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Exogenous and endogenous factors in seasonality of reproduction in buffalo: A review

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A R T I C L E I N F O

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ABSTRACT

Seasonal breeding in buffalo is influenced by exogenous (photoperiod, climate, nutrition, management) and endogenous (hormones, genotype) factors, Buffalo are negatively photoperiodic and show a natural increase in fertility during decreasing day length. The hormone melatonin is produced by the pineal gland and has a fundamental role in photoperiodic time measurement within the brain. This drives annual cycles of gonadotropin secretion and gonadal function in buffaloes. Some melatonin is released into the systemic circulation and, together with peripherally produced melatonin, acts at somatic tissues. In the ovaries and testes of buffalo, melatonin acts as an antioxidant and scavenges oxygen free radicals to reduce both oxidative stress and apoptosis. This has beneficial effects on gametogenesis and steroidogenesis. Female buffalo treated with melatonin show an improved response to estrus synchronization protocols in out-of-season breeding. Melatonin acts through melatonin receptors MT₁ and MT₂ and the gene for MT₁ (MTNR1A) is polymorphic in buffaloes. Single nucleotide polymorphisms (SNPs) in gene MTNR1A have been associated with fertility in female buffalo. The knowledge and tools are available to lift the reproductive performance of buffalo. This is highly important as the global demand for nutritious buffalo food products has undergone a sharp rise, and continues to grow. Buffalo can make an important contribution to affordable, nutritious animal protein. This will help address global nutritional security.

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1. Introduction

Buffalo have been an important farm animal for more than 5000 years and their significance in global food systems continues to grow [1-6]. Today, buffalo-derived products are found at the very expensive and less expensive extremes of food markets. Buffalo contribute premium, niche food such as mozzarella cheese [7-9] and they are also a source of affordable, highly nutritious meat protein [10,11]. The latter is particularly important in nutritional security in Asia and The Middle East [10-14]. The growth in demand for affordable buffalo 'beef' allows smallholder and landless

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https://doi.org/10.1016/j.theriogenology.2020.01.044 0093-691X/© 2020 Elsevier Inc. All rights reserved. farmers to participate in global food markets. This creates the opportunity for poverty alleviation and improvement in rural livelihoods in developing countries, and is highly relevant to the 2017 Sustainable Development Goals [15]. In a global food system, vulnerable farmers need to be protected by ethical supply-chains and governance structures.

Growth in buffalo production is constrained by their reproductive biology. Buffalo are generally considered to be late sexually maturing [16–19] and to have an extended postpartum anestrus [16,20,21] compared with cattle. Selection over 30 years in the Italian Mediterranean buffalo reduced age at first calving from 42 to 36 months and, with management, age at first calving was further reduced to 28 months [22]. Buffalo also have a longer gestation period (typically 300–320 days) compared with beef cattle (*Bos taurus* 279–283 day; *Bos indicus*, 290–293 days) [23]. The







reproductive characteristics of female buffalo make it very difficult for buffalo to produce an offspring yearly. Added to these features is the fact buffalo are negatively photoperiodic seasonal breeders and show natural annual fluctuations in fertility [24–31]. In addition, the vast majority of buffalo are in tropical and subtropical areas where high temperatures and humidity in summer, together with relatively poor nutrition, cause seasonal anestrus [32–35]. Environmental factors can, however, be relative as Italian Mediterranean buffalo are bred out-of-season during summer-spring when the temperature-humidity index is at its highest [29,36-38]. Notwithstanding the reproductive challenges in buffalo, the recent sharp rise in demand for affordable and nutritious buffalo food products has made it necessary to develop new strategies that optimize the reproductive performance of buffalo. This needs to occur in both endowed and non-endowed production systems. Genetic selection for seasonality of breeding and fertility is one strategy that is mentioned below.

This review seeks to bring together information on exogenous and endogenous factors that contribute to seasonality of reproduction in buffalo. Exogenous factors include photoperiod, nutrition, temperature, humidity and management. Endogenous factors include IGF1, insulin, prolactin, thyroid hormones, melatonin, progesterone, and genotype. Melatonin is given particular attention as recent studies have demonstrated a direct beneficial effect on male and female gametes in buffalo, and treatment with melatonin has shown promise in improving fertility in anestrous buffalo.

2. Exogenous factors in buffalo reproduction

Exogenous factors that influence reproductive function in buffalo have been extensively reviewed [5,6,24,25] and this section provides only a general overview. Photoperiod is arguably the main exogenous regulator of reproduction in buffalo. Buffalo are negatively photoperiodic and show a natural increase in fertility in response to decreasing day length [24,26–29]. This is particularly evident at higher latitudes [27] but also occurs in the sub-tropics [39,40]. Buffalo show a decline in fertility in response to increasing day length [26]. The decline in fertility induced by long day length can be partly overcome with assisted reproductive technology (ART), including treatments for ovulation induction coupled with AI [31,39-41]. The potential for melatonin to enhance the response to ovulation induction protocols in anestrous buffalo is discussed below. Assisted reproductive technology is necessary to maintain continuity of reproduction in female buffalo because the 300-320 day gestation, and postpartum anestrous period, causes a decoupling of reproductive function from annual cycles of decreasing day length. There are also market drivers such as the yearly cycle in peak demand for mozzarella cheese which requires Italian Mediterranean buffalo to be bred outside the stimulatory window of decreasing day length [36].

In the tropics and sub-tropics, high temperatures and humidity in summer combine to induce anestrus in buffalo [5,25,42,43]. Management strategies to mitigate these stressors, such as active cooling, are necessary to maintain a degree of productivity. Also during summer, both the quantity and quality of feed are typically reduced, which adds a further nutritional stressor to reproductive function [5,25]. However, as noted above, market demand for mozzarella cheese requires breeding during summer-spring in intensively managed Italian Mediterranean buffalo.

Other than photoperiod, external factors that have a negative impact on reproduction must be managed in order to realize the opportunity presented by the global demand for buffalo food products. Approaches include strategic nutritional supplementation, the use of ART, and the implementation of management strategies (e.g. cooling, loafing space) that optimize the wellbeing of buffalo across endowed and non-endowed production systems.

3. Endogenous factors in buffalo reproduction

3.1. Prolactin, TRH, insulin, IGF1, P₄

Endogenous factors that regulate reproduction in buffalo are generally influenced by exogenous stimuli. An example is prolactin which is elevated during long day length [44]. Female buffalo that enter the condition of 'hyperprolactinemia' in summer due to day length, and heat stress, have reduced gonadotropin secretion and become anestrus [45,46]. A cause-and-effect of elevated prolactin under these conditions was suggested by the improved response to estrus synchronization in buffalo treated with norprolac, which lowered prolactin [45]. Notwithstanding the latter study, it was suggested that hypothyroidism, with elevated thyrotropin releasing hormone (TRH), may combine with elevated prolactin to suppress reproduction in female buffalo [20,47]. The interrelationship of endocrine disruption (prolactin, TRH, thyroid hormones) with stress and reproduction requires further study in seasonal buffalo. Factors associated with metabolism, and which impact gonadal function, such as insulin and IGF1, are typically influenced by nutrition [43,48,49]. IGF1 is particularly important in ovarian folliculogenesis and, in one study, Italian Mediterranean buffalo heifers maintained on a constant nutritional regimen had greater follicular IGF1 during short days compared to long days [50]. This finding suggested that photoperiod can directly affect follicular IGF1 in buffalo, although the mechanism is unknown.

The influence of day length on development of the corpus luteum and secretion of P₄, essential for the maintenance of a pregnancy, is well established in buffalo [27–29,31]. During long days, the corpus luteum is less vascularized and secretes less P₄ [29]. This is associated with greater embryonic mortality in buffalo during long days [27]. These studies were undertaken in Italian Mediterranean buffalo under a relatively constant nutritional regimen and further illustrated the important role of day length in seasonal breeding in buffalo. Nili-Ravi buffalo in Pakistan bred at spontaneous [51] or synchronized [52] estrus had a lower pregnancy rate [51,52] and greater pregnancy failure [52] during summer compared with winter.

3.2. Melatonin

Melatonin, produced by the pineal gland in the brain, has an important role centrally in photoperiodic time measurement which drives seasonal breeding in ruminants, including buffalo [53–56]. Seasonal changes in the circadian pattern of brain melatonin impact on the function of GnRH neurons in the hypothalamus that, in turn, influence pituitary gonadotropin secretion and reproductive function in ruminants [57,58].

Some brain melatonin is released into the general circulation and acts peripherally [59,60]. Melatonin is also produced by somatic tissues [54,55,61] and melatonin receptors (MT₁, MT₂) are expressed peripherally [62–66]. It is now recognized that locallyproduced melatonin can have autocrine and paracrine actions at somatic tissues including the ovaries and testes [67–73]. An important property of melatonin, and its metabolites, is a powerful antioxidant action [74–79]. Indeed, melatonin is a more powerful scavenger of oxygen and nitrogen free radicals than vitamin E [76]. The antioxidant property of melatonin protects cells from free radical damage and apoptotic mechanisms. The beneficial effects of melatonin on sperm, oocytes and embryos are conferred, at least in part, by its antioxidant action [80] (Fig. 1). Acyclic buffalo had greater amounts of follicular reactive oxygen species [81] and treatment with melatonin reduced oxidative stress in summer



Fig. 1. Actions of melatonin at the brain, ovaries and testes. Within the brain melatonin is involved in photoperiodic time measurement which drives seasonal changes in the activity of GnRH neurons and gonadotrophin secretion. Some melatonin is released into the systemic circulation and, together with locally produced melatonin, influences the function of follicles and oocytes, Sertoli cells, Leydig cells, and sperm. Melatonin is a strong antioxidant and scavenges reactive free radicals to reduce oxidative stress and apoptosis. Melatonin acts through melatonin receptors MT₁ and MT₂ which have been demonstrated on follicles, Sertoli cells, Leydig cells and sperm.

anestrous buffalo [82].

Melatonin is present in seminal plasma and the inclusion of melatonin had a protective effect during cryopreservation of buffalo semen [83,84]. Melatonin also facilitated *in vitro* sperm capacitation in buffalo [85]. Murrah buffalo bulls implanted with melatonin showed an improvement in semen quality during the nonbreeding season [86]. Melatonin treatment also improved semen quality in seasonal Mithun bulls [87]. It can be concluded from these findings that peripherally-derived and testicular melatonin has a role in sperm survival and function in buffalo and other ruminants. The European bison, a seasonal breeder, shows annual changes in testicular melatonin and melatonin receptors MT₁ and MT₂ [88]. Melatonin receptors MT₁ and MT₂ are expressed in sperm of seasonal and non-seasonal breeders [89].

In sheep, melatonin and its receptors MT_1 and MT_2 are expressed by cumulus-oocyte complexes [90] and the corpus

luteum [91]. Receptor MT₁ is also expressed by luteal cells in mares [92]. In cattle, the addition of melatonin increased the expression of LH receptors in cultured granulosa cells [93], increased steroidogenic enzymes and P₄ production by cultured thecal cells [94], and decreased apoptosis in cultured granulosa cells [93]. Melatonin also increased steroidogenesis and P₄ production by luteal cells in mares [92] and sows [95]. In buffalo, melatonin increased steroidogenesis and reduced apoptosis in cultured granulosa cells [96]. The supplementation of maturation media with melatonin improved maturation and in vitro fertilization of buffalo oocytes [97,98]. Melatonin also improved oocyte maturation and IVF outcome in pigs [99] and sheep [100–102]. Improved oocyte maturation in pigs was associated with lesser reactive oxygen species [99]. In contrast, the presence of melatonin did not improve in vitro oocyte maturation or embryo development in cattle [103,104]. Returning to the antioxidant properties of melatonin, follicular fluid of acyclic buffalo had greater amounts of oxygen free radicals and lesser antioxidant capacity compared with follicular fluid of cyclic buffalo [81]. It can be hypothesized that an additional beneficial effect of melatonin is to neutralize free radicals in follicular fluid (Fig. 1). Cryopreservation was associated with greater amounts of reactive oxygen species in Bos indicus embryos compared with Bos taurus embryos [105], suggesting that the inclusion of melatonin could potentially be beneficial in embryo technologies in Bos indicus cattle.

Melatonin is secreted during darkness in 24 h light-dark cycles [106,107]. This means that the duration of melatonin secretion is greater in short days than in long days [108]. An increased period of melatonin secretion in short days has a positive effect on reproductive function in short day breeders (e.g. buffalo, deer, sheep). A decreased period of melatonin secretion in long days has a positive effect on reproductive function in long day breeders (e.g. hamster, horse). The positive relationship between increased duration of exposure to melatonin and stimulation of reproductive function in buffalo [109–111] led to a series of studies that examined the ability of melatonin treatment to overcome seasonal anestrus. A typical design of these studies involved the administration of melatonin (18 mg/50 kg LW) around 40–45 days before the implementation of either a P₄-based [112-114] or GnRH-based [115] estrus synchronization protocol. The general finding from these studies was that prior exposure to melatonin is associated with increased functionality of the corpus luteum and greater capacity to maintain a pregnancy in summer anestrous buffalo heifers [112,113,115] and lactating buffalo cows [114]. The majority of these studies were undertaken in Indian Murrah buffalo in a tropical environment and further studies should be carried out in different environments, including higher latitudes. Seasonally anestrous Anglo-Nubian does, pre-treated with 18 mg melatonin for 42 days before a P₄based estrus synchronization protocol, had a greater conception rate and greater fecundity compared with does that did not receive melatonin [116]. A similar improvement in fertility was observed in seasonally anestrous New Zealand Romney composite ewes pretreated with melatonin for 35 days before a P₄-based estrus synchronization protocol [117]. The broad conclusion from these studies in short day breeders is that melatonin has a beneficial effect on fertility in seasonally anestrous females subjected to an estrus synchronization protocol.

The gene for melatonin receptor MT_1 (*MTRN1A*) is polymorphic and has been associated with productive and reproductive traits in buffalo [118,119]. Single nucleotide polymorphisms (SNPs) in gene *MTRN1A* were associated with conception rates in Indian Murrah buffalo heifers treated with melatonin during summer [120]. The gene *MTRN1A* also has a polymorphic site in Italian Mediterranean buffalo [121–123]. The latter was associated with differences in seasonal patterns of mating [121] and the capacity for out-ofseason breeding in females [122]. The polymorphism was not associated with puberty in Italian Mediterranean buffalo heifers [123]. A polymorphism in exon II of gene MTRN1A was associated with seasonality of breeding in Indian Murrah buffalo cows [124]. However, a polymorphism in exon 1 of gene MTRN1A was not associated with reproductive traits in Brazilian Amazon buffalo cows [125,126]. Polymorphism in gene MTRN1A was also reported for Indian Chokla sheep [127], Italian Sarda sheep [128], French Mérino d'Arles sheep [129], Greek local breed sheep [130], and Dorset crossbred sheep [131]. In most sheep breeds there was an association between MTRN1A polymorphism and out-of-season breeding. The studies on polymorphism in gene MTRN1A present the interesting possibility of selecting female buffalo that can be mated at different times of the year in order to meet production and market requirements. Recent genome wide association studies [132–134] and RNA-seg analyses [134] have uncovered polymorphism in other genes associated with reproductive function in female buffalo.

Two features of melatonin in buffalo are worth noting. First, the rise in melatonin during darkness was less pronounced in Italian Mediterranean buffalo heifers compared with older cows [135]. It was suggested that this might explain, at least in part, why seasonal effects on reproduction are less apparent in heifers [22,24,135]. Second, multiparous Italian Mediterranean buffalo that showed clear seasonal changes in reproductive function had a greater increase in melatonin during darkness compared to buffalo that showed less pronounced seasonal changes in reproduction [136]. The difference in nighttime melatonin between seasonal and less seasonal buffalo was observed throughout the year [136]. Buffalo with relatively high nighttime melatonin and seasonal breeding retained these features when relocated to a buffalo farm where local animals had relatively low nighttime melatonin and less pronounced seasonal breeding [137]. It would appear from these studies that both the magnitude (above a threshold) and duration of melatonin secretion during darkness are involved in photoperiodic time measurement which cues reproduction to day length in buffalo. The magnitude of the rise in melatonin in darkness most likely has a genetic component and could potentially be used to select for seasonality, similar to polymorphism in gene MTRN1A which codes for melatonin receptor MT₁, as discussed above.

4. Conclusions

The action of melatonin in the brain to regulate photoperiodic time measurement and seasonal breeding is well established in ruminants. More recent evidence indicates that melatonin also acts peripherally at somatic tissues. A direct action of melatonin on ovarian function is strongly supported by the expression of its receptors in follicles and corpus luteum. The benefits conferred by melatonin in out-of-season mating, and the response to estrus synchronization and AI in summer, could result from a combination of (i) systemic and/or local antioxidant effects, (ii) stimulation of steroidogenesis, and/or (iii) suppression of apoptotic mechanisms. The potential to use polymorphism in the melatonin receptor gene MTRN1A to select for female buffalo that show optimal reproduction at different times of the year presents an interesting possibility. Efforts to improve fertility in buffalo are a worthy pursuit given the sharp rise in demand for affordable, nutritious buffalo food products. Those to benefit must include smallholder and landless farmers who are increasingly participating in the global food system.

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References

- Borghese A, Mazzi M. Buffalo population and strategies in the world. In: Borghese A, editor. Buffalo production and research. FAO Reu Technical Series; 2005. p. 1–39. vol. 67.
- [2] Moioli B, Borghese A. Buffalo breeds and management systems. In: Borghese A, editor. Buffalo production and research. vol. 67. FAO Reu Technical Series; 2005. p. 51–76.
- [3] Nanda AS, Nakao T. Role of buffalo in the socioeconomic development of rural Asia: current status and future prospectus. Anim Sci J 2003;74:443–55. https://doi.org/10.1046/j.1344-3941.2003.00138.x.
- [4] Nanda AS, Brar PS, Prabhakar S. Enhancing reproductive performance in dairy buffalo: major constraints and achievements. Reproduction 2003;Suppl 61:27–36. https://doi.org/10.1530/biosciprocs.5.003.
- [5] Perera BMAO. Reproductive cycles of buffalo. Anim Reprod Sci 2011;124: 194–9. https://doi.org/10.1016/j.anireprosci.2010.08.022.
- [6] Vale WG. Effects of environment on buffalo reproduction. Ital J Anim Sci 2007;6(Suppl):130–42. https://doi.org/10.4081/ijas.2007.s2.130.
- Zicarelli L. Buffalo milk: its properties, dairy yield and mozzarella production. Vet Res Commun 2004;28(Suppl 1). https://doi.org/10.1023/B: VERC.0000045390.81982.4d. 127–35.
- [8] Altieri S, Saiano K, Biondi M, Ricci P, Lubritto C. Traceability of 'Mozzarella di Bufala Campana' production chain by means of carbon, nitrogen and oxygen stable isotope ratios. J Sci Food Agric 2019. https://doi.org/10.1002/ jsfa.10100.
- [9] Mazzei P, Piccolo A. 1H HRMAS-NMR metabolomic to assess quality and traceability of mozzarella cheese from Campania buffalo milk. Food Chem 2012;132:1620–7. https://doi.org/10.1016/j.foodchem.2011.11.142.
- [10] Meat & Livestock Australia. Global snapshot beef 2019. https://www.mla. com.au/globalassets/mla-corporate/prices-markets/documents/os-markets/ export-statistics/jan-2019-snapshots/global-beef-snapshot-jan2019.pdf.
- [11] Naveena BM, Kiran M. Buffalo meat quality, composition, and processing characteristics: contribution to the global economy and nutritional security. Anim Front 2014;4:18–24. https://doi.org/10.2527/af.2014-0029.
- [12] El Debaky HA, Kutchy NA, Ul-Husna A, Indriastuti R, Akhter S, Purwantara B, et al. Review: potential of water buffalo in world agriculture: challenges and opportunities. Appl Anim Sci 2019;35:255–68. https://doi.org/10.15232/ aas.2018-01810.
- [13] Deb GK, Nahar TN, Duran PG, Presicce GA. Safe and sustainable traditional production: the water buffalo in Asia. Front Environ Sci 2016;4:38. https:// doi.org/10.3389/fenvs.2016.00038.
- [14] Escarcha JF, Lassa JA, Palacpac EP, Zander KK. Understanding climate change impacts on water buffalo production through farmers' perceptions. Clim Risk Manag 2018;20:50–63. https://doi.org/10.1016/j.crm.2018.03.003.
- [15] UN sustainable development Goals. https://www.un.org/ sustainabledevelopment/sustainable-development-goals/.
- [16] Barile VL. Improving reproductive efficiency in female buffalo. Livest Prod Sci 2005;92:183-94. https://doi.org/10.1016/j.livprodsci.2004.06.014.
- [17] Warriach HM, McGill DM, Bush RD, Wynn PC, Chohan KR. A review of recent developments in buffalo reproduction. Asian Aust J Anim Sci 2015;28:451–5. https://doi.org/10.5713/ajas.14.0259.
- [18] Pirondi AN, Teixeira CMC, Lima ES, Valente TNP, Deminicis BB, Bezerra FC, et al. Reproductive characteristics of buffaloes: a review. J Agric Sci 2019;11: 167–77. https://doi.org/10.5539/jas.v11n13p167.
- [19] Terzano GM, Barile VL, Borghese A. Overview on reproductive endocrine aspects in buffalo. J Buffalo Sci 2012;1:126–38.
- [20] Campanile G, Neglia G, Vecchio D, Russo M, Zicarelli L. Pregnancy in buffalo cows. In: O'Leary Marie, Arnett John, editors. Pregnancy protein research. Nova Science Publishers; 2009. p. 31–91.
- [21] Prakash BS, Sarkar M, Paul V, Mishra DP, Mishra A, Meyer HHD. Postpartum endocrinology and prospects for fertility improvement in the lactating riverine buffalo (*Bubalus bubalis*) and yak (*Poephagus grunniens L.*). Livest Prod Sci 2005;98:13–23. https://doi.org/10.1016/j.livprodsci.2005.10.014.
- [22] Zicarelli L. Can we consider buffalo a non-precocious and hypofertile species? Ital J Anim Sci 2007;6(Suppl 2):143–54. https://doi.org/10.4081/ ijas.2007.s2.143.
- [23] Randel RD. Unique reproductive traits of Brahman and Brahman based cows. Tex. A &M Univ Agric Res Ext Cent 1990:60–82. https://animal.ifas.ufl.edu/ beef_extension/bcsc/1990/docs/randel_cows.pdf.
- [24] Zicarelli L. Reproductive seasonality in buffalo. In: Proceedings of third international course of biotechnology in buffalo reproduction, napoli 6-10 october. Bubalus bubalis; 1997. p. 29–52. 4(Suppl).
- [25] Phogat JB, Pandey AK, Singh I. Seasonality in buffaloes reproduction. Int J Plant Anim Environ Sci 2016;6:46–54.
- [26] Wankhade PR, Diwakar Kumar V, Talokar AJ, Aderao GN, Miranda GD, Gourkhede DP. Effect of photoperiod on the performance of buffaloes: a review. J Entomol Zool Stud 2019;7:177–80.
- [27] Campanile G, Neglia G, D'Occhio MJ. Embryonic and fetal mortality in river buffalo (Bubalus bubalis). Theriogenology 2016;86:207–13. https://doi.org/

10.1016/j.theriogenology.2016.04.033.

- [28] Campanile G, Vecchio D, Neglia G, Bella A, Prandi A, Senatore EM, et al. Effect of season, late embryonic mortality and progesterone production on pregnancy rates in pluriparous buffaloes (Bubalus bubalis) after artificial insemination with sexed semen. Theriogenology 2013;79:653–9. https:// doi.org/10.1016/j.theriogenology.2012.11.020.
- [29] Di Francesco S, Neglia G, Vecchio D, Rossi P, Russo M, Zicarelli L, et al. Influence of season on corpus luteum structure and function and Al outcome in the Italian Mediterranean buffalo (*Bubalus bubalis*). Theriogenology 2012;78: 1839–45. https://doi.org/10.1016/j.theriogenology.2012.07.022.
- [30] Zicarelli L. Influence of season on buffalo production. In: Presicce GA, editor. The buffalo (*Bubalus bubalis*) – production and research. Bentham Books; 2017. p. 196–224.
- [31] Campanile G, Baruselli PS, Neglia G, Vecchio D, Gasparrini B, Gimenes LU, et al. Ovarian function in the buffalo and implications for embryo development and assisted reproduction. Anim Reprod Sci 2010;121:1–11. https:// doi.org/10.1016/j.anireprosci.2010.03.012.
- [32] Das GK, Khan FA. Summer anoestrus in buffalo a review. Reprod Domest Anim 2010;45:e483–94. https://doi.org/10.1111/j.1439-0531.2010.01598.x.
- [33] Dash S, Chakravarty AK, Singh A, Upadhyay A, Singh M, Yousuf S. Effect of heat stress on reproductive performances of dairy cattle and buffaloes: a review. Vet World 2016;9. EISSN:2231-0916, veterinaryworld.org/vol.9/ march-2016/3.pdf.
- [34] Aksoy M, Kaya A, Ucar M, Lehimcioglu N, Tekeli T. Effect of seasonal condition on oestrus occurrence and postpartum period in Anatolian water buffaloes. Dtsch Tierarztl Wochenschr 2002;109:416–8.
- [35] Ramadan SI. Effect of some genetic and non-genetic factors on productive and reproductive traits of Egyptian buffaloes. J Adv Vet Anim Res 2018;5: 372–80. https://doi.org/10.5455/javar.2018.e287.
 [36] Rossi P, Vecchio D, Neglia G, Di Palo R, Gasparrini B, D'Occhio MJ, et al.
- [36] Rossi P, Vecchio D, Neglia G, Di Palo R, Gasparrini B, D'Occhio MJ, et al. Seasonal fluctuations in the response of Italian Mediterranean buffaloes to synchronization of ovulation and timed artificial insemination. Theriogenology 2014;82:132–7. https://doi.org/10.1016/ j.theriogenology.2014.03.005.
- [37] Vecchio D, Neglia G, Gasparrini B, Russo M, Pacelli C, Prandi A, et al. Corpus luteum development and function and relationship to pregnancy during the breeding season in the Mediterranean buffalo. Theriogenology 2012;77: 1811–5. https://doi.org/10.1016/j.theriogenology.2011.12.025.
- [38] Neglia G, Gasparrini B, Vecchio D, Boccia L, Varricchio E, Di Palo R, et al. Long term effect of ovum pick-up in buffalo species. Anim Reprod Sci 2011;123: 180–6. https://doi.org/10.1016/j.anireprosci.2011.01.011.
- [39] de Carvalho NAT, Soares JG, Baruselli PS. Strategies to overcome seasonal anestrus in water buffalo. Theriogenology 2016;86:200-6. https://doi.org/ 10.1016/j.theriogenology.2016.04.032.
- [40] Monteiro BM, de Souza DC, de Vasconcellos GSFM, de Carvalho NAT, Baruselli PS. Effect of season on dairy buffalo reproductive performance when using P4/E2/eCG-based fixed-time artificial insemination management. Theriogenology 2018;119:275–81. https://doi.org/10.1016/ j.theriogenology.2018.07.004.
- [41] Baruselli PS, Soares JG, Bayeux BM, Silva JCB, Mingoti RD, Carvalho NAT. Assisted reproductive technologies (ART) in water buffalo. Anim Reprod 2018;15(Suppl1):971–83. https://doi.org/10.21451/1984-3143-AR2018-0043.
- [42] Abdoon AS, Gabler C, Holder C, Kandil OM, Einspanier R. Seasonal variations in developmental competence and relative abundance of gene transcripts in buffalo (*Bubalus bubalis*) oocytes. Theriogenology 2014;82:1055–67. https:// doi.org/10.1016/j.theriogenology.2014.07.008.
- [43] Khan FA, Das GK, Pande M, Sarkar M, Mahapatra RK, Shankar U. Alterations in follicular fluid estradiol, progesterone and insulin concentrations during ovarian acyclicity in water buffalo (*Bubalus bubalis*). Anim Reprod Sci 2012;130:27–32. https://doi.org/10.1016/j.anireprosci.2011.12.020.
- [44] Sheth AR, Wadadekar KB, Moodbidri SB, Janakiraman K, Parameswaran M. Seasonal alterations in the serum prolactin and LH levels in the water buffalo. Curr Sci 1978;47:75–7.
- [45] Roy KS, Prakash BS. Changes in endocrine profiles during ovsynch and ovsynch plus norprolac treatment in Murrah buffalo heifers at hot summer season. Trop Anim Health Prod 2009;41:677–87. https://doi.org/10.1007/ s11250-008-9241-3.
- [46] Roy KS, Prakash BS. Seasonal variation and circadian rhythmicity of the prolactin profile during the summer months in repeat-breeding Murrah buffalo heifers. Reprod Fertil Dev 2007;19:569–75. https://doi.org/10.1071/ rd06093.
- [47] Campanile G, Avallone L, d'Angelo A, Di Palo R, Di Meo C. Influence of the season and of the number of days after calving on the pattern of thyroid hormones in buffalo cows. Fourth World Buffalo Congr 1994;3:564–6.
- [48] D'Occhio MJ, Baruselli PS, Campanile G. Influence of nutrition, body condition, and metabolic status on reproduction in female beef cattle: a review. Theriogenology 2019;125:277–84. https://doi.org/10.1016/ j.theriogenology.2018.11.010.
- [49] D'Occhio MJ, Baruselli PS, Campanile G. Metabolic health, the metabolome and reproduction in female cattle: a review. Ital J Anim Sci 2019;18:858–67. https://doi.org/10.1080/1828051X.2019.1600385.
- [50] Salzano A, Gasparrini B, Vecchio D, Longobardi V, Baruselli PS, Balestrieri A, et al. Effect of photoperiod on follicular IGF-1 and oocyte quality independently of metabolic status in buffalo heifers. Ital J Anim Sci 2019;18:949–56.

https://doi.org/10.1080/1828051X.2019.1588793.

- [51] Warriach HM, Channa AA, Ahmad N. Effect of oestrus synchronization methods on oestrus behavior, timing of ovulation and pregnancy rate during the breeding and low breeding seasons in Nili-Ravi buffaloes. Anim Reprod Sci 2008;107:62-7. https://doi.org/10.1016/j.anireprosci.2007.06.007.
- [52] Qayyum A, Arshad U, Yousuf MR, Ahmad N. Effect of breeding method and season on pregnancy rate and embryonic and fetal losses in lactating Nili-Ravi buffaloes. Trop Anim Health Prod 2018;50:555–60. https://doi.org/ 10.1007/s11250-017-1468-4.
- [53] Ramadan T, Ghuman S, Singh I. Melatonin and reproductive seasonality in buffalo. Lap Lambert Academic Publishing; 2016, ISBN 978-3-659-97479-3.
- [54] Emet M, Ozcan H, Ozel L, Yayla M, Halici Z, Hacimuftuoglu A. A review of melatonin, its receptors and drugs. Eurasian J Med 2016;48:135–41. https:// doi.org/10.5152/eurasianjmed.2015.0267.
- [55] Zhao D, Yu T, Shen Y, Liu Q, Zhao Z, Sharma R, et al. Melatonin synthesis and function: evolutionary history in animals and plants. Front Endocrinol 2019;10:249. https://doi.org/10.3389/fendo.2019.00249.
- [56] Ramadan TA. Role of melatonin in reproductive seasonality in buffalo. Intech Open Sci 2017;5:87–106. https://doi.org/10.5772/intechopen.69549.
 [57] Reiter RJ, Tan DX, Manchester LC, Paredes SD, Mayo JC, Sainz RM. Melatonin
- [57] Reiter RJ, Tan DX, Manchester LC, Paredes SD, Mayo JC, Sainz RM. Melatonin and reproduction revisited. Biol Reprod 2009;81:445–56. https://doi.org/ 10.1095/biolreprod.108.075655.
- [58] Weems PW, Goodman RL, Lehman MN. Neural mechanisms controlling seasonal reproduction: principles derived from the sheep model and its comparison with hamsters. Front Neuroendocrinol 2015;37:43–51. https:// doi.org/10.1016/j.yfrne.2014.12.002.
- [59] Pandi-Perumal SR, Srinivasan V, Maestroni GJM, Cardinali DP, Poeggeler B, Hardeland R. Melatonin: nature's most versatile biological signal? FEBS J 2006;273:2813-38. https://doi.org/10.1111/j.1742-4658.2006.05322.x.
- [60] Singh M, Jadhav HR. Melatonin: functions and ligands. Drug Discov Today 2014;19:1410–8. https://doi.org/10.1016/j.drudis.2014.04.014.
- [61] Gonzalez-Arto M, Hamilton TRS, Gallego M, Gaspar-Torrubia E, Aguilar D, Serrano-Blesa E, et al. Evidence of melatonin synthesis in the ram reproductive tract. Andrology 2016. https://doi.org/10.1111/andr.12117. 4_163-71.
- [62] Dubocovich ML, Markowska M. Functional MT₁ and MT₂ melatonin receptors in mammals. Endocrine 2005;27:101–10. https://doi.org/10.1385/ENDO:27: 2:101.
- [63] Prendergast BJ. MT1 melatonin receptors mediate somatic, behavioral, and reproductive neuroendocrine responses to photoperiod and melatonin in Siberian hamsters (*Phodopus sungorus*). Endocrinology 2010;151:714–21. https://doi.org/10.1210/en.2009-0710.
- [64] Liu J, Clough SJ, Hutchinson AJ, Adamah-Biassi EB, Popovska-Gorevski M, Dubocovich ML. MT1 and MT2 melatonin receptors: a therapeutic perspective. Annu Rev Pharmacol Toxicol 2016;56:361–83. https://doi.org/10.1146/ annurev-pharmtox-010814-124742.
- [65] Gobbi G, Comai S. Differential function of melatonin MT1 and MT2 receptors in REM and NREM sleep. Front Endocrinol 2019;10:87. https://doi.org/ 10.3389/fendo.2019.00087.
- [66] Soni N, Pandey AK, Kumar A, Verma A, Kumar S, Gunwant P, et al. Expression of MTNR1A, steroid (ERα, ERβ, and PR) receptor gene transcripts, and the concentration of melatonin and steroid hormones in the ovarian follicles of buffalo. Domest Anim Endocrinol 2019;(Jul 3):106371. https://doi.org/ 10.1016/j.domaniend.2019.06.003 [Epub ahead of print].
- [67] Frungieri MB, Calandra RS, Rossi SP. Local actions of melatonin in somatic cells of the testis. Int J Mol Sci 2017;18:1170. https://doi.org/10.3390/ ijms18061170.
- [68] Yu K, Deng SL, Sun TC, Li YY, Liu YX. Melatonin regulates the synthesis of steroid hormones on male reproduction: a review. Molecules 2018;23:447. https://doi.org/10.3390/molecules23020447.
- [69] Tabecka-Lonczynska A, Mytych J, Solek P, Kulpa M, Koziorowski M. New insight on the role of melatonin receptors in reproductive processes of seasonal breeders on the example of mature male European bison (*Bison bonasus*, Linnaeus 1758). J Photochem Photobiol B Biol 2017;173:84–91. https://doi.org/10.1016/j.jphotobiol.2017.05.026.
- [70] McGuire NL, Kangas K, Bentley GE. Effects of melatonin on peripheral reproductive function: regulation of testicular GnIH and testosterone. Endocrinology 2011;152:3461–70. https://doi.org/10.1210/en.2011-1053.
- [71] Tamura H, Nakamura Y, Korkmaz A, Manchester LC, Tan DX, Sugino N, et al. Melatonin and the ovary: physiological and pathophysiological implications. Fertil Steril 2009;92:328–43. https://doi.org/10.1016/ j.fertnstert.2008.05.016.
- [72] Tamura H, Takasaki A, Taketani T, Tanabe M, Kizuka F, Lee L, et al. The role of melatonin as an antioxidant in the follicle. J Ovarian Res 2012;5:5. ovarianresearch.com/content/5/1/5.
- [73] Minguini IP, Luquetti CM, Baracat MCP, Maganhin CC, Nunes CO, Simoes RS, et al. Melatonin effects on ovarian follicles: a systematic review. Rev Assoc Med Bras 2019;65:1122–7. https://doi.org/10.1590/1806-9282.65.8.1122.
- [74] Reina M, Martinez A. A new free radical scavenging cascade involving melatonin and three of its metabolites (3OHM, AFMK and AMK). Comput Theor Chem 2018;1123:111–8. https://doi.org/10.1016/ j.comptc.2017.11.017.
- [75] Reiter RJ, Tan DX, Fuentes-Broto L. Melatonin: a multitasking molecule. Prog Brain Res 2010;181:127–51. https://doi.org/10.1016/S0079-6123(08)81008-4.

- [76] Pieri C, Marra M, Moroni F, Recchioni R, Marcheselli F. Melatonin: a peroxyl radical scavenger more effective than vitamin E. Life Sci 1994;55:271–6. https://doi.org/10.1016/0024-3205(94)00666-0.
- [77] Rodriguez C, Mayo JC, Sainz RM, Antolin I, Herrera F, Martin V, et al. Regulation of antioxidant enzymes: a significant role for melatonin. J Pineal Res 2004;36:1-9. https://doi.org/10.1046/j.1600-079x.2003.00092.x.
- [78] Reiter RJ, Tan DX, Manchester LC, Qi W. Biochemical reactivity of melatonin with reactive oxygen and nitrogen Species: a review of the evidence. Cell Biochem Biophys 2001;34:237–56. https://doi.org/10.1385/CBB:34:2:237.
- [79] Tan DX, Manchester LC, Terron MP, Flores LJ, Reiter RJ. One molecule, many derivatives: a never-ending interaction of melatonin with reactive oxygen and nitrogen species? J Pineal Res 2007;42:28–42. https://doi.org/10.1111/ j.1600-079X.2006.00407.x.
- [80] Medrano A, Contreras CF, Herrera F, Alcantar-Rodriguez A. Melatonin as an antioxidant preserving sperm from domestic animals. Asian Pac J Reprod 2017;6:241-6. https://doi.org/10.4103/2305-0500.217317.
- [81] Jan MH, Das GK, Khan FA, Singh J, Bashir ST, Khan S, et al. Evaluation of follicular oxidant-antioxidant balance and oxidative damage during reproductive acyclicity in water buffalo (*Bubalus bubalis*). Asian Pac J Reprod 2014;3:35–40. https://doi.org/10.1016/S2305-0500(13)60182-7.
- [82] Kumar A, Mehrotra S, Singh G, Narayanan K, Das GK, Soni YK, et al. Sustained delivery of exogenous melatonin influences biomarkers of oxidative stress and total antioxidant capacity in summer-stressed anestrous water buffalo (*Bubalus bubalis*). Theriogenology 2015;83:1402–7. https://doi.org/10.1016/ j.theriogenology.2014.12.023.
- [83] Cebrian-Perez JA, Casao a, Gonzalez-Arto M, dos Santos Hamilton TR, Perez-Pe R, Muino-Blanco T. Melatonin in sperm biology: breaking paradigms. Reprod Domest Anim 2014;49:11–21. https://doi.org/10.1111/rda.12378.
- [84] El-Raey M, Badr MR, Rawash ZM, Darwish GM. Evidences for the role of melatonin as a protective additive during buffalo semen freezing. Am J Anim Vet Sci 2014;9:252–62. https://doi.org/10.3844/ajavssp.2014.252.262.
- [85] Di Francesco S, Mariotti E, Tsantarliotou M, Sattar A, Venditto I, Rubessa M, et al. Melatonin promotes *in vitro* sperm capacitation in buffalo (Bubalus bubalis). Reprod Fertil Dev 2009;22:311–2. https://doi.org/10.1071/ RDv22n1Ab311.
- [86] Ramadan TA, Kumar D, Ghuman SS, Singh I. Melatonin-improved buffalo semen quality during nonbreeding season under tropical condition. Domest Anim Endocrinol 2019;68:119–25. https://doi.org/10.1016/ j.domaniend.2019.01.010.
- [87] Perumal P, Chang S, Baruah KK, Srivastava N. Administration of slow release exogenous melatonin modulates oxidative stress profiles and in vitro fertilizing ability of the cryopreserved mithun (*Bos frontalis*) spermatozoa. Theriogenology 2018;120:79–90. https://doi.org/10.1016/ j.theriogenology.2018.07.033.
- [88] Tabecka-Longczynska A, Mytych J, Solek P, Kulpa M, Koziorowski M. New insight on the role of melatonin receptors in reproductive processes of seasonal breeders on the example of mature male European bison (*Bison Bonasis*, Linnaeus 1758). J Photochem Photobiol B Biol 2017;173:84–91. https://doi.org/10.1016/j.jphotobiol.2017.05.026.
- [89] Gonzalez-Arto M, Vicente-Carrillo A, Martinez-Pastor F, Fernandez-Alegre E, Roca J, Miro J. et al. Melatonin receptors MT₁ and MT₂ are expressed in spermatozoa from several seasonal and nonseasonal breeder species. Theriogenology 2016;86:1958–68. https://doi.org/10.1016/ j.theriogenology.2016.06.016.
- [90] Xiao L, Hu J, Song L, Zhang Y, Dong W, Jiang Y, et al. Profile of melatonin and its receptors and synthesizing enzymes in cumulus—oocyte complexes of the developing sheep antral follicle – a potential estradiol-mediated mechanism. Reprod Biol Endocrinol 2019;17:1. https://doi.org/10.1186/s12958-018-0446-7.
- [91] Xiao L, Hu J, Zhao X, Song L, Zhang Y, Dong W, et al. Expression of melatonin and its related synthase and membrane receptors in the oestrous corpus luteum and corpus luteum verum of sheep. Reprod Domest Anim 2018;53: 1142–8. https://doi.org/10.1111/rda.13218.
- [92] Pedreros M, Ratto M, Guerra M. Expression of functional melatonin MT1 receptors in equine luteal cells: *in vitro* effects of melatonin on progesterone secretion. Reprod Fertil Dev 2011;23:417–23. https://doi.org/10.1071/ RD10137.
- [93] Wang SJ, Liu WJ, Wang LK, Pang XS, Yang LG. The role of Melatonin receptor MTNR1A in the action of melatonin on bovine granulosa cells. Mol Reprod Dev 2017;84:1140–54. https://doi.org/10.1002/mrd.22877.
- [94] Wang X, Meng K, He Y, Wang H, Zhang Y, Quan F. Melatonin stimulates STAR expression and progesterone production via activation of the PI3K/AKT pathway in bovine theca cells. Int J Biol Sci 2019;15:404–15. https://doi.org/ 10.7150/ijbs.27912.
- [95] Zhang W, Wang Z, Zhang L, Zhang Z, Chen J, Chen W, et al. Melatonin stimulates the secretion of progesterone along with the expression of cholesterol side-chain cleavage enzyme (P450scc) and steroidogenic acute regulatory protein (StAR) in corpus luteum of pregnant sows. Theriogenology 2018;108:297–305. https://doi.org/10.1016/ j.theriogenology.2017.12.026.
- [96] Riaz H, Yousuf MR, Liang A, Hua GH, Yang L. Effect of melatonin on regulation of apoptosis and steroidogenesis in cultured buffalo granulosa cells. Anim Sci J 2019;90:473–80. https://doi.org/10.1111/asj.13152.
- [97] Nagina G, Asima A, Nemat U, Shamim A. Effect of melatonin on maturation capacity and fertilization of Nili-Ravi buffalo (Bubalus bubalis) oocytes. Open

Vet J 2016;6:128-34. https://doi.org/10.4314/ovj.v6i2.9.

- [98] Arul V, Gomathy VS, Brindha V. Effect of melatonin on *in vitro* maturation of buffalo (Bubalus Bubalis) oocytes. Int J Chem Stud 2017;5:1134–40.
- [99] Kang JT, Kool OJ, Kwon DK, Park HJ, Jang G, Kang SK, et al. Effects of melatonin on in vitro maturation of porcine oocyte and expression of melatonin receptor RNA in cumulus and granulosa cells. J Pineal Res 2009;46:22–8. https://doi.org/10.1111/j.1600-079X.2008.00602.x.
- [100] Casao A, Abecia JA, Cebrian-Perez JA, Muino-Blanco T, Vazquez MI, Forcada F. The effects of melatonin on *in vitro* oocyte competence and embryo development in sheep. Spanish J Agric Res 2010;8:35–41. https://doi.org/10.5424/ sjar/2010081-1141.
- [101] Tian X, Wang F, Zhang L, He C, Ji P, Wang J, et al. Beneficial effects of melatonin on the in vitro maturation of sheep oocytes and its relation to melatonin receptors. Int J Mol Sci 2017;18:834. https://doi.org/10.3390/ ijms18040834.
- [102] Barros VRP, Monte APO, Santos JM, Lins TLBG, Cavalcante AYP, Gouveia BB, et al. Melatonin improves development, mitochondrial function and promotes the meiotic resumption of sheep oocytes from *in vitro* grown secondary follicles. Theriogenology 2020. https://doi.org/10.1016/ j.theriogenology.2019.12.006 (in press).
- [103] Tsantarliotoun MP, Attanasio L, De Rosa A, Boccia L, Pellerano G, Gasparrini B. The effect of melatonin on bovine *in vitro* embryo development. Ital J Anim Sci 2007;6(Suppl 1):488–9. https://doi.org/10.4081/ijas.2007.1s.488.
- [104] Takada L, Junior AM, Mingoti GZ, Balieiro JCC, Coelho LA. Melatonin in maturation media fails to improve oocyte maturation, embryo development rates and DNA damage of bovine embryos. Sci Agric 2010;67:393–8. https:// doi.org/10.1590/S0103-90162010000400003.
- [105] Lopez-Damian EP, Jimenez-Medina JA, Alarcon MA, Lammoglia MA, Hernandez A, Galina CS, et al. Cryopreservation induces higher oxidative stress levels in *Bos indicus* embryos compared with *Bos Taurus*. Theriogenology 2020;143:74–81. https://doi.org/10.1016/ j.theriogenology.2019.12.001.
- [106] Earl CR, D'Occhio MJ, Kennaway DJ, Seamark RF. Temporal changes in the pattern of melatonin secretion in sheep held in constant darkness. J Pineal Res 1990;8:115–21.
- [107] Earl CR, D'Occhio MJ, Kennaway DJ, Seamark RF. Mechanisms controlling the offset of melatonin secretion in the Ewe. J Pineal Res 1990;8:49–56.
- [108] D'Occhio MJ, Suttie JM. The role of the pineal gland and melatonin in reproduction in male domestic ruminants. Anim Reprod Sci 1992;30: 135–55.
- [109] Singh A. Study on anoestrus and fertility response by using melatonin hormone in dairy buffaloes. Master Vet Sci, Nanaji Deshmukh veterinary science university, Jabalpur https://pdfs.semanticscholar.org/ebf6/ 8a92eadad80aea757bd663ef0e9ea258d899.pdf.
- [110] Ghuman SPS, Singh J, Honparkhe M, Dadarwal D, Dhaliwal GS, Jain AK. Induction of ovulation of ovulatory size non-ovulatory follicles and initiation of ovarian cyclicity in summer anoestrous buffalo heifers (*Bubalus bubalis*) using melatonin implants. Reprod Domest Anim 2010;45:600-7. https:// doi.org/10.1111/j.1439-0531.2008.01310.x.
- [111] Kumar A, Mehrotra S, Singh G, Maurya VP, Narayanan K, Mahla AS, et al. Supplementation of slow-release melatonin improves recovery of ovarian cyclicity and conception in summer anoestrous buffaloes (*Bubalus bubalis*). Reprod Domest Anim 2016;51:10–7. https://doi.org/10.1111/rda.12639.
- [112] Ramadan TA, Sharma RK, Phulia SK, Balhara AK, Ghuman SS, Singh I. Effectiveness of melatonin and controlled internal drug release device treatment on reproductive performance of buffalo heifers during out-of-breeding season under tropical conditions. Theriogenology 2014;82:1296–302. https://doi.org/10.1016/j.theriogenology.2014.08.014.
- [113] Ramadan TA, Sharma RK, Phulia SK, Balhara AK, Ghuman SS, Singh I. Effects of melatonin and controlled internal drug release device treatment on blood metabolites of buffalo heifers during out-of-breeding season under tropical conditions. Egypt J Anim Prod 2015;(Suppl):9–17.
- [114] Ramadan TA, Sharma RK, Phulia SK, Balhara AK, Chuman SS, Singh I. Manipulation of reproductive performance of lactating buffaloes using melatonin and controlled internal drug release device treatment during outof-breeding season under tropical conditions. Theriogenology 2016;86: 1048-53. https://doi.org/10.1016/j.theriogenology.2016.03.034.
- [115] Kavita, Phogat JB, Pandey AK, Balhara AK, Ghuman SS, Gunwant P. Effects of melatonin supplementation prior to Ovsynch protocol on ovarian activity and conception rates in anestrous Murrah buffalo heifers during out of breeding season. Reprod Biol 2018;18:161–8. https://doi.org/10.1016/ j.repbio.2018.03.001.
- [116] El-Mokadem MY, Nour El-Din ANM, Ramadan TA, Rashad AMA, Taha TA, Samak MA. Manipulation of reproductive seasonality using melatonin implantation in Anglo-Nubian does treated with controlled internal drug release and equine chorionic gonadotropin during the nonbreeding season. J Dairy Sci 2017;100:5028–39. https://doi.org/10.3168/jds.2016-12240.
- [117] deNicolo G, Morris ST, Kenyon PR, Morel PCH, Parkinson TJ. Melatoninimproved reproductive performance in sheep bred out of season. Anim Reprod Sci 2008;109:124–33. https://doi.org/10.1016/ j.anireprosci.2007.10.012.
- [118] Cheema RS, Kaur A, Ghuman SPS, Dhindsa S. Melatonin receptor 1A gene polymorphism in Murrah buffaloes with possible impact on summer anestrous. J Bio Innovat 2016;5:386–94.
- [119] Zetouni L, de Camargo GMF, Fonseca PDS, Cardoso DF, Gil FMM, Hurtado-

Lugo NA, et al. Polymorphisms in the *MTRN1A* gene and their effects on the productive and reproductive traits in buffaloes. Trop Anim Health Prod 2014;46:337–40. https://doi.org/10.1007/s11250-013-0493-1.

- [120] Pandey AK, Gunwant P, Soni N, Kavita Kumar S, Kumar A, Magotra A, et al. Genotype of MTNR1A gene regulates the conception rate following melatonin treatment in water buffalo. Theriogenology 2019;128:1–7. https:// doi.org/10.1016/j.theriogenology.2019.01.018.
- [121] Carcangiu V, Mura MC, Pazzola M, Vacca GM, Paludo M, Marchi B, et al. Characterization of the Mediterranean Italian buffaloes melatonin receptor 1A (MTNR1A) gene and its association with reproductive seasonality. Theriogenology 2011;76:419–26. https://doi.org/10.1016/ j.theriogenology.2011.02.018.
- [122] Luridiana S, Mura MC, Pazzola AM, Paludo M, Cosso G, Dettori ML, et al. Association between melatonin receptor 1A (*MTNR1A*) gene polymorphism and the reproductive performance of Mediterranean Italian buffaloes. Reprod Fertil Dev 2012;24:983-7. https://doi.org/10.1071/RD11297.
- [123] Paludo M, Mura MC, Luridiana S, Pazzola M, Daga C, Vacca GM, et al. Genotype of melatonin receptor MT1 (MTNR1A) and puberty in Mediterranean Italian Buffalo. Agric Conspectus Sci 2011;76:157–60. https://hrcak.srce.hr/ 72025.
- [124] Gunwant P, Pandey AK, Kumar A, Singh I, Kumar S, Phogat JB, et al. Polymorphism of melatonin receptor (*MTNR1A*) gene and its association with seasonal reproduction in water buffalo (*Bubalus bubalis*). Anim Reprod Sci 2018;199:51–9. https://doi.org/10.1016/j.anireprosci.2018.10.006.
- [125] Barbosa EM, Souza BB, Guimaraes RC, Azevedo JSN, Goncalves EC, Ribeiro HFL, et al. Novel polymorphism in exon 1 of the melatonin receptor gene unassociated with reproductive characteristics of buffaloes in the Amazon Region. Genet Mol Res 2016;15. https://doi.org/10.4238/ gmr.15028309.
- [126] Barbosa EM, Souza BB, Guimaraes RC, Azevedo JSN, Goncalves EC, Ribeiro HFL, et al. Polymorphism in the melatonin receptor gene in buffalo populations of the Brazilian Amazon. Genet Mol Res 2016;15. https://doi.org/ 10.4238/gmr.15027960.
- [127] Saxena VK, Jha BK, Meena AS, Naqvi SMK. Sequence analysis and identification of new variations in the coding sequence of melatonin receptor gene (MTNR1A) of Indian Chokla sheep breed. Meta Gene 2014;2:450–8. https://

doi.org/10.1016/j.mgene.2014.05.005.

- [128] Carcangiu V, Mura MC, Vacca GM, Pazzola M, Dettori ML, Luridiana S, et al. Polymorphism of the melatonin receptor MT1 gene and its relationship with seasonal reproductive activity in the Sarda sheep breed. Anim Reprod Sci 2009;116:65–72. https://doi.org/10.1016/j.anireprosci.2009.01.005.
- [129] Pelletier J, Bodin L, Hanocq E, Malpaux B, Teyssier J, Thimonier J, et al. Association between expression of reproductive seasonality and alleles of the gene for Mel_{1a} receptor in the Ewe. Biol Reprod 2000;62:1096–101. https://doi.org/10.1095/biolreprod62.4.1096.
- [130] Giantsis IA, Laliotis GP, Stoupa O, Avdi M. Polymorphism of the melatonin receptor 1A (MNTR1A) gene and association with seasonality of reproductive activity in a local Greek sheep breed. J Biol Res-Thessaloniki 2016;23:9. https://doi.org/10.1186/s40709-016-0050-y.
- [131] Notter DR, Cockett NE, Hadfield TS. Evaluation of *melatonin receptor 1a* as a candidate gene influencing reproduction in an autumn-lambing sheep flock. J Anim Sci 2003;81:912–7. https://doi.org/10.2527/2003.814912x.
- [132] de Camargo GMF, Aspilcueta-Borquis RR, Fortes MRS, Porto-Neto R, Cardoso DF, Santos DJA, et al. Prospecting major genes in dairy buffaloes. BMC Genom 2015;16:872. https://doi.org/10.1186/s12864-015-1986-2.
 [133] Li J, Liu J, Campanile G, Plastow G, Zhang C, Wang Z, et al. Novel insights into
- [133] Li J, Liu J, Campanile G, Plastow G, Zhang C, Wang Z, et al. Novel insights into the genetic basis of buffalo reproductive performance. BMC Genom 2018;19: 814. https://doi.org/10.1186/s12864-018-5208-6.
- [134] Li J, Liu S, Plastow G, Zhang C, Wang Z, et al. Integrating RNA-seq and GWAS reveals novel genetic mutations for buffalo reproductive traits. Anim Reprod Sci 2018;197:290–5. https://doi.org/10.1016/ j.anireprosci.2018.08.041.
- [135] Barile VL. Reproductive efficiency in female buffaloes. In: Borghese A, editor. Buffalo production and research. FAO Reu Technical Series; 2005. p. 77–108. vol. 67.
- [136] Parmeggiani A, Di Palo R, Zicarelli L, Campanile G, Esposito L, Seren E, et al. Melatonina e stagionalità riproduttiva della bufala. Agric Ric 1994;53:41–8.
- [137] Di Palo R, Parmeggiani A, Spadetta M, Campanile G, Esposito L, Seren E, et al. Influence of changing farm on the repeatability of melatonin plasma level in Italian Mediterranean buffalo. In: Proc 5th world buffalo congress, Caserta, Italy; October 13-16, 1997. p. 758–61.