soluble extracellular domain bound to ligand (sAxl or sMer) and destroys TAM receptor function. Conversely, secretory leucocyte protease inhibitor increases the expression of MerTK on the cell surface, likely by inhibiting its cleavage.⁵⁴ The molecular regulation of TAM receptors is illustrated in *Figure 1*.

3. Axl in CVD

3.1 Gas6-Axl and regulation of cardiovascular remodelling

Vascular remodelling refers to a dynamic process that causes changes to the structure of the vascular wall. This can occur in response to certain stimuli, such as injury or local production of inflammatory mediators. In addition, chronic conditions such as *hypertension* or *atherosclerosis* can drive the process of vascular remodelling. Although vascular remodelling likely represents a response to correct environmental changes to vascular flow and maintain homeostasis, it can have pathological effects, as described below. Several vascular cell types, including endothelial cells (ECs), vascular smooth muscle cells (VSMCs), fibroblasts and myofibroblasts, contribute to vascular remodelling. Vascular remodelling occurs



Figure 1 Molecular regulation of the TAM receptors. The expression and activity of the TAM receptors is controlled by various factors at the transcriptional, post-transcriptional, and protein levels. (A) TAM receptor gene transcription can be up- or down-regulated by various factors, including cytokines. The figure shows mediators that specifically regulate MerTK transcription; the other TAM receptors are regulated by other mediators.^{43,44,46–48} (B) Post-transcriptional regulation by micro-RNAs such as miR-34a inhibition of Axl expression.⁵⁰ (C) At the protein level, TAM receptors are rendered dysfunctional by cleavage of their extracellular domain by metalloprotease ADAM17.^{52,53} This process which can be driven by other environmental factors such as reactive oxygen species (ROS).^{53,55,56} (D) The soluble byproduct released may act as a decoy for the receptor ligands, thus inhibiting TAM receptor activity.⁵⁷ (E) Activation of the receptors can also be enhanced by various environmental factors.^{58–60} (F) Activation of the TAM receptors subsequently induces various molecular pathways affecting cell function.^{16,60–62}

through four main processes: cellular migration, proliferation, survival; and extracellular matrix synthesis and degradation. For each of these processes, Axl and its ligands have been shown to play a role, while the contributions of MerTK and Tyro3 are less clear.

Both Pros1 and Gas6 were shown to be secreted by and enhance the proliferation of VSMCs prior to their identification as the ligands for TAM receptors.^{58,63} Subsequently, Axl expression was found to be increased in VSMCs following balloon-injury in rat carotid artery, and both

Axl and Gas6 expression were temporally correlated with neointima formation.⁶⁴ The key role of Gas6/Axl pathway in the regulation of vascular remodelling was confirmed in animal studies, which showed reduced intimal thickening following vascular injury in Axl^{-/-} mice compared with wild type control mice.^{59,65,66} Similarly, in the deoxycorticosterone acetate (DOCA)-salt hypertensive mouse model, Axl deficiency led to reduced systolic blood pressure^{67,68} and reduced remodelling index of the mesenteric artery.⁶⁸ Emerging studies suggest that (i) induction of proliferation of VSMCs,^{58,63} (ii) stimulation of migration of VSMCs,⁶⁹ and (iii) protection from apoptosis^{70,71} are underlying mechanisms linking Gas6/Axl signalling to vasculature remodelling.

Angiotensin II (Ang II) is a hormone produced as an end-product of the renin-angiotensin system and is an important mediator of numerous cardiovascular pathologies, such as hypertension, vascular remodelling, and neointima formation.⁷² Ang II has been demonstrated to activate the Gas6-Axl pathway and is necessary for facilitating the effects of Axl on VSMC proliferation.^{73,74} In a similar vein, reactive oxygen species (ROS) such as hydrogen peroxide (H_2O_2) elicit pathological effects on the vasculature,⁷⁵ at least partially through Axl in VSMCs.⁵⁹ ROS have been shown to induce interaction between Axl and glutathiolated non-muscle myosin heavy chain (MHC)-IIB, which may mediate increased migration in vascular injury,⁷⁶ and pharmacological inhibition of Axl attenuated the pathological effects of oxidative stress and reduced VSMC migration in vitro.⁷⁷ In addition, AxI appears to prolong VSMC survival, and Gas6 was shown to protect VSMCs from apoptosis.^{70,71} In this context, one study observed increased VSMC apoptosis in Axl deficient mice, leading to reduced intima-media thickening following ligation of the left carotid artery.66

In addition to regulating VSMC biology, activation of Axl via Gas6 exerted a mitogenic effect on serum-starved fibroblasts *in vitro* and protected these cells from cell death by apoptosis.⁷⁸ Similarly, Gas6-Axl signalling can promote the survival and vascular endothelial growth factor Amediated migration of ECs.^{79–83} Finally, Axl has also been shown to directly regulate cytokine/chemokine expression and extracellular matrix remodelling in the vessel wall.⁸⁴ Axl regulates these multiple aspects of vascular function through its ability to interact with multiple downstream signalling pathways. For example, Axl-mediated effects on cell proliferation and migration are facilitated through activation of phosphatidylinositol-3-OH kinase (PI3K)/protein kinase B (Akt), sarcoma (SRC) signalling pathways, and extracellular signal-regulated kinases (ERKs), which is similar to other RTK-mediated processes.^{60,61}

Remarkably, differing levels of glucose in vitro can affect Axl behaviour, demonstrated by the comparison of functional Axl interactions in VSMCs exposed to 5.5 mmol/L 'low glucose' or 27.5 mmol/L 'high glucose' culturing conditions.⁶⁰ Axl was found to preferentially interact with proteins involved in the PI3K signalling pathway under 'low glucose' conditions, stimulating anti-apoptotic signalling and enhancing survival of the VSMCs. In contrast, Axl associated with signalling proteins of the ERK1/2 pathway in 'high glucose' conditions, driving VSMC migration. Ultimately, this demonstrates that the function of Axl may be altered depending on the physiological conditions.⁶⁰ In addition to this, Axl expression is significantly lower in left internal mammary artery tissue from diabetic compared with non-diabetic patients⁸⁵ and Axl overexpression was able to reverse the effects of high glucose-induced dysfunction in ECs in vitro.⁸⁵ The precise mechanisms by which Axl signalling and function respond to varying glucose levels remains unclear. However, these findings may have important implications in terms of understanding the pathological effects of chronic high blood glucose levels in diabetic patients.

Vascular calcification, which is the process by which calcium builds up within the vasculature, is particularly prevalent in advanced, inflammatory atherosclerosis and correlates with worse clinical outcomes.^{86–88} Conversely, absence of calcification in coronary arteries predicts a low risk of CVD even in subjects with a high level of traditional risk factors.⁸⁹ During calcification, vascular pericytes undergo a process of osteogenic differentiation, and Axl was identified as one of the genes which is down-regulated when this process was explored *in vitro*.⁹⁰ Similarly, Axl

1289

expression was shown to be down-regulated as cultured VSMCs calcify their matrix, and Axl overexpression or activation inhibited calcification *in vitro*.^{91,92} More recently, miR-34a has been shown to promote VSMC calcification in mice and in VSMCs *in vitro*, with the *in vitro* effect of miR-34a showing a correlation with decreased Axl expression on the cultured VSMCs.⁹³ Interestingly, work by another group has revealed that the ability of hydroxy-3-methylglutaryl coenzyme A (HMG CoA) reductase inhibitors (statins) to prevent phosphate-induced calcification by VSMCs *in vitro* occurs via restoration of the Gas6-Axl mediated survival pathway.^{94,95} Whether Axl affects vascular calcification *in vivo* has not yet been determined.

Axl has also been reported to be expressed by cardiomyocytes.³⁶ In patients with heart failure Axl levels are amplified both in terms of the cardiac tissue and circulating soluble Axl (sAxl).³⁵ Furthermore, Gas6 and sAxl levels are found to increase in patients following ST-segment elevation MI.⁹⁶ In both studies, the levels of Axl were predictive of adverse pathology, such as the extent of left ventricular remodelling and of further cardiovascular events. One study tested the effect of both KO and cardiac-specific overexpression of Gas6 in a cardiac stress murine model.⁹⁷ They found that Gas6-deficient mice had decreased hypertrophy, fibrosis, and contractile dysfunction in the chronic stress overload induced by aortic banding. Whereas cardiac-specific overexpression of Gas6 enhanced these pathologies. This was ultimately attributed to Gas6 activation of the ERK signalling pathway, driving cardiac hypertrophy. Interestingly, this process was reversed with the use of a pharmacological inhibitor of ERK. As discussed previously, Gas6 activation of Axl can induce the ERK signalling cascade,⁹⁸ which combined with evidence that Axl levels are elevated in heart failure patients, points to the Gas6-Axl axis as a potential novel therapeutic target in heart failure.

In summary, multiple studies suggest that GAS6/Axl drives cardiovascular remodelling by regulating the biology of VSMCs, ECs, cardiomyocytes, and potentially fibroblasts, thereby facilitating pathological processes such as neointima formation, and remodelling in both the heart and vasculature. Most importantly, Axl suppression can dampen these adverse effects, suggesting possible therapeutic implications of these studies.

3.2 Gas6-Axl and inflammation in CVD

Axl performs biphasic roles in the cells of the vasculature and immune system. Axl expression has actually been suggested to drive proinflammatory activation of VSMCs during vein-graft remodelling.⁶⁵ In this study, vein-graft surgeries were performed to examine the effect of Axl deficiency in both the vein graft donor and recipient mouse to compare the effect of vascular vs. systemic expression of Axl. The authors found that Axl depletion in all groups led to lower levels of MHC class II expression in the vein graft, indicating lesser immune activation. They also observed down-regulation of various pro-inflammatory cytokines and chemokines in $Axl^{-/-}$ SMCs compared with $Axl^{+/+}$ with and without interferon gamma (IFN- γ) stimulation. Remarkably, they found Axl deficiency to increase the expression of SOCS1 in VSMCs, i.e., opposite to what is observed in immune cells.^{10,16,50}

The transfer of Axl deficient haematopoietic myeloid cells to western diet-fed low-density lipoprotein receptor KO ($Ldlr^{-/-}$) mice had no effect on atherosclerotic disease progression.⁹⁹ To date no studies have examined the effect of global Axl deficiency on atherosclerosis pathology, and thus the effect of eliminating Axl expression in the vasculature has not been addressed. This remains an important point for study, as Axl has been shown to be present in human vessels with expression down-regulated in atherosclerotic plaques compared with normal carotids.¹⁰⁰

Furthermore, Axl expression is higher in vessels that are less prone to develop atherosclerosis such as the left internal mammary artery compared with the aorta.¹⁰¹ Concomitantly, elevated serum levels of sAxl have been detected in the setting of acute coronary syndromes.¹⁰² The Axl ligand Gas6 is expressed by ECs, VSMCs, and macrophages and its expression increases with atherosclerosis development.^{103,104} Genetic KO of Gas6 increases plaque stability in *ApoE^{-/-}* mice, leading to increased plaque content of SMCs and collagen and to reduced numbers of macrophages.¹⁰⁵ However, data from an epidemiological studies showed that low levels of Gas6 correlate with increased risk of coronary artery disease (CAD) in patients with psoriasis.¹⁰⁶

An interesting study used chimeric mice with haematopoietic or nonhaematopoietic Axl deficiency to dissect the role of Axl expression in immune vs. vasculature cells in hypertension.⁶⁷ The data indicated that Axl-expressing immune cells drove pro-inflammatory gene expression and increased immune cell infiltration in the kidney at early stages of hypertension and that Axl expression in both immune and vascular cells was detrimental in the later phases of hypertensive disease. Continuing on from this, the same group demonstrated that Axl is critical for survival of T lymphocytes, affecting vascular remodelling and inflammation in DOCA-salt induced hypertension.¹⁰⁷

Axl has been suggested to influence natural killer (NK) cell development. However, this conclusion is controversial, with studies supporting a role for Axl in both the suppression and promotion of NK cells.^{108–110} Although none of these studies are focused on cardiovascular disease, this could be an interesting area of future study. On the one hand, NK cells may have a potentially protective role in CAD based on strong correlations between coronary heart disease and low NK levels in humans.^{111,112} However, animal studies suggest that NK cells are atherogenic.¹¹³ The role of Axl in NK cells and the role of NK cells themselves in cardiovascular disease represent areas that will benefit from further studies, which is necessary for a better understanding of Axl's role in vascular pathology and its potential as a therapeutic target.

4. MerTK in CVD

4.1 MerTK-mediated efferocytosis in atherosclerosis

MerTK deficiency leads to enhanced pathology in mouse models of atherosclerosis.^{30,32} One study backcrossed MerTK KOs into $ApoE^{-/-}$ mice, which were maintained on western diet for either 10 or 16 weeks to examine the differential effect of MerTK deficiency on early or advanced lesions.³² The pathological effects of MerTK-deficiency were apparent in advanced, as opposed to early, lesions owing to a marked decrease in the clearance of apoptotic bodies and the subsequent accelerated plaque necrosis. Interestingly, the study found no apparent effect on overall lesion size or plasma cholesterol profile, despite other studies showing that effective efferocytosis is beneficial early on in lesion development.^{25–27} Thus, other phagocytic receptors may play a role in effero-cytosis in early atherosclerosis, although Axl is not likely among these receptors, as mentioned above, the transfer of Axl deficient bone marrow cells to $Ldlr^{-/-}$ mice had no effect on atherosclerotic disease progression.⁹⁹

The effects of MerTK in atherosclerosis progression to date has been investigated in phagocytic immune cells—primarily macrophages. This can be inferred from a study which utilized a chimeric mouse model, in which bone marrow cells deficient in MerTK were transferred into *Ldlr^{-/-}* mice.³⁰ Similar to the previous study, MerTK deficiency lead to

significantly higher levels of apoptotic debris accumulation within the plaque after 15 weeks of western diet. The authors were also able to show increased inflammation in the lesions of these mice. In this study, there was a 60% increased lesion size in the $Mertk^{-/-}Ldlr^{-/-}$ mice compared with $Mertk^{+/+}Ldlr^{-/-}$. It is possible the differences in effect on lesion size may be due to differences between the mouse models and/or composition of the diets used in the two studies. More recently, a macrophage Ca²⁺/calmodulin-dependent protein kinase II γ (CaMKII γ) pathway was shown to play a key role in the development of necrotic atherosclerotic plaques by preventing MerTK expression through the inhibition of the transcription factors ATF6 and LXR α .¹¹⁴

MerTK is expressed in macrophages in human atherosclerotic arteries,¹⁰⁰ and MerTK induction is required for clearance of apoptotic cells by human macrophages.¹¹⁵ Interestingly, it has been suggested that macrophage MerTK deficiency can occur near the necrotic cores of human plaques via the action of ADAM17-mediated MerTK cleavage.⁵⁵ ADAM17 can be activated by the byproducts of polyunsaturated fatty acid oxidation and by inflammatory mediators, which are known to be present within the necrotic core of atherosclerotic plaques.^{53,55} Levels of sMer within individual plaques were shown to correlate with the extent of necrosis, and mice expressing genetically modified MerTK resistant to cleavage (MerTK^{Δ 483-488}) showed improved lesional efferocytosis, more stable plaques, and, interestingly, improved resolution of inflammation³¹ (see below).

Although research in this area has mainly focused on the role of MerTK in macrophages, it should also be noted that brain microvascular ECs have been shown to express MerTK,¹¹⁶ and that it is required for tightening the blood–brain barrier during viral infection.¹¹⁷ In atherosclerosis, inflammation and oxidative stress can destabilize the endothelial barrier, contributing to pathology.¹¹⁸ Therefore, MerTK, by maintaining the integrity of the endothelial barrier, may in principle also contribute to impeding the development of atherosclerosis; however, further work needs to be done to explore this aspect.

4.2 MerTK-mediated efferocytosis in myocardial infarction

Another important facet of cardiovascular pathology that relies on efficient efferocytosis, includes the clearing of dead cardiomyocytes following MI.^{119–121} This process is orchestrated by various immune cells, and the inflammatory profile of these cells can have a major influence on the functional outcome and subsequent progression of heart failure.¹²² MerTK-expressing monocyte/macrophages are key for the clearance of injured cardiomyocytes and improve remodelling following MI in mice.¹²³ Conversely, genetic deficiency of MerTK led to an increase in the accumulation of apoptotic cardiac cells following experimental MI, resulting in larger infarct sizes and worse cardiac functional outcomes.¹²⁴ Cardiac extracts from the *Mertk*^{+/+} control mice showed the presence of sMer following MI, which is likely due to the presence of post-MI ADAM17-activating factors that promote MerTK cleavage.¹²⁵

In a study investigating adverse effects of post-MI ischaemia–reperfusion (IR) in mice and humans using therapeutic interventions to restore blood-flow, such as coronary stents and thrombolytic agents, ^{126,127} serum levels of sMer were found to be elevated.⁵⁶ In the mouse models, this finding correlated with lower expression of intact MerTK on the surface of cardiac macrophages. The study went on to show that mice possessing genetically modified cleavage-resistant MerTK (MerTK^{Δ 483-488}) displayed improved levels of efferocytosis, reduced infarct size, and improved cardiac function following IR. It was concluded that the crucial

role of MerTK in facilitating phagocytic clearance of cardiac debris following MI was hindered by IR-induced MerTK cleavage. They hypothesized that the trigger for this was recruitment of monocytes from the circulation, which they tested by treating mice with an antagonist to block C-C chemokine receptor type 2 (CCR2), a chemokine receptor expressed by infiltrating circulatory monocytes, but not in cardiacresident macrophages. This treatment improved post-IR infarct size in $Mertk^{+/+}$ but not $Mertk^{-/-}$ mice, suggesting that CCR2-mediated infiltration negatively affects ability of MerTK to effectively drive phagocytic repair following IR. The exact mechanism for CCR2-dependent MerTK cleavage is still ambiguous. One suggestion is activation of ADAM17mediated cleavage of MerTK by CCR2⁺ monocyte production of ROS.^{53,55,56}

Interestingly, dying cardiomyocytes in the setting of MI promote the release of MerTK from the surface of macrophages, which prevents engulfment of dying cardiomyocytes.¹²⁵ The mechanism involves upregulation on cardiomyocytes of the 'don't-eat-me' marker CD47 following MI.¹²⁸ Although this process during MI is pathologic, as uncleared apoptotic cardiomyocytes drive cardiac inflammation and further cardiomyocyte death, it should be noted that in non-pathological conditions, evading efferocytosis may preserve cardiomyocyte numbers in view of their normally low regenerative capacity. In the pathologic setting of MI, treatment with anti-CD47 subsequently enhanced cardiomyocyte phagocytosis and reduced infarct size.¹²⁸ Accordingly, further investigation into the mechanism by which cardiomyocytes facilitate MerTK shedding and how this can be prevented may present a novel target for therapeutic intervention in improving post-MI recovery.

4.3 MerTK mediated resolution of inflammation in atherosclerosis

The MerTK studies in the Apoe^{-/-} and chimeric Ldlr^{-/-} atherosclerosis models found a heightened inflammatory pathology associated with loss of MerTK.^{30,32} Ait-Oufella et al. showed that ex vivo cultured splenocytes from *Mertk^{-/-}* mice had increased production of pro-inflammatory cytokines IFN- γ , IL-12, and tumour necrosis factor (TNF)- α and decreased production of anti-atherogenic IL-10, showing this phenotype to be inherent in these cells and not just a consequence of the defective apoptotic debris clearance. Mice expressing cleavage-resistant MerTK generate increased levels of specialized pro-resolving mediators such as TGF- β and IL-10 and lipid mediators such as resolvins in atherosclerotic lesions,^{31,37} which, together with enhanced efferocytosis, contribute to attenuation of cardiovascular pathology in atherosclerosis. In terms of lipid mediators, when MerTK engages a ligand, a particular tyrosine residue in the cytoplasmic tail of MerTK signals inhibition of a calcium/ CaMKII pathway that normally favours the biosynthesis of long-chain unsaturated fatty acid-derived inflammatory mediators, notably leukotrienes, over resolution mediators such as lipoxins and resolvins.^{37,62} Thus, MerTK signalling increases the ratio of pro-resolving-to-proinflammatory lipid mediators.

Activation of LXR expression occurs as a result of apoptotic cell clearance, particularly in the presence of excess-lipoprotein derived cholesterol, as would be expected within advanced atherosclerotic lesions.¹²⁹ This process has been shown to promote expression of MerTK, which mitigates pro-inflammatory cytokine release upon subsequent exposure to cholesterol-loaded apoptotic macrophages.^{24,130} In addition, human protein S inhibits the expression of macrophage scavenger receptor A through MerTK, leading to reduced uptake of modified lipoproteins.¹³¹ Therefore, MerTK appears to sit at the interface between lipid metabolism and inflammation within the plaque and functions to attenuate inflammation in this environment.

5. Tyro3 in CVD

Apart from a study showing an association between Tyro3 and MerTK variants and carotid atherosclerosis,¹³² the net contribution of Tyro3 to CVD has not been addressed to date. This may be because Tyro3 is mainly localized to the central nervous system and reproductive organs, and doesn't show high levels of expression in the vasculature.^{36,133,134} It's ligand Pros1 has been found to correlate with coronary heart disease risk and is expressed in atherosclerotic lesions.^{100,135}

Interestingly, Tyro3 is believed to negatively regulate T helper type 2 (Th2) cells via the suppression of a specific subset of CD11c⁺ DCs expressing programmed cell death protein 2 (PDL2).¹³⁶ PDL2⁺ DCs are associated with driving Type 2 (Th2-driven) immune responses, and expression of Tyro3 in these cells was shown to decrease their Th2associated molecule production. This pathway functions as part of a negative feedback loop, whereby Th2-associated cytokine IL-4 induces sustained expression of Pros1, which then activated Tyro3-mediated suppression of Th2 activation. In terms of relevance to CVD, while IL-4 and IL-5 have been shown to be atheroprotective, ^{137,138} IL-9 may be pro-atherogenic.¹³⁹ IL-13 production exerts adverse effects on cardiovascular pathology by driving fibrosis of heart tissue in the context of both ageing and IR injury,^{140–142} but IL-13 secreted by regulatory T cells may actually promote efferocytosis in atherosclerotic lesions.¹⁴³ These combined observations provide a strong rationale for future studies on the role of Tyro3 in CVD.

6. Conclusions and future questions

The TAM receptors—particularly MerTK and Axl—appear to have very different roles in suppressing or driving elements of cardiovascular pathology (*Figure 2*). In terms of what is currently known, MerTK is fundamentally protective in its role. This is mediated by distinct but complementary functions: suppression of the inflammatory response, enhancing the inflammation resolution response, and facilitating the clearance of apoptotic cell debris. By these criteria, as well as the protective role of MerTK in the heart itself, enhancing MerTK synthesis or functioning, and/or blocking its cleavage, may represent novel therapeutic approaches to CVD. However, this approach must consider the possible adverse effects of heightened MerTK function on cancer^{144,145} and possibly pathologic liver fibrosis.¹⁴⁶

There is controversy within the literature in terms of a protective vs. a detrimental role of Axl in CVD. This largely stems from apparent differences in Axl function, particularly in terms of activating inflammation and the cell type in which it is expressed. While there is a molecular signalling pathway by which Axl has been shown to suppress the immune response, the role of this anti-inflammatory pathway in vascular cells is controversial.^{16,65} On the other hand, Axl signalling may be protective against vascular calcification.^{90,91,95} Therefore, future work will be required to sort out the mechanisms that facilitate Axl's opposing roles in vascular and immune cell types and how these roles affect overall cardiovascular pathology.

Tyro3 remains to be the least studied of the TAM receptors, particularly in the area of cardiovascular disease. Other than one study which found an association of a Tyro3 SNP in carotid atherosclerosis¹³²





we could find no published work to date investigating the role of Tyro3 in the context of cardiovascular pathology. However, Tyro3 may possess a protective function in its ability to suppress of type 2 immune responses, which promotes cardiac fibrosis.^{140–142} Thus, with further work in this area, it is possible that targeting Tyro could be considered as a fibrosis-preventing therapy post-cardiac injury.

Finally, there are studies suggesting the Gas6/TAM signalling pathway is essential for platelet activation and thrombus stabilization.

For example, mice deficient in Gas6 or TAM receptors or treated with inhibitors of Gas6/TAM receptor pathways develop less arterial and venous thrombosis. The GAS6/TAM receptor role in haemostasis and thrombosis has been recently fully reviewed elsewhere¹⁴⁷ and is therefore not the focus of the current review. Nonetheless, this role of TAM receptors needs to be considered in view of the key contribution of thrombosis to atherosclerosis and its clinical complications.

Conflict of interest: none declared.

Funding

This work was supported by the British Heart Foundation [PG/12/81/29897 to P.M., RE/13/5/30177, and FS/16/55/32731]; the Engineering and Physical Sciences Research Council (EPSRC) [EP/L014165/1 to P.M.]; the European Commission Marie Skłodowska-Curie Individual Fellowships [661369 to P.M.]; the Arthritis Research UK [RACE20298 to M.K.S.]; and National Institutes of Health [HL075662, HL127464, and HL132412 to I.T.].

References

- Harrison DG, Guzik TJ, Lob H, Madhur M, Marvar PJ, Thabet S, Vinh A, Weyand C. Inflammation, immunity and hypertension. *Hypertension* 2011;57:132–140.
- 2. Hu D, Mohanta SK, Yin C, Peng L, Ma Z, Srikakulapu P, Grassia G, MacRitchie N, Dever G, Gordon P, Burton FL, Ialenti A, Sabir SR, McInnes IB, Brewer JM, Garside P, Weber C, Lehmann T, Teupser D, Habenicht L, Beer M, Grabner R, Maffia P, Weih F, Habenicht A. Artery tertiary lymphoid organs control aorta immunity and protect against atherosclerosis via vascular smooth muscle cell lymphotoxin β receptors. *Immunity* 2015;**42**:1100–1115.
- Welsh P, Grassia G, Botha S, Sattar N, Maffia P. Targeting inflammation to reduce cardiovascular disease risk: a realistic clinical prospect? Br J Pharmacol 2017;174:3898–3913.
- 4. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, Fonseca F, Nicolau J, Koenig W, Anker SD, Kastelein JJP, Cornel JH, Pais P, Pella D, Genest J, Cifkova R, Lorenzatti A, Forster T, Kobalava Z, Vida-Simiti L, Flather M, Shimokawa H, Ogawa H, Dellborg M, Rossi PRF, Troquay RPT, Libby P, Glynn RJ. Antiinflammatory therapy with canakinumab for atherosclerotic disease. N Engl J Med 2017;**377**:1119–1131.
- Prasad D, Rothlin CV, Burrola P, Burstyn-Cohen T, Lu Q, Garcia de Frutos P, Lemke G. TAM receptor function in the retinal pigment epithelium. *Mol Cell Neurosci* 2006;**33**:96–108.
- Nagata K, Ohashi K, Nakano T, Arita H, Zong C, Hanafusa H, Mizuno K. Identification of the product of growth arrest-specific gene 6 as a common ligand for Axl, Sky, and Mer receptor tyrosine kinases. J Biol Chem 1996;271:30022–30027.
- Stitt TN, Conn G, Goret M, Lai C, Bruno J, Radzlejewski C, Mattsson K, Fisher J, Gies DR, Jones PF, Masiakowski P, Ryan TE, Tobkes NJ, Chen DH, DiStefano PS, Long GL, Basilico C, Goldfarb MP, Lemke G, Glass DJ, Yancopoulos GD. The anticoagulation factor protein S and its relative, Gas6, are ligands for the Tyro 3/Axl family of receptor tyrosine kinases. *Cell* 1995;80:661–670.
- Lew ED, Oh J, Burrola PG, Lax I, Zagórska A, Través PG, Schlessinger J, Lemke G. Differential TAM receptor–ligand–phospholipid interactions delimit differential TAM bioactivities. *eLife* 2014;3:e03385.
- 9. Lemke G. Biology of the TAM receptors. Cold Spring Harb Perspect Biol 2013;5: a009076.
- Rothlin CV, Carrera-Silva EA, Bosurgi L, Ghosh S. TAM receptor signaling in immune homeostasis. Annu Rev Immunol 2015;33:355–391.
- Nagata S, Suzuki J, Segawa K, Fujii T. Exposure of phosphatidylserine on the cell surface. Cell Death Differ 2016;23:952–961.
- Sasaki T, Knyazev PG, Clout NJ, Cheburkin Y, Göhring W, Ullrich A, Timpl R, Hohenester E. Structural basis for Gas6-Axl signalling. *EMBO J* 2006;**25**:80–87.
- Lu Q, Gore M, Zhang Q, Camenisch T, Boast S, Casagranda F, Lai C, Skinner MK, Klein R, Matsushima GK, Earp HS, Goff SP, Lemke G. Tyro-3 family receptors are essential regulators of mammalian spermatogenesis. *Nature* 1999;**398**:723–728.
- Duncan JL, LaVail MM, Yasumura D, Matthes MT, Yang H, Trautmann N, Chappelow AV, Feng W, Earp HS, Matsushima GK, Vollrath D. An RCS-like retinal dystrophy phenotype in Mer knockout mice. *Invest Ophthalmol Vis Sci* 2003;**44**:826–838.
- Lu Q, Lemke G. Homeostatic regulation of the immune system by receptor tyrosine kinases of the Tyro 3 family. *Science* 2001;293:306–311.
- Rothlin CV, Ghosh S, Zuniga EI, Oldstone MBA, Lemke G. TAM receptors are pleiotropic inhibitors of the innate immune response. *Cell* 2007;**131**:1124–1136.
- Scott RS, McMahon EJ, Pop SM, Reap EA, Caricchio R, Cohen PL, Earp HS, Matsushima GK. Phagocytosis and clearance of apoptotic cells is mediated by MER. *Nature* 2001;411:207–211.
- Seitz HM, Camenisch TD, Lemke G, Earp HS, Matsushima GK. Macrophages and dendritic cells use different Axl/Mertk/Tyro3 receptors in clearance of apoptotic cells. J Immunol 2007;178:5635–5642.
- Elliott MR, Ravichandran KS. Clearance of apoptotic cells: implications in health and disease. J Cell Biol 2010;189:1059–1070.
- Muñoz LE, Lauber K, Schiller M, Manfredi AA, Herrmann M. The role of defective clearance of apoptotic cells in systemic autoimmunity. Nat Rev Rheumatol 2010;6:280.
- Henson PM. Cell removal: efferocytosis. Annu Rev Cell Dev Biol 2017;33:127–144.
 Lemke G, Burstyn-Cohen T. TAM receptors and the clearance of apoptotic cells. Ann N Y Acad Sci 2010;1209:23–29.
- Lemke G, Rothin CV. Immunobiology of the TAM receptors. Nat Rev Immunol 2008;8:327–336.

- Gonzalez L, Trigatti BL. Macrophage apoptosis and necrotic core development in atherosclerosis: a rapidly advancing field with clinical relevance to imaging and therapy. Can J Cardiol 2017;33:303–312.
- Linton MF, Babaev VR, Huang J, Linton EF, Tao H, Yancey PG. Macrophage apoptosis and efferocytosis in the pathogenesis of atherosclerosis. *Circ J* 2016;80: 2259–2268.
- Schrijvers DM, De Meyer GRY, Herman AG, Martinet W. Phagocytosis in atherosclerosis: molecular mechanisms and implications for plaque progression and stability. *Cardiovasc Res* 2007;**73**:470–480.
- Tabas I. Consequences and therapeutic implications of macrophage apoptosis in atherosclerosis: the importance of lesion stage and phagocytic efficiency. Arterioscler Thromb Vasc Biol 2005;25:2255–2264.
- Van Vré EA, Ait-Oufella H, Tedgui A, Mallat Z. Apoptotic cell death and efferocytosis in atherosclerosis. Arterioscler Thromb Vasc Biol 2012;32:887–893.
- Bentzon JF, Otsuka F, Virmani R, Falk E. Mechanisms of plaque formation and rupture. Circ Res 2014;114:1852–1866.
- Ait-Oufella H, Pouresmail V, Simon T, Blanc-Brude O, Kinugawa K, Merval R, Offenstadt G, Lesèche G, Cohen PL, Tedgui A, Mallat Z. Defective Mer receptor tyrosine kinase signaling in bone marrow cells promotes apoptotic cell accumulation and accelerates atherosclerosis. *Arterioscler Thromb Vasc Biol* 2008;28:1429–1431.
- Cai B, Thorp EB, Doran AC, Sansbury BE, Daemen MJAP, Dorweiler B, Spite M, Fredman G, Tabas I. MerTK receptor cleavage promotes plaque necrosis and defective resolution in atherosclerosis. J Clin Invest 2017;127:564–568.
- Thorp E, Cui D, Schrijvers DM, Kuriakose G, Tabas I. Mertk receptor mutation reduces efferocytosis efficiency and promotes apoptotic cell accumulation and plaque necrosis in atherosclerotic lesions of Apoe^{-/-} mice. Arterioscler Thromb Vasc Biol 2008;28:1421–1428.
- Han CZ, Juncadella JJ, Kinchen JM, Buckley MW, Klibanov AL, Dryden K, Onengut-Gumuscu S, Erdbrügger U, Turner SD, Shim YM, Tung KS, Ravichandran KS. Macrophages redirect phagocytosis by non-professional phagocytes and influence inflammation. *Nature* 2016;**539**:570–574.
- 34. Nakaya M, Watari K, Tajima M, Nakaya T, Matsuda S, Ohara H, Nishihara H, Yamaguchi H, Hashimoto A, Nishida M, Nagasaka A, Horii Y, Ono H, Iribe G, Inoue R, Tsuda M, Inoue K, Tanaka A, Kuroda M, Nagata S, Kurose H. Cardiac myofibroblast engulfment of dead cells facilitates recovery after myocardial infarction. J Clin Invest 2016;**127**:383–401.
- Batlle M, Recarte-Pelz P, Roig E, Castel MA, Cardona M, Farrero M, Ortiz JT, Campos B, Pulgarín MJ, Ramírez J, Pérez-Villa F, García de Frutos P. AXL receptor tyrosine kinase is increased in patients with heart failure. *Int J Cardiol* 2014;**173**: 402–409.
- Su AI, Wiltshire T, Batalov S, Lapp H, Ching KA, Block D, Zhang J, Soden R, Hayakawa M, Kreiman G, Cooke MP, Walker JR, Hogenesch JB. A gene atlas of the mouse and human protein-encoding transcriptomes. *Proc Natl Acad Sci USA* 2004; 101:6062–6067.
- Cai B, Thorp EB, Doran AC, Subramanian M, Sansbury BE, Lin C-S, Spite M, Fredman G, Tabas I. MerTK cleavage limits proresolving mediator biosynthesis and exacerbates tissue inflammation. *Proc Natl Acad Sci USA* 2016;**113**:6526–6531.
- Yoshimura A, Naka T, Kubo M. SOCS proteins, cytokine signalling and immune regulation. Nat Rev Immunol 2007;7:454.
- Carrera Silva EA, Chan PY, Joannas L, Errasti AE, Gagliani N, Bosurgi L, Jabbour M, Perry A, Smith-Chakmakova F, Mucida D, Cheroutre H, Burstyn-Cohen T, Leighton JA, Lemke G, Ghosh S, Rothlin CV. T cell-derived protein S engages TAM receptor signaling in dendritic cells to control the magnitude of the immune response. *Immunity* 2013;**39**:160–170.
- Caligiuri G, Paulsson G, Nicoletti A, Maseri A, Hansson GK. Evidence for antigendriven T-cell response in unstable angina. *Circulation* 2000;**102**:1114–1119.
- Koltsova EK, Garcia Z, Chodaczek G, Landau M, McArdle S, Scott SR, von Vietinghoff S, Galkina E, Miller YI, Acton ST, Ley K. Dynamic T cell–APC interactions sustain chronic inflammation in atherosclerosis. J Clin Invest 2012;122: 3114–3126.
- Sage AP, Murphy D, Maffia P, Masters LM, Sabir SR, Baker LL, Cambrook H, Finigan AJ, Ait-Oufella H, Grassia G, Harrison JE, Ludewig B, Reith W, Hansson GK, Reizis B, Hugues S, Mallat Z. MHC class II-restricted antigen presentation by plasmacytoid dendritic cells drives proatherogenic T cell immunity. *Circulation* 2014;**130**: 1363–1373.
- 43. Bauer T, Zagórska A, Jurkin J, Yasmin N, Köffel R, Richter S, Gesslbauer B, Lemke G, Strobl H. Identification of Axl as a downstream effector of TGF-β1 during Langerhans cell differentiation and epidermal homeostasis. J Exp Med 2012;209: 2033–2047.
- 44. Fujimori T, Grabiec AM, Kaur M, Bell TJ, Fujino N, Cook PC, Svedberg FR, MacDonald AS, Maciewicz RA, Singh D, Hussell T. The Axl receptor tyrosine kinase is a discriminator of macrophage function in the inflamed lung. *Mucosal Immunol* 2015;8:1021–1030.
- Zagórska A, Través PG, Lew ED, Dransfield I, Lemke G. Diversification of TAM receptor tyrosine kinase function. Nat Immunol 2014;15:920–928.
- Zizzo G, Cohen PL. IL-17 stimulates differentiation of human anti-inflammatory macrophages and phagocytosis of apoptotic neutrophils in response to IL-10 and glucocorticoids. *J Immunol* 2013;**190**:5237–5246.

- McColl A, Bournazos S, Franz S, Perretti M, Morgan BP, Haslett C, Dransfield I. Glucocorticoids induce protein S-dependent phagocytosis of apoptotic neutrophils by human macrophages. *J Immunol* 2009;**183**:2167–2175.
- 48. A-Gonzalez N, Bensinger SJ, Hong C, Beceiro S, Bradley MN, Zelcer N, Deniz J, Ramirez C, Díaz M, Gallardo G, Galarreta CR, de Salazar J, Lopez F, Edwards P, Parks J, Andujar M, Tontonoz P, Castrillo A. Apoptotic cells promote their own clearance and immune tolerance through activation of the nuclear receptor LXR. *Immunity* 2009;**31**:245–258.
- Tontonoz P, Mangelsdorf DJ. Liver X receptor signaling pathways in cardiovascular disease. Mol Endocrinol 2003;17:985–993.
- 50. Kurowska-Stolarska M, Alivernini S, Melchor EG, Elmesmari A, Tolusso B, Tange C, Petricca L, Gilchrist DS, Di Sante G, Keijzer C, Stewart L, Di Mario C, Morrison V, Brewer JM, Porter D, Milling S, Baxter RD, McCarey D, Gremese E, Lemke G, Ferraccioli G, McSharry C, McInnes IB. MicroRNA-34a dependent regulation of AXL controls the activation of dendritic cells in inflammatory arthritis. *Nat Commun* 2017;**8**:15877.
- 51. Kabir TD, Ganda C, Brown RM, Beveridge DJ, Richardson KL, Chaturvedi V, Candy P, Epis M, Wintle L, Kalinowski F, Kopp C, Stuart LM, Yeoh GC, George J, Leedman PJ. A microRNA-7/growth arrest specific 6/TYRO3 axis regulates the growth and invasiveness of sorafenib-resistant cells in human hepatocellular carcinoma. *Hepatology* 2018;**67**:216–231.
- 52. Orme JJ, Du Y, Vanarsa K, Mayeux J, Li L, Mutwally A, Arriens C, Min S, Hutcheson J, Davis LS, Chong BF, Satterthwaite AB, Wu T, Mohan C. Heightened cleavage of Axl receptor tyrosine kinase by ADAM metalloproteases may contribute to disease pathogenesis in SLE. *Clin Immunol* 2016;**169**:58–68.
- 53. Thorp E, Vaisar T, Subramanian M, Mautner L, Blobel C, Tabas I. Shedding of the Mer tyrosine kinase receptor is mediated by ADAM17 protein through a pathway involving reactive oxygen species, protein kinase Cδ, and p38 mitogen-activated protein kinase (MAPK). J Biol Chem 2011;286:33335–33344.
- 54. Triantafyllou E, Pop OT, Possamai LA, Wilhelm A, Liaskou E, Singanayagam A, Bernsmeier C, Khamri W, Petts G, Dargue R, Davies SP, Tickle J, Yuksel M, Patel VC, Abeles RD, Stamataki Z, Curbishley SM, Ma Y, Wilson ID, Coen M, Woollard KJ, Quaglia A, Wendon J, Thursz MR, Adams DH, Weston CJ, Antoniades CG. MerTK expressing hepatic macrophages promote the resolution of inflammation in acute liver failure. *Gut* 2018;**67**:333–347.
- 55. Garbin U, Baggio E, Stranieri C, Pasini A, Manfro S, Mozzini C, Vallerio P, Lipari G, Merigo F, Guidi G, Cominacini L, Fratta Pasini A. Expansion of necrotic core and shedding of Mertk receptor in human carotid plaques: a role for oxidized polyunsaturated fatty acids? *Cardiovasc Res* 2013;**97**:125–133.
- 56. DeBerge M, Yeap XY, Dehn S, Zhang S, Grigoryeva L, Misener S, Procissi D, Zhou X, Lee DC, Muller WA, Luo X, Rothlin C, Tabas I, Thorp EB. MerTK cleavage on resident cardiac macrophages compromises repair after myocardial ischemia reperfusion injurynovelty and significance. *Circ Res* 2017;**121**:930–940.
- Sather S, Kenyon KD, Lefkowitz JB, Liang X, Varnum BC, Henson PM, Graham DK. A soluble form of the Mer receptor tyrosine kinase inhibits macrophage clearance of apoptotic cells and platelet aggregation. *Blood* 2007;**109**:1026–1033.
- Nakano T, Higashino K, Kikuchi N, Kishino J, Nomura K, Fujita H, Ohara O, Arita H. Vascular smooth muscle cell-derived, Gla-containing growth-potentiating factor for Ca-mobilizing growth factors. J Biol Chem 1995;270:5702–5705.
- Konishi A, Aizawa T, Mohan A, Korshunov VA, Berk BC. Hydrogen peroxide activates the Gas6-Axl pathway in vascular smooth muscle cells. J Biol Chem 2004;279: 28766–28770.
- Cavet ME, Smolock EM, Ozturk OH, World C, Pang J, Konishi A, Berk BC. Gas6-Axl receptor signaling is regulated by glucose in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 2008;28:886–891.
- Goruppi S, Ruaro E, Varnum B, Schneider C. Requirement of phosphatidylinositol 3-kinase-dependent pathway and Src for Gas6-Axl mitogenic and survival activities in NIH 3T3 fibroblasts. *Mol Cell Biol* 1997;**17**:4442–4453.
- Cai B, Kasikara C, Doran AC, Ramakrishnan R, Birge RB, Tabas I. MerTK signaling in macrophages promotes the synthesis of inflammation resolution mediators by suppressing CaMKII activity. Sci Signal 2018;11:eaar3721.
- Gasic GP, Arenas CP, Gasic TB, Gasic GJ. Coagulation factors X, Xa, and protein S as potent mitogens of cultured aortic smooth muscle cells. *Proc Natl Acad Sci USA* 1992;89:2317–2320.
- Melaragno MG, Wuthrich DA, Poppa V, Gill D, Lindner V, Berk BC, Corson MA. Increased expression of Axl tyrosine kinase after vascular injury and regulation by G protein-coupled receptor agonists in rats. *Circ Res* 1998;83:697–704.
- Batchu SN, Xia J, Ko KA, Doyley MM, Abe J-I, Morrell CN, Korshunov VA. Axl modulates immune activation of smooth muscle cells in vein graft remodeling. *Am J Physiol Heart Circ Physiol* 2015;**309**:H1048–H1058.
- Korshunov VA, Mohan AM, Georger MA, Berk BC. Axl, a receptor tyrosine kinase, mediates flow-induced vascular remodeling. *Circ Res* 2006;**98**:1446–1452.
- Batchu SN, Hughson A, Gerloff J, Fowell DJ, Korshunov VA. Role of Axl in early kidney inflammation and progression of salt-dependent hypertension. *Hypertension* 2013;62:302–309.
- Korshunov VA, Daul M, Massett MP, Berk BC. Axl mediates vascular remodeling induced by deoxycorticosterone acetate-salt. *Hypertension* 2007;50:1057–1062.
- Fridell Y-W, Villa J, Attar EC, Liu ET. GAS6 induces Axl-mediated chemotaxis of vascular smooth muscle cells. J Biol Chem 1998;273:7123–7126.

- Melaragno MG, Cavet ME, Yan C, Tai L-K, Jin Z-G, Haendeler J, Berk BC. Gas6 inhibits apoptosis in vascular smooth muscle: role of Axl kinase and Akt. J Mol Cell Cardiol 2004;37:881–887.
- Nakano T, Kawamoto K, Higashino K, Arita H. Prevention of growth arrest-induced cell death of vascular smooth muscle cells by a product of growth arrest-specific gene, gas6. FEBS Lett 1996;387:78–80.
- Prescott MF, Webb RL, Reidy MA. Angiotensin-converting enzyme inhibitor versus angiotensin II, AT1 receptor antagonist. Effects on smooth muscle cell migration and proliferation after balloon catheter injury. Am J Pathol 1991;139:1291–1296.
- Berk BC. Angiotensin II signal transduction in vascular smooth muscle: pathways activated by specific tyrosine kinases. J Am Soc Nephrol 1999;10: S62–S68.
- Ishida M, Berk BC. Angiotensin II signal transduction in vascular smooth muscle cells: role of tyrosine kinases. *Keio J Med* 1997;46:61–68.
- Griendling KK, Ushio-Fukai M. Redox control of vascular smooth muscle proliferation. J Lab Clin Med 1998;132:9–15.
- Cavet ME, Smolock EM, Menon P, Konishi A, Korshunov VA, Berk BC. Gas6-Axl pathway. *Hypertension* 2010;56:105–111.
- Smolock EM, Korshunov VA. Pharmacological inhibition of Axl affects smooth muscle cell functions under oxidative stress. *Vascul Pharmacol* 2010;53:185–192.
- Goruppi S, Ruaro E, Schneider C. Gas6, the ligand of Axl tyrosine kinase receptor, has mitogenic and survival activities for serum starved NIH3T3 fibroblasts. *Oncogene* 1996;**12**:471–480.
- Bellosta P, Zhang Q, Goff SP, Basilico C. Signaling through the ARK tyrosine kinase receptor protects from apoptosis in the absence of growth stimulation. *Oncogene* 1997;**15**:2387–2397.
- D'Arcangelo D, Gaetano C, Capogrossi MC. Acidification prevents endothelial cell apoptosis by Axl activation. *Circ Res* 2002;91:e4–e12.
- D'Arcangelo D, Ambrosino V, Giannuzzo M, Gaetano C, Capogrossi MC. Axl receptor activation mediates laminar shear stress anti-apoptotic effects in human endothelial cells. *Cardiovasc Res* 2006;**71**:754–763.
- Hasanbasic I, Cuerquis J, Varnum B, Blostein MD. Intracellular signaling pathways involved in Gas6-Axl-mediated survival of endothelial cells. Am J Physiol Heart Circ Physiol 2004;287:H1207–H1213.
- Ruan G-X, Kazlauskas A. Axl is essential for VEGF-A-dependent activation of PI3K/ Akt. EMBO J 2012;31:1692–1703.
- Gerloff J, Korshunov VA. Immune modulation of vascular resident cells by Axl orchestrates carotid intima-media thickening. Am J Pathol 2012;180:2134–2143.
- Lee C-H, Shieh Y-S, Hsiao F-C, Kuo F-C, Lin C-Y, Hsieh C-H, Hung Y-J. High glucose induces human endothelial dysfunction through an Axl-dependent mechanism. *Cardiovasc Diabetol* 2014;**13**:53.
- Abedin M, Tintut Y, Demer LL. Vascular calcification: mechanisms and clinical ramifications. Arterioscler Thromb Vosc Biol 2004;24:1161–1170.
- Nakahara T, Dweck MR, Narula N, Pisapia D, Narula J, Strauss HW. Coronary artery calcification: from mechanism to molecular imaging. *JACC Cardiovasc Imaging* 2017;**10**:582–593.
- Shao J-S, Cai J, Towler DA. Molecular mechanisms of vascular calcification: lessons learned from the aorta. Arterioscler Thromb Vasc Biol 2006;26:1423–1430.
- Blaha MJ, Cainzos-Achirica M, Greenland P, McEvoy JW, Blankstein R, Budoff MJ, Dardari Z, Sibley CT, Burke GL, Kronmal RA, Szklo M, Blumenthal RS, Nasir K. Role of coronary artery calcium score of zero and other negative risk markers for cardiovascular disease: the Multi-Ethnic Study of Atherosclerosis (MESA). *Circulation* 2016;**133**:849–858.
- Collett G, Wood A, Alexander MY, Varnum BC, Boot-Handford RP, Ohanian V, Ohanian J, Fridell Y-W, Canfield AE. Receptor tyrosine kinase Axl modulates the osteogenic differentiation of pericytes. *Circ Res* 2003;**92**:1123–1129.
- Collett GDM, Sage AP, Kirton JP, Alexander MY, Gilmore AP, Canfield AE. Axl/ Phosphatidylinositol 3-kinase signaling inhibits mineral deposition by vascular smooth muscle cells. *Circ Res* 2007;**100**:502–509.
- Qiu C, Zheng H, Tao H, Yu W, Jiang X, Li A, Jin H, Lv A, Li H. Vitamin K2 inhibits rat vascular smooth muscle cell calcification by restoring the Gas6/Axl/Akt antiapoptotic pathway. *Mol Cell Biochem* 2017;433:149–159.
- Badi I, Mancinelli L, Polizzotto A, Ferri D, Zeni F, Burba I, Milano G, Brambilla F, Saccu C, Bianchi ME, Pompilio G, Capogrossi MC, Raucci A. miR-34a promotes vascular smooth muscle cell calcification by downregulating SIRT1 (Sirtuin 1) and Axl (AXL Receptor Tyrosine Kinase). Arterioscler Thromb Vasc Biol 2018;38:2079–2090.
- Son B-K, Kozaki K, lijima K, Eto M, Kojima T, Ota H, Senda Y, Maemura K, Nakano T, Akishita M, Ouchi Y. Statins protect human aortic smooth muscle cells from inorganic phosphate-induced calcification by restoring Gas6-Axl survival pathway. *Circ* Res 2006;**98**:1024–1031.
- Son B-K, Kozaki K, lijima K, Eto M, Nakano T, Akishita M, Ouchi Y. Gas6/AxI-PI3K/ Akt pathway plays a central role in the effect of statins on inorganic phosphateinduced calcification of vascular smooth muscle cells. *Eur J Pharmacol* 2007;556:1–8.
- 96. Caldentey G, García De Frutos P, Cristóbal H, Garabito M, Berruezo A, Bosch X, San Antonio R, Flores-Umanzor E, Perea RJ, De Caralt TM, Rodríguez J, Ortiz-Pérez JT. Serum levels of Growth Arrest-Specific 6 protein and soluble AXL in patients with ST-segment elevation myocardial infarction. Eur Heart J Acute Cardiovasc Care 2017; doi:10.1177/2048872617740833.
- Zhao YF, Xu DC, Zhu GF, Zhu MY, Tang K, Li WM, Xu YW. Growth arrestspecific 6 exacerbates pressure overload-induced cardiac hypertrophy. *Hypertension* 2016;67:118–129.

- Goruppi S, Ruaro E, Varnum B, Schneider C. Gas6-mediated survival in NIH3T3 cells activates stress signalling cascade and is independent of Ras. Oncogene 1999;18: 4224–4236.
- Subramanian M, Proto JD, Matsushima GK, Tabas I. Deficiency of AXL in bone marrow-derived cells does not affect advanced atherosclerotic lesion progression. *Sci Rep* 2016;6:39111.
- 100. Hurtado B, Muñoz X, Recarte-Pelz P, Garcia N, Luque A, Krupinski J, Sala N, Frutos P. D. Expression of the vitamin K-dependent proteins GAS6 and protein S and the TAM receptor tyrosine kinases in human atherosclerotic carotid plaques. *Thromb Haemost* 2011;**105**:873–882.
- Lee C-H, Shieh Y-S, Tsai C-S, Hung Y-J, Tsai Y-T, Lin C-Y. Expression of growth arrest-specific protein 6 and Axl molecules in the left internal mammary artery of patients undergoing coronary artery bypass grafting. J Clin Pathol 2014;67:506–511.
- Liu Y-W, Yang Q-F, Zuo P-Y, Xiao C-L, Chen X-L, Liu C-Y. Elevated serum levels of soluble axl in acute coronary syndrome. Am J Med Sci 2015;349:124–129.
- Clauser S, Meilhac O, Bièche I, Raynal P, Bruneval P, Michel J-B, Borgel D. Increased secretion of Gas6 by smooth muscle cells in human atherosclerotic carotid plaques. *Thromb Haemost* 2012;**107**:140–149.
- Tjwa M, Moons L, Lutgens E. Pleiotropic role of growth arrest-specific gene 6 in atherosclerosis. *Curr Opin Lipidol* 2009;20:386–392.
- Lutgens E, Tjwa M, Frutos PG, de Wijnands E, Beckers L, Dahlbäck B, Daemen M, Carmeliet P, Moons L. Genetic loss of Gas6 induces plaque stability in experimental atherosclerosis. J Pathol 2008;216:55–63.
- 106. Sunbul M, Cagman Z, Gerin F, Ozgen Z, Durmus E, Seckin D, Ahmad S, Uras F, Agirbasli M. Growth arrest-specific 6 and cardiometabolic risk factors in patients with psoriasis. *Cardiovasc Ther* 2015;**33**:56–61.
- Batchu N, Hughson A, Wadosky KM, Morrell CN, Fowell DJ, Korshunov VA. The role of Axl in T lymphocyte survival in salt-dependent hypertension. Arterioscler Thromb Vasc Biol 2016;36:1638–1646.
- 108. Paolino M, Choidas A, Wallner S, Pranjic B, Uribesalgo I, Loeser S, Jamieson AM, Langdon WY, Ikeda F, Fededa JP, Cronin SJ, Nitsch R, Schultz-Fademrecht C, Eickhoff J, Menninger S, Unger A, Torka R, Gruber T, Hinterleitner R, Baier G, Wolf D, Ullrich A, Klebl BM, Penninger JM. The E3 ligase Cbl-b and TAM receptors regulate cancer metastasis via natural killer cells. *Nature* 2014;**507**:508–512.
- 109. Park I-K, Giovenzana C, Hughes TL, Yu J, Trotta R, Caligiuri MA. The AxI/Gas6 pathway is required for optimal cytokine signaling during human natural killer cell development. Blood 2009;113:2470–2477.
- Kim EM, Lee EH, Lee HY, Choi HR, Ji KY, Kim SM, Kim KD, Kang HS. Axl signaling induces development of natural killer cells *in vitro* and *in vivo*. *Protoplasma* 2017;254: 1091–1101.
- 111. Backteman K, Ernerudh J, Jonasson L. Natural killer (NK) cell deficit in coronary artery disease: no aberrations in phenotype but sustained reduction of NK cells is associated with low-grade inflammation. *Clin Exp Immunol* 2014;**175**:104–112.
- Ong S, Rose NR, Čiháková D. Natural killer cells in inflammatory heart disease. *Clin Immunol* 2017;**175**:26–33.
- Selathurai A, Deswaerte V, Kanellakis P, Tipping P, Toh B-H, Bobik A, Kyaw T. Natural killer (NK) cells augment atherosclerosis by cytotoxic-dependent mechanisms. *Cardiovasc Res* 2014;**102**:128–137.
- 114. Doran AC, Ozcan L, Cai B, Zheng Z, Fredman G, Rymond CC, Dorweiler B, Sluimer JC, Hsieh J, Kuriakose G, Tall AR, Tabas I. CAMKIIγ suppresses an efferocytosis pathway in macrophages and promotes atherosclerotic plaque necrosis. J Clin Invest 2017;**127**:4075–4089.
- Zizzo G, Hilliard BA, Monestier M, Cohen PL. Efficient clearance of early apoptotic cells by human macrophages requires M2c polarization and MerTK induction. *J Immunol* 2012;**189**:3508–3520.
- 116. Fourgeaud L, Través PG, Tufail Y, Leal-Bailey H, Lew ED, Burrola PG, Callaway P, Zagórska A, Rothlin CV, Nimmerjahn A, Lemke G. TAM receptors regulate multiple features of microglial physiology. *Nature* 2016;**532**:240–244.
- 117. Miner JJ, Daniels BP, Shrestha B, Proenca-Modena JL, Lew ED, Lazear HM, Gorman MJ, Lemke G, Klein RS, Diamond MS. The TAM receptor Mertk protects against neuroinvasive viral infection by maintaining blood-brain barrier integrity. *Nat Med* 2015;**21**:1464–1472.
- Chistiakov DA, Orekhov AN, Bobryshev YV. Endothelial barrier and its abnormalities in cardiovascular disease. Front Physiol 2015;6:365.
- 119. Bujak M, Kweon HJ, Chatila K, Li N, Taffet G, Frangogiannis NG. Aging-related defects are associated with adverse cardiac remodeling in a mouse model of reperfused myocardial infarction. J Am Coll Cardiol 2008;51:1384–1392.
- 120. Chen W, Frangogiannis NG. The role of inflammatory and fibrogenic pathways in heart failure associated with aging. *Heart Fail Rev* 2010;**15**:415–422.
- 121. Thorp EB. Contrasting inflammation resolution during atherosclerosis and post myocardial infarction at the level of monocyte/macrophage phagocytic clearance. *Front Immunol* 2012;**3**:39.
- Frangogiannis NG. Regulation of the inflammatory response in cardiac repair. *Circ* Res 2012;110:159–173.
- 123. Howangyin K-Y, Zlatanova I, Pinto C, Ngkelo A, Cochain C, Rouanet M, Vilar J, Lemitre M, Stockmann C, Fleischmann BK, Mallat Z, Silvestre J-S. Myeloid-epithelialreproductive receptor tyrosine kinase and milk fat globule epidermal growth factor 8 coordinately improve remodeling after myocardial infarction via local delivery of vascular endothelial growth factor. *Circulation* 2016;**133**:826–839.

- 124. Wan E, Yeap XY, Dehn S, Terry R, Novak M, Zhang S, Iwata S, Han X, Homma S, Drosatos K, Lomasney J, Engman DM, Miller SD, Vaughan DE, Morrow JP, Kishore R, Thorp EB. Enhanced efferocytosis of apoptotic cardiomyocytes through myeloid-epithelial-reproductive tyrosine kinase links acute inflammation resolution to cardiac repair after infarction. *Circ Res* 2013;**113**:1004–1012.
- 125. Zhang S, Yeap X-Y, Grigoryeva L, Dehn S, DeBerge M, Tye M, Rostlund E, Schrijvers D, Zhang ZJ, Sumagin R, Tourtellotte WG, Lee D, Lomasney J, Morrow J, Thorp EB. Cardiomyocytes induce macrophage receptor shedding to suppress phagocytosis. J Mol Cell Cardiol 2015;87:171–179.
- 126. Scarabelli T, Stephanou A, Rayment N, Pasini E, Comini L, Curello S, Ferrari R, Knight R, Latchman D. Apoptosis of endothelial cells precedes myocyte cell apoptosis in ischemia/reperfusion injury. *Circulation* 2001;**104**:253–256.
- Yellon DM, Hausenloy DJ. Myocardial reperfusion injury. N Engl J Med 2007;357: 1121–1135.
- 128. Zhang S, Yeap X-Y, DeBerge M, Naresh NK, Wang K, Jiang Z, Wilcox JE, White SM, Morrow JP, Burridge PW, Procissi D, Scott EA, Frazier W, Thorp EB. Acute CD47 blockade during ischemic myocardial reperfusion enhances phagocytosisassociated cardiac repair. JACC Basic Transl Sci 2017;2:386–397.
- Castrillo A, Tontonoz P. Nuclear receptors in macrophage biology: at the crossroads of lipid metabolism and inflammation. Annu Rev Cell Dev Biol 2004;20: 455–480.
- 130. Li Y, Gerbod-Giannone M-C, Seitz H, Cui D, Thorp E, Tall AR, Matsushima GK, Tabas I. Cholesterol-induced apoptotic macrophages elicit an inflammatory response in phagocytes, which is partially attenuated by the Mer receptor. *J Biol Chem* 2006;**281**:6707–6717.
- 131. Liao D, Wang X, Li M, Lin PH, Yao Q, Chen C. Human protein S inhibits the uptake of AcLDL and expression of SR-A through Mer receptor tyrosine kinase in human macrophages. *Blood* 2009;**113**:165–174.
- Hurtado B, Abasolo N, Muñoz X, García N, Benavente Y, Rubio F, García de Frutos P, Krupinski J, Sala N. Association study between polymorphims in GAS6-TAM genes and carotid atherosclerosis. *Thromb Haemost* 2010;**104**:592–598.
- Dai W, Pan H, Hassanain H, Gupta SL, Murphy MJ. Molecular cloning of a novel receptor tyrosine kinase, TIF, highly expressed in human ovary and testis. *Oncogene* 1994;9:975–979.
- 134. Mark MR, Scadden DT, Wang Z, Gu Q, Goddard A, Godowski PJ. rse, a novel receptor-type tyrosine kinase with homology to AxI/Ufo, is expressed at high levels in the brain. J Biol Chem 1994;269:10720–10728.
- 135. Ken-Dror G, Cooper JA, Humphries SE, Drenos F, Ireland HA. Free protein S level as a risk factor for coronary heart disease and stroke in a prospective cohort study of healthy United Kingdom men. Am J Epidemiol 2011;**174**:958–968.
- 136. Chan PY, Carrera Silva EA, De Kouchkovsky D, Joannas LD, Hao L, Hu D, Huntsman S, Eng C, Licona-Limón P, Weinstein JS, Herbert DR, Craft JE, Flavell RA, Repetto S, Correale J, Burchard EG, Torgerson DG, Ghosh S, Rothlin CV. The TAM family receptor tyrosine kinase TYRO3 is a negative regulator of type 2 immunity. *Science* 2016;**352**:99–103.
- Zhao W, Lei T, Li H, Sun D, Mo X, Wang Z, Zhang K, Ou H. Macrophage-specific overexpression of interleukin-5 attenuates atherosclerosis in LDL receptordeficient mice. *Gene Ther* 2015;22:645–652.
- Zhao XN, Li YN, Wang YT. Interleukin-4 regulates macrophage polarization via the MAPK signaling pathway to protect against atherosclerosis. *Genet Mol Res* 2016;**15**. doi:10.4238/gmr.15017348.
- 139. Zhang W, Tang T, Nie D, Wen S, Jia C, Zhu Z, Xia N, Nie S, Zhou S, Jiao J, Dong W, Lv B, Xu T, Sun B, Lu Y, Li Y, Cheng L, Liao Y, Cheng X. IL-9 aggravates the development of atherosclerosis in ApoE^{-/-} mice. *Cardiovasc Res* 2015;**106**: 453–464.
- 140. Cieslik KA, Taffet GE, Carlson S, Hermosillo J, Trial J, Entman ML. Immune-inflammatory dysregulation modulates the incidence of progressive fibrosis and diastolic stiffness in the aging heart. J Mol Cell Cardiol 2011;50:248–256.
- 141. Kaviratne M, Hesse M, Leusink M, Cheever AW, Davies SJ, McKerrow JH, Wakefield LM, Letterio JJ, Wynn TA. IL-13 activates a mechanism of tissue fibrosis that is completely TGF- β independent. *J Immunol* 2004;**173**:4020–4029.
- 142. Trial J, Cieslik KA, Haudek SB, Duerrschmid C, Entman ML. Th1/M1 conversion to Th2/M2 responses in models of inflammation lacking cell death stimulates maturation of monocyte precursors to fibroblasts. *Front Immunol* 2013;4:287.
- 143. Proto JD, Doran AC, Gusarova G, Yurdagul A, Sozen E, Subramanian M, Islam MN, Rymond CC, Du J, Hook J, Kuriakose G, Bhattacharya J, Tabas I. Regulatory T cells promote macrophage efferocytosis during inflammation resolution. *Immunity* 2018; 49:666–677.e6.
- Cummings CT, DeRyckere D, Earp HS, Graham DK. Molecular pathways: MERTK signaling in cancer. *Clin Cancer Res* 2013;19:5275–5280.
- Vouri M, Hafizi S. TAM receptor tyrosine kinases in cancer drug resistance. Cancer Res 2017;77:2775–2778.
- 146. Petta S, Valenti L, Marra F, Grimaudo S, Tripodo C, Bugianesi E, Cammà C, Cappon A, Di Marco V, Di Maira G, Dongiovanni P, Rametta R, Gulino A, Mozzi E, Orlando E, Maggioni M, Pipitone RM, Fargion S, Craxì A. MERTK rs4374383 polymorphism affects the severity of fibrosis in non-alcoholic fatty liver disease. J Hepatol 2016;64: 682–690.
- 147. Law LA, Graham DK, Di Paola J, Branchford BR. GAS6/TAM pathway signaling in hemostasis and thrombosis. *Front Med (Lausanne)* 2018;**5**:137.