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92	Abstract	<p><b>Purpose of Review:</b> We highlight new entities of congenital diarrheal disorders (CDDs) and progresses in understanding of functionally related genes, opening new diagnostic and therapeutic perspectives.</p> <p><b>Recent Findings:</b> The more significant advances have been made in field of pathogenesis, encouraging a better understanding not only of these rare diseases but also of more common pathogenetic mechanisms.</p> <p><b>Summary:</b> CDDs represent an evolving group of rare chronic</p>	

enteropathies with a typical onset early in the life. Usually, severe chronic diarrhea is the main clinical manifestation, but in other cases, diarrhea is only a component of a more complex systemic disease. The number of conditions has gradually increased, and many new genes have been indentified and functionally related to CDDs, opening new diagnostic and therapeutic perspectives. Advances in molecular analysis procedures have modified the diagnostic approach in CDDs, leading to a reduction in invasive and expensive procedures.

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93	Keywords separated by ' - '	Chronic diarrhea - Genes - Molecular analysis - Mutations - Children
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# New Insights and Perspectives in Congenital Diarrheal Disorders

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## Abstract

*Purpose of Review* We highlight new entities of congenital diarrheal disorders (CDDs) and progresses in understanding of functionally related genes, opening new diagnostic and therapeutic perspectives.

*Recent Findings* The more significant advances have been made in field of pathogenesis, encouraging a better understanding not only of these rare diseases but also of more common pathogenetic mechanisms.

*Summary* CDDs represent an evolving group of rare chronic enteropathies with a typical onset early in the life. Usually, severe chronic diarrhea is the main clinical manifestation, but in other cases, diarrhea is only a component of a more complex systemic disease. The number of conditions has gradually increased, and many new genes have been identified and functionally related to CDDs, opening new

diagnostic and therapeutic perspectives. Advances in molecular analysis procedures have modified the diagnostic approach in CDDs, leading to a reduction in invasive and expensive procedures.

**Keywords** Chronic diarrhea · Genes · Molecular analysis · Mutations · Children

## Introduction

Congenital diarrheal disorders (CDDs) are a group of rare hereditary enteropathies, characterized by a typical onset during the first days of life [1••]. Although, most of these diseases present similar clinical features, the causes, the management, and prognosis of various forms of CDDs are very different. For most of these conditions, a severe chronic diarrhea is the primary clinical manifestation; more rarely, diarrhea is only one component of a multiorgan more complex picture. In most cases, an appropriate therapy should be initiated immediately in order to prevent dehydration and serious short- and long-term complications [1••]. There are also milder forms of CDDs, with a less severe clinical picture, diagnosed in later ages, typically due to mutations that less severely impair the residual activity of the disease-protein. To date, genes responsible for disease are known in most cases of CDDs. Therefore, molecular analysis has assumed a key role in the diagnostic approach to a patient suspected of CDDs and, in some cases, in the prediction of the outcome of the disease (genotype–phenotype correlation). Evolving knowledge of the pathogenesis of CDDs suggests the utility of a classification system based on the main pathogenetic mechanism, which could help the approach to these patients (Fig. 1). This classification comprises four groups of disorders:

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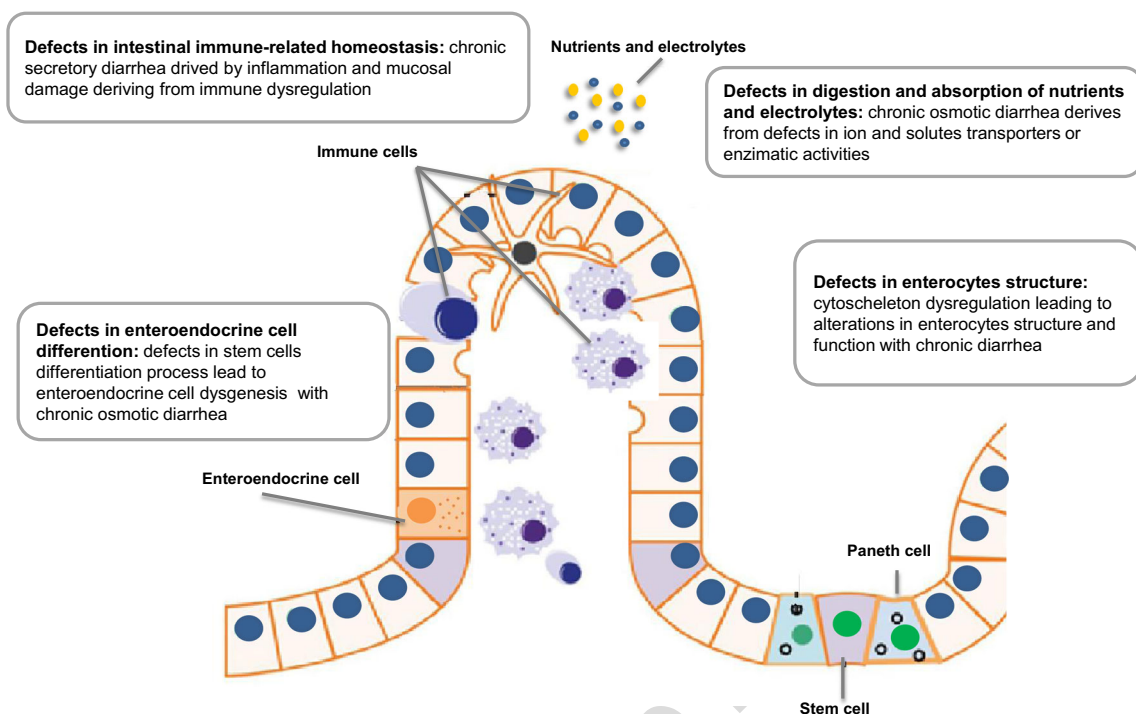
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**Fig. 1** The improving knowledge about the pathogenesis of congenital diarrheal disorders has inspired a new classification that could help the diagnostic and therapeutic approach to these conditions

- 57 I. Defects in digestion and absorption of nutrients and
- 58 electrolytes
- 59 II. Defects in enterocyte structure
- 60 III. Defects in enteroendocrine cell differentiation
- 61 IV. Defects in intestinal immune-related homeostasis.

conditions have been described. No histological or ultrastructural defects are generally observed in these patients at gut level [1•].

**Familial Diarrhea Syndrome**

**Main Clinical Features** This condition has been described in 32 members of a Norwegian family, and is characterized by early-onset chronic diarrhea and meteorism. In a subset of patients, abdominal pain and dysmotility have been described as main features [4•]. Patients have an increased risk to develop Crohn’s disease and intestinal obstruction resulting from volvulus, adhesional bands, and/or ileal inflammation [5].

**Genotype** All affected members have an activating heterozygous missense mutation (p.Ser840Ile) in the GUCY2C gene, which encodes for the intestinal guanylate cyclase receptor for uroguanylin, guanylin, and heat-stable enterotoxins [4•]. Activation of the guanylate cyclase C (GC-C) receptor increases cellular levels of cyclic guanosine monophosphate (cGMP), leading to phosphorylation of the cystic fibrosis transmembrane conductance regulator (CFTR) channel [5]. The efflux of Cl<sup>-</sup> and water into the intestinal lumen, with reduced sodium ion (Na<sup>+</sup>) absorption owing to inhibition of the Na<sup>+</sup>-H<sup>+</sup> exchanger 3 (NHE3) [5], leads to a severe chronic secretory diarrhea. GC-C signaling also has implications for renal electrolyte homeostasis, intestinal cell proliferation, apoptosis, intestinal barrier function, and inflammation [5].

We review new CDD entities and advances understanding of functionally related genes that are opening new diagnostic and therapeutic perspectives, underlining the crucial role of molecular analysis.

**Defects in Absorption and Transport of Nutrients and Electrolytes**

Most of the CDDs belong to this group. These diseases derive from a defect in one of the main mechanisms of digestion or transport that leads to severe diarrhea and subsequent dehydration and weight loss early after birth. Specifically, the defect can be charged to brush border membrane, membrane carriers, pancreatic enzymes, lipid transport and metabolism, ribosomal proteins, and mitochondrial DNA [2••] (Table 1). The alteration of digestion or absorption of carbohydrates, proteins, and electrolytes results in osmotic diarrhea. Fecal pH <5, ion gap >50, and molecular analysis confirm the diagnosis. The prototypes of this group are glucose-galactose malabsorption and congenital chloride diarrhea [1••], but new

**Q3** t1.1 **Table 1** Main features of congenital diarrheal disorders: genetics and epidemiology

t1.2	Disease	Gene			Protein	Inheritance and incidence
		Name	OMIM number	Position		
t1.4	Defects in absorption and transport of nutrients and electrolytes					
t1.5	Congenital chloride diarrhea	SLC26A3	126650	7q31.1	Cl <sup>-</sup> /base exchanger	AR, sporadic; common in some ethnic groups
t1.6	Congenital sodium diarrhea <sup>a</sup>	SLC29A3	182307	5p15.33	Na <sup>+</sup> -H <sup>+</sup> exchanger	AR, <1:1,000,000
t1.7	Congenital lactase deficiency	LCT	603202	2q21.3	Lactase-pherorizin hydrolase	AR, 1:60,000 in Finland; lower in other ethnic groups
t1.8	Sucrase-isomaltase deficiency	SI	609845	3q26.1	Isomaltase-sucrase	AR, 1:5000; higher in Greenland, Alaska, and Canada
t1.9	Maltase-glucoamylase deficiency	MGAM	154360	7q34	Maltase-glucoamylase	Only few cases described
t1.10	Glucose-galactose malabsorption	SLC5A1	182380	22q13.1	Na <sup>+</sup> /glucose cotransporter	AR, a few hundred cases described
t1.11	Fanconi-Bickel syndrome	SLC2A2	138160	3q26.2	Basolateral glucose transporter	AR, rare
t1.12	Acrodermatitis enteropathica	SLC39A4	607059	8q24.3	Zn <sup>2+</sup> transporter	AR, 1:500.000
t1.13	Lysinuric protein intolerance	SLC7A7	603593	14q11.2	Cationic amino acid transporter	AR, approximately 1:60,000 in Finland and Japan; rare in other ethnic groups
t1.14	Primary bile acid diarrhea	SLC10A2	601295	13q33.1	Ileal Na <sup>+</sup> /bile salt transporter	AR
t1.15		FGF-19	603891	11q13.3	Bile acids negative feedback	Only few cases described
t1.16	Enterokinase deficiency	TMPRSS15	606635	21q21.1	Proenterokinase	AR
t1.17	Abetalipoproteinemia	MTTP	157147	4q23	Microsomal triglyceride transfer protein	AR, about 100 cases described; higher frequency among Ashkenazi
t1.18	Hypobetalipoproteinemia	Apo B	107730	2p24.1	Apolipoprotein B 100/48	Autosomal codominant
t1.19	Chylomicron retention disease	SAR1B	607690	5q31.1	Intracellular chylomicron trafficking	AR, about 40 cases described
t1.20	Familial diarrhea syndrome	GUCY2C	601330	12p13.1-p12.3	Receptor for heat-stable enterotoxins	Described in 32 members of a Norwegian family
t1.21	Diarrhea-associated DGAT1 mutation	DGAT1	604900	8q24.3	Diacylglycerol acyltransferases	One family has been reported
t1.22	Defects in enterocyte structure					
t1.23	Microvillous inclusion disease	MYO5B	606540	18q21.1	Myosin VB	AR; rare; highest frequency among Navajo
t1.24		STX3	600876	11q12.1	Syntaxin3	AR; two patients described
t1.25	Congenital tufting enteropathy <sup>b</sup>	EPCAM	185535	2p21	Protein for cell-cell interaction	AR; 1:50-100.000; higher among Arabians
t1.26		SPINT2	605124	19q13.2	Serine protease inhibitor	<1/1,000,000
t1.27	Trichohepatoenteric syndrome (syndromic diarrhea)	TTC37	614589	5q15	Component of the SKI complex	AR; <1/1,000,000
t1.28		SKIV2L	600478	6p21.33	Helicase	AR; <1/1,000,000



t1.29 **Table 1** (continued)

Disease	Gene			Protein	Inheritance and incidence
	Name	OMIM number	Position		
t1.31	Defects in enteroendocrine cell differentiation				
t1.32	Enteric anendocrinosis	NEUROG3	604882	10q22.1	Transcriptional regulator AR; few cases described
t1.33	X-linked lissencephaly and MR	ARX	300382	Xp21.3	Homeodomain transcription factors X-linked
t1.34	Proprotein convertase 1/3 deficiency	PCSK1	162150	5q15	Neuroendocrine convertase AR; <1/1,000,000
t1.35	Mitchell–Riley Syndrome	RFX6	612659	6q22.1	Transcription factors AR
t1.36	Defects in intestinal immune-related homeostasis				
t1.37	Immunodysregulation, polyendocrinopathy, enteropathy, X-linked syndrome	FOXP3	300292	Xp11.23	Transcription factor X-linked
t1.38	IPEX-like disorders	CD25	147730	10p15.1	Interleukin-2 receptor, alpha chain AR
t1.39		STAT5b	604260	17q21.2	Transcriptional regulator AR
t1.40		STAT-1	600555	2q32.2	Transcriptional regulator AD, loss/gain of function
t1.41		ITCH	606409	20q11.22	Ubiquitin protein ligase AR (one family)
t1.42		LRBA	606453	4q31.3	Protein involving in apoptosis AR, three family described
t1.43		MALT-1	604860	18q21.32	Protein involving in NF-κB activation AR (one family)
t1.44	Early onset enteropathy with colitis	IL-10 IL-10Rα IL-10Rβ	124092 146933 123889	1q32.1 11q23.3 21q23.3	Cytokine or cytokine receptors AR

<sup>a</sup> Analysis of the intestinal brush border membrane of affected patients revealed that the condition is caused by a SLC9A3 loss-of-function mutations. SLC9A3 encodes Na<sup>+</sup>/H<sup>+</sup> antiporter 3 (NHE3), the major intestinal brush border Na<sup>+</sup>/H<sup>+</sup> exchanger. A syndromic form of CSD is characterized by the presence of choanal and intestinal atresias as well as recurrent corneal erosions. Small bowel histology frequently detects an epithelial “tufting” dysplasia. It is autosomal recessively inherited and associated to SPINT2 mutations [3]

<sup>b</sup> Congenital tufting enteropathy associated to EPCAM mutation is characterized by only intestinal involvement, while mutation in SPINT2 leads to a syndromic form with dysmorphic features, wooly hair, small birth weight and immune deficiency, and diarrhea with high sodium content in the stools

106	<b>Treatment</b> Total parenteral nutrition is the only therapeutic option at the moment [4•, 5].	maybe acting as bioactive signaling lipids or via a detergent-like action [7•]. Several differences, however, are apparent among the reported cases and regard the serum lipid profile, including triglycerides, the presence of digital clubbing and the onset of diarrhea. These phenotypic differences may reflect genotypic differences [7•].	120 121 122 123 124 125
108	<b>DGAT1-Deficiency-Related Diarrhea</b>		
109	<b>Main Clinical Features</b> Protein-losing enteropathy (PLE) is a clinical disorder of protein loss from the gastrointestinal system that results in hypoproteinemia and malnutrition.		
112	<b>Genotype</b> Patients with mutations in DGAT1 (which encodes acyl CoA/diacylglycerol acyltransferases 1) present aspects of both PLE and CDD [6]. DGATs catalyze the final step of triglyceride synthesis. Animal models lacking DGAT1 show delayed fat absorption with more fat reaching the distal gut. However, how DGAT1 deficiency causes diarrhea and protein losing enteropathy is still unknown, but it is possible that excess diacylglycerols or fatty acids could have a toxic role,	<b>Treatment</b> Total parenteral nutrition is the only therapeutic option at the moment [7•].	126 127
114		<b>Defects in Enterocyte Structure</b>	128
116		This group of CDDs includes microvillus inclusion disease (MVID), congenital tufting enteropathy (CTE), more recently trichohepatoenteric syndrome (THE) has added to the group (Table 1). During the last years, mutations in genes, involved	129 130 131 132

133 in intestinal epithelial physiology, have been associated with  
 134 different CDDs, opening new perspectives in understanding  
 135 the pathogenetic mechanism and in the clinical approach [1••].  
 136 Parenteral nutrition and intestinal transplantation are the only  
 137 therapeutic strategies available at the moment; they have re-  
 138 duced the morbidity and mortality rates of these diseases [1••].

139 **Microvillus Inclusion Disease**

140 **Main Clinical Features** Loss of apical microvilli and forma-  
 141 tion of microvillus inclusion in the cytoplasm of enterocytes  
 142 are the main hallmarks of MVID [8•]. These alterations lead to  
 143 persistent diarrhea, nutrient malabsorption, and failure to  
 144 thrive. In most cases (95%), symptoms develop within days  
 145 after birth, but there is a late-onset variant, which presents 2–  
 146 3 months postnatally. Extra intestinal symptoms could be  
 147 intrahepatic cholestasis and renal Fanconi syndrome. Some  
 148 individuals with MVID present less-severe digestive symp-  
 149 toms for reasons that are not clear [9••]. Intestinal biopsy is  
 150 the most important method to diagnose this disease. This will  
 151 display: features of villus atrophy, microvillus atrophy, and the  
 152 redistribution of CD10 and periodic acid Schiff (PAS)-stained  
 153 material from the brush border to intracellular sites in the  
 154 enterocytes. A definitive diagnosis includes analysis by elec-  
 155 tron microscopy (EM) for microvillus inclusions in the cyto-  
 156 plasm of enterocytes. It is interesting that microvillus inclu-  
 157 sions are also present in rectal biopsies, facilitating diagnosis  
 158 if a duodenal biopsy is not available [9••].

159 **Genotype** Loss of function mutations in the actin motor my-  
 160 osin Vb (MYO5B) is responsible for most cases of MVID.  
 161 Recently, mutations in the SNARE fusion protein syntaxin 3  
 162 (STX3) and STXBP2 were reported in the milder MVID var-  
 163 iant [10]. MYO5B encodes the actin-based motor protein myo-  
 164 sin Vb, which consists of an N-terminal actin binding motor  
 165 domain and a C-terminal tail domain that includes the  
 166 cargobinding domain. The myosin Vb cargobinding domain  
 167 binds selectively to small Rab GTPases, among which  
 168 RAB11A and RAB8A. MYO5b, in concert with RAB11A  
 169 and RAB8A associated with apical recycling endosomes  
 170 (AREs), in polarized epithelial cells controls the activity of  
 171 the small GTPase CDC42, and it modulates intestinal epithe-  
 172 lial cells polarity, apical trafficking, and microvilli growth  
 173 [10]. At the basis of MVID's pathogenesis, it was demonstrat-  
 174 ed an uncoupling of myosin Vb from RAB11A and RAB8A,  
 175 caused from the mutation of myosin Vb. Two other mutations  
 176 involved in MVID interested STX, which encodes the trans-  
 177 membrane protein syntaxin-3, or STXBP2, which encodes  
 178 Munc18-2 [11]. In enterocytes, syntaxin-3 is localized at the  
 179 apical cell-surface domain, where it, in concert with SNAP23  
 180 and Munc18-2, has the function of mediating the fusion of  
 181 transport vesicles with the apical plasma membrane. The mu-  
 182 tation STX3, responsible for MVID, leads to depletion of the

syntaxin-3 or the expression of a syntaxin-3 protein that lacks  
 its transmembrane domain, with the loss of its function.  
 STXBP2 mutations abolish the interaction of Munc18-2 with  
 syntaxin proteins [11].

**THE**

**Main Clinical Features** Trichohepatoenteric syndrome, also  
 called syndromic diarrhea, is a rare life-limiting autosomal  
 recessive bowel disorder [12]. Main symptoms are chronic  
 diarrhea, facial dysmorphism, trichothiodystrophy associated  
 or not with liver disease, hepatomegaly, siderosis, congenital  
 cardiac defects, and platelet anomalies [12]. Affected people  
 are susceptible to infection, because they might fail to produce  
 antibodies upon vaccination, or present with low immuno-  
 globulin levels. In 50% of all cases, it is possible to find mild  
 intellectual deficiency. Clinical dates and via biopsies of the  
 small intestine are useful to make diagnosis. The biopsies  
 display the typical histological and ultrastructural defects: vil-  
 lus atrophy and variable immune cell infiltration of the thin  
 layer of loose connective tissue that lies beneath the epitheli-  
 um [12].

**Genotype** Recent studies have shown that the mutations in-  
 volved in this disease are associated with TTC37 or SKIV2L  
 [13]. The gene's products of both TTC37 and SKIV2L are  
 human homologs of components of the yeast Ski complex,  
 which is linked with exosome-mediated degradation of aber-  
 rant messenger RNA (mRNA) and associated with transcrip-  
 tionally active genes. TTC37 (also called Thespin) encodes  
 the tetratricopeptide repeat protein 37 and it is expressed in  
 many tissues like vascular endothelium, lung and intestine,  
 but not in the liver. In enterocytes with TTC37 mutations,  
 the brush-border-associated NHE-2 and NHE-3, aquaporin  
 7, the Na<sup>+</sup>/Γ symporter, and the H<sup>+</sup>/K<sup>+</sup>-ATPase show reduced  
 expression or mislocalization to the apical cytoplasm, with  
 different patterns of mislocalization relative to their normal  
 pattern [12]. In conclusion, the loss of TTC37 results in the  
 defective trafficking and/or decreased expression of apical  
 transport proteins, including aquaporin 7.

The other mutation identified in the THE regard SKIV2L,  
 which encodes SKI2 homolog, superkiller viralicidic activity  
 2-like protein, which might be involved in antiviral activity by  
 blocking translation of poly (A)-deficient mRNAs. The mech-  
 anism, which is the base of the disease, is associated with loss  
 of function a cytoplasmic-exosome cofactor involved in vari-  
 ous mRNA decay pathways and required for normal cell  
 growth [12].

**Congenital Tufting Enteropathy**

**Main Clinical Features** Typical is the presence of epithelial  
 tufts that can be localized from the duodenum to the large

231	intestine [1••]. Patients affected by this condition have persistent diarrhea that presents immediately or just after birth, despite bowel rest, and total parenteral nutrition.	280
232		281
233		282
234	A subset of individuals with CTE displays a syndromic form of the disease that includes other signs and symptoms like dysmorphic features, wooly hair, punctate keratitis, atresias, reduced body size, and immune deficiency [14].	
235		
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238	Histological features of CTE reveal various degrees of villus atrophy, basement membrane abnormalities, disorganization of enterocytes and focal crowding at the villus tips, resembling tufts.	
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242	<b>Genotype</b> In CTE, the absence of epithelial cell adhesion molecule (EPCAM) in enterocytes is considered the most important diagnostic marker [15•]. EPCAM, expressed along the basolateral membranes, is a multifunctional transmembrane glycoprotein, and it has an important role in cell–cell adhesion, proliferation, and differentiation. In this disease, EPCAM protein levels in the intestine are decreased and all CTE-associated EPCAM mutations lead to loss of cell-surface EPCAM, either because of impaired plasma membrane targeting or because of truncation of the protein, both of which result in its secretion [15•].	
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253	Recent studies have demonstrated that in the EPCAM KO mouse intestine, E-cadherin, and beta-catenin, two adherens junction-associated proteins are also mislocalized, leading to disorganized transition from crypts to villi [15•]. Recently, it has been demonstrated that a second group of CTE individuals is characterized by mutations in SPINT2 [14]. SPINT2 encodes the transmembrane protein Kunitz-type 2 serine-protease inhibitor which is involved in epithelial regeneration, in the Nf-Kb and TGF-beta signaling pathways. The inhibition of trypsin-family serine peptidases, which are encoded by SPINT2, abolishes the stimulation of apical Na <sup>+</sup> transport by nonvoltage-gated sodium channel-1-alpha (SCNN1A) in polarized intestinal epithelial cells, which could contribute to secretory diarrhea. It is possible that such a mechanism is the basis of the syndromic form of congenital sodium diarrhea that is associated with SPINT2 mutations [14].	
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270	<b>Defects in Enteroendocrine Cell Differentiation</b>	
271	Abnormal enteroendocrine cell development or function and congenital malabsorptive diarrhea, associated or not with other systemic endocrine abnormalities, are the main features of this group of extremely rare forms of CDDs. Various null mouse models of each of these genes are associated with early postnatal mortality and occasionally diarrhea [1••]. Genes involved in this group are as follows: neurogenin-3 (NEUROG3), regulatory factor X-6 (RFX6), aristaless-related homeobox (ARX), and proprotein convertase subtilisin/kexin type 1 (PCSK1) (Table 1). Parenteral nutrition and intestinal transplant are the only two therapeutic options in these patients [1••, 16–19].	283
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	<b>Enteric Anendocrinosis and Mitchell–Riley Syndrome</b>	283
	<b>Main Clinical Features and Genotype</b> NEUROG3 controls the fate of enteric cells in both the pancreas and the intestine. Biallelic mutations in NEUROG3 are known to cause a rare but well-defined clinical syndrome characterized by severe malabsorptive diarrhea from early life and mild nonketotic diabetes with a variable age of onset [16–18]. Few reports suggest that NEUROG3 may influence pancreatic exocrine function, possibly through its activation of NEUROD1 [20]. Homozygous mutations of RFX6 are associated with a complex clinical phenotype characterized by duodenal atresia, biliary abnormalities, neonatal diabetes mellitus, and malabsorptive diarrhea (Mitchell–Riley Syndrome) [20]. RFX6 is a winged helix transcription factor that is downstream of NEUROG3, and it is required for islet cell development and for enteroendocrine cells function [21•]. Mutation of RFX6 is associated with normal enteroendocrine cells number. The intestinal atresia associated with RFX6 mutations is probably related to a not yet fully characterized role in early gut endoderm. Furthermore, while murine studies suggest that RFX6 is exclusively expressed in enteroendocrine K-cells that express gastric inhibitory polypeptide and others hormones, it remains uncertain if this subset of cells are depleted in humans with RFX6 deficiency [22].	284 285 286 287 288 289 290 291 292 293 294 295 296 297 298 299 300 301 302 303 304 305 306
	<b>Other Defects of Enteroendocrine Cell Differentiation</b>	307
	<b>Main Clinical Fetures and Genotype</b> A complex clinical phenotype of X-linked mental retardation, seizures, lissencephaly, abnormal genitalia, and occasionally congenital diarrhea is due to mutations in the ARX gene (a prd-homeodomain transcription factor) [23, 24]. The ARX gene is a down-stream target of NEUROG3 and is expressed in a subgroup of enteroendocrine cells including those that express CCK, secretin, and glucagon [25]. More than 50% of patients described with loss-of-function ARX mutations present a polyalanine expansion that may be responsible for the highly variable neurologic and intestinal clinical phenotypes associated with this condition [26]. All active hormones produced by endocrine cells are processed by a specific Ca <sup>2+</sup> -dependent serine endoprotease named prohormone convertase 1/3 (PC1/3). Homozygote loss-of-function mutations in PCSK1 gene encoding for PC1/3 have been associated with malabsorptive diarrhea and other endocrinopathies, including adrenal insufficiency, hypothyroidism, and hypogonadism [27]. PCSK1 is also expressed at hypothalamic level, producing various central orexigenic hormones that control appetite. Children presenting mutations of PCSK1 are extremely polyphagic [28].	308 309 310 311 312 313 314 315 316 317 318 319 320 321 322 323 324 325 326 327 328

329 In a cohort of children with this disorder (enteric  
330 dysendocrinosis) was found severe failure to thrive, and PN  
331 was required for the first several years of life [28]. These  
332 children also develop diabetes insipidus and growth hormone  
333 deficiency that distinguish PCSK1 deficiency from other en-  
334 teric endocrinopathies. These findings suggest that enteric  
335 hormones may be particularly important to facilitate nutrient  
336 absorption during infancy when caloric requirement (per body  
337 weight) is at its highest [29].

338 **Defects in Intestinal Immune-Related Homeostasis**

339 **IPEX Syndrome**

Q6 340 **Main Clinical Features** Immunodysregulation,  
341 polyendocrinopathy, enteropathy, X-Linked (IPEX) syndrome  
342 is a monogenic autoimmune disease with early life onset that  
343 is considered the prototype of the defects in intestinal  
344 immune-related homeostasis. This rare syndrome is character-  
345 ized by multiorgan autoimmunity, including severe diarrhea  
346 due to autoimmune enteropathy, chronic dermatitis, and  
347 endocrinopathy (type 1 diabetes mellitus, hypothyroidism).  
348 In more severe cases, symptoms onset starts in the immediate  
349 fetal period with hydrops [30]. In addition to the intestinal  
350 architectural changes due to dysregulation of Treg cells activ-  
351 ity, patients with IPEX syndrome present serum autoanti-  
352 bodies to enterocyte antigens harmonin and villin that are  
353 uniquely found in IPEX and have high diagnostic value [31,  
354 32]. IPEX syndrome is often fatal early in infancy; therefore, a  
355 prompt diagnosis is essential to start treatment as soon as  
356 possible, before tissue damage spreads to multiple organs  
357 [33].

358 **Genotype** IPEX is caused by loss of function mutations in the  
359 gene encoding the forkhead box P3 (FOXP3) on X-  
360 chromosome (Xp11.23) transcription factor for thymic-  
361 derived CD4<sup>+</sup>CD25<sup>+</sup> regulatory T (Treg) cell (Table 1).  
362 These cells play a key role in the establishment and mainte-  
363 nance of immune tolerance [34]. Recently, a novel mutation in  
364 the FOXP3 gene is identified at Phe367 residue level. Phe367  
365 is a key structural residue of the DNA-binding domain of  
366 FOXP because of its contribution to the dimeric state of  
367 FOXP3, and this mutation may have a disruptive effect on  
368 the interaction network whose integrity is essential for  
369 FOXP3 regulatory activity [34]. As a consequence,  
370 FOXP3mut Tregs normally differentiate in the thymus and  
371 can be detected in the peripheral blood and tissues of IPEX  
372 patients, but they are unable to suppress Teff cells. The early  
373 neonatal onset and severity of IPEX enteropathy suggests that  
374 likely the damage is established even during fetal life, inde-  
375 pendently from external environmental factors, such as nutri-  
376 ents and gut microbiota. Whole exome sequencing identified a

novel nonsense mutation in the FOXP3 gene, c.1009C>T, 377  
which was inherited from the mother. FOXP3 is also transiently 378  
expressed by any activated T cells, in which it controls cell 379  
cycle and Th development [35••]. Functional data demonstrate 380  
that Treg cells isolated from IPEX patients are dysfunctional, 381  
as they cannot inhibit proliferation and cytokine production 382  
[33]. Indeed, IPEX patients manifest lymphoproliferation, 383  
skewing towards Th2 and increased frequency of peripheral 384  
IL-17 producing T cells, frequently found in autoimmune dis- 385  
eases. The increase in both Th2 and Th17 cells is involved in 386  
the pathogenesis of the disease and in tissues damage at dif- 387  
ferent target organ levels. In addition to FOXP3 Treg cells, 388  
type 1 regulatory T (Tr1) cells represent nonthymic-derived 389  
Treg cells, able to provide immunoregulation. Tr1 cells main- 390  
tain their function independently of FOXP3. In IPEX patients, 391  
the rapid development of immune disease after birth indicates 392  
that Tr1 cells are not sufficient by themselves to control auto- 393  
immunity [35••]. Lentiviral-mediated overexpression of wild- 394  
type FOXP3 successfully conveys stable regulatory function 395  
to FOXP3mut T cells, opening new therapeutic perspectives 396  
for IPEX patients [36]. 397

**IPEX-Like Syndromes** 398

**Main Clinical Features** This group of CDDs includes an 399  
expanding spectrum of genetic defects that compromise T 400  
regulatory cell function that underlies human disorders of im- 401  
mune dysregulation and autoimmunity. Collectively, these 402  
disorders offer novel insights into pathways of peripheral tol- 403  
erance and their disruption in autoimmunity. However, auto- 404  
immune symptoms phenotypically resembling IPEX often oc- 405  
cur in the absence of detectable FOXP3 mutations; in fact, 406  
patients with clinical manifestations of IPEX have a normal 407  
Foxp3 gene. IPEX-like forms of autoimmune enteropathy 408  
manifestations have been associated with mutations in genes 409  
that are important for Treg maintenance, signaling, and expan- 410  
sion (Table 1) [37]. 411

**Genotype** A number of other gene defects that affect T 412  
regulatory cell function also give rise to IPEX-related 413  
phenotypes, including loss-of-function mutations in 414  
CD25, STAT5b, and ITCH. Recent progress includes the 415  
identification of gain-of-function mutations in STAT1 as a 416  
cause of an IPEX-like disease, emerging FOXP3 geno- 417  
type/phenotype relationships in IPEX [38–41]. The major- 418  
ity of IPEX-like patients, however, still lack a clear diag- 419  
nosis. Mutations in the IL-2receptor alpha subunit (CD25) 420  
are responsible for early-onset enteropathy manifesting 421  
with severe diarrhea. These patients are more susceptible 422  
to early infections, like CMV infection. Probably, CD25 is 423  
essential not only for the Treg cells function but also to 424  
mount an appropriate immune response, concomitant with 425  
the immune-dysregulation. It is described that a patient 426



427 with clinical manifestations of IPEX had a normal Foxp3  
 428 gene, but who had CD25 deficiency due to autosomal  
 429 recessive mutations in this gene. This patient exhibited  
 430 defective IL-10 expression from CD4 lymphocytes,  
 431 whereas a Foxp3-deficient patient expressed normal levels  
 432 of IL-10. These data show that CD25 deficiency results in  
 433 an IPEX-like syndrome and suggests that although Foxp3  
 434 is not required for normal IL-10 expression by human  
 435 CD4 lymphocytes, CD25 expression is important [38].  
 436 Mutations in STAT5b, responsible for transactivation of  
 437 the IL-2 signal from CD25 to FOXP3, have been de-  
 438 scribed associated with reduced Treg cell number [37].  
 439 Children with STAT5b mutation have symptoms other than  
 440 enteropathy that can help in establishing the diagnosis. Early-  
 441 onset chronic/recurrent enteropathy is described in patients  
 442 with either loss- or gain-of-function mutations in STAT1,  
 443 which also impinges effective immunity [39]. Some patients  
 444 with IPEX-like disorder, profound Treg cell deficiency, and a  
 445 normal FOXP3 gene sequence were found to have a homozy-  
 446 gous nonsense mutation in the LPS-responsive beige-like an-  
 447 chor (LRBA) gene, which was previously implicated as a  
 448 cause of autoimmunity. In fact, some patients with LRBA  
 449 deficiency manifest increased levels of autoantibodies against  
 450 autologous antigens in association with a dramatic decreased  
 451 numbers of circulating Treg cells [39]. The clinical features  
 452 observed in patients with LRBA deficiency are heterogeneous  
 453 with the age of presentation ranging from 2 months to  
 454 12 years. The most common features of the patients are chron-  
 455 ic diarrhea, organomegaly, respiratory tract infections, and  
 456 hypogammaglobulinemia [40]. Infections and progressive  
 457 loss of T cells number and function, including Treg cells, is  
 458 reported in patients with loss-of-function mutations, whereas  
 459 Treg instability has been suggested as consequence of the  
 460 gain-of-function variants, which is characterized by chronic  
 461 mucocutaneous candidiasis [37]. Another mutation associated  
 462 with inflammatory unbalance has been described in an extend-  
 463 ed family with recurrence of lymphoproliferation, inflam-  
 464 mation, and dysmorphisms; this concerns ITCH gene, encoding a  
 465 ubiquitin ligase implicated in several T-cell functions [37].  
 466 Whole exome sequencing performed in two affected children  
 467 and their parents, have identified a homozygous missense mu-  
 468 tation in MALT1 gene (mucosa associated lymphoid tissue  
 469 lymphoma translocation 1), which inhibits protein expression  
 470 [42]. NF-κB-dependent lymphocyte activation was resulted  
 471 severely impaired and there was a drastic reduction in  
 472 FOXP3 Treg accounting for the IPEX-like phenotype.  
 473 Following identification of the mutation, both children re-  
 474 ceived hematopoietic stem cell transplantation, which permit-  
 475 ted full clinical recovery. Immunological controls at 6 and  
 476 12 months after transplantation showed normal NF-κB acti-  
 477 vation and correction of Treg frequency [42]. Mutation in IL-  
 478 10 receptor alpha and beta is observed in patients with colitis  
 479 early in life typically associated with skin perianal ulcers and

strong local inflammation. This condition underlines the  
 antiinflammatory action of IL10 and possibly peripheral de-  
 velopment of Tr1 cells [41]. Fistula and abscesses can also be  
 present, with recurrence, requiring multiple surgical interven-  
 tions. Gene mutations in either the alpha or beta subunit of the  
 IL-10 receptor (IL-10R1 and 2) abrogate response to IL-10,  
 and this causes persistent colonic inflammation [41].  
 Currently, the most effective therapy to cure the disease is  
 hematopoietic stem cell transplantation [41]. Although studies  
 of Tr1 cells in these patients have not been directly performed,  
 this disorder illustrates the essential role of IL-10 in control-  
 ling the intestinal homeostasis [43]. This confirms the  
 nonredundancy of both regulatory pathways in the intestine  
 and the importance of considering genetic screening in the  
 presence of early-onset disease [43]. Indeed, hematopoietic  
 stem cell transplantation could be a valid therapeutic option  
 for several of not only these disorders but also novel gene  
 therapy approaches using CRISPR/Cas9 or other technologies  
 could be pursued. However, external factors, like nutrients or  
 intestinal microbiota could influence the immune system, con-  
 tribute to reduce intestinal inflammation, and induce tolerance  
 that provide useful therapeutic insights for the benefit of pa-  
 tients with congenital defects [44].

**Main Therapeutic Strategies for CDDs Deriving from  
 Dysregulation of Intestinal Immune Response** The current  
 treatments available for IPEX and IPEX-like syndrome pa-  
 tients include supportive therapy, immunosuppressive thera-  
 py, and hematopoietic stem cell transplantation (HSCT) [36].  
 Positive long-term outcome for IPEX patients can be obtained  
 with long term immunosuppressive treatment. However, stud-  
 ies demonstrate that immunosuppression does not cure the  
 disease and can induce severe side effects, like osteoporosis,  
 dyslipidemia secondary to corticosteroids, and also chronic  
 renal dysfunction linked to cyclosporine or tacrolimus [45].  
 Early HSTC provides the best outcome, before organs are  
 damaged by autoimmunity [46]. Gene correction of autolo-  
 gous stem cells will hopefully become an option for IPEX  
 patients. Other therapeutic approaches, alternative to multiple  
 immunosuppression, could also be envisaged, aiming to re-  
 establish tolerance in a FOXP3-independent manner. For ex-  
 ample, IPEX patient cells can secrete IL-10 and IL-10-  
 dependent type 1 T regulatory (Tr1) cells, playing important  
 role in peripheral regulation, which can be differentiated de-  
 spite the presence of FOXP3mut. Interestingly, patients with  
 FOXP3 mutations and late onset or unusual clinical presenta-  
 tion are frequently reported. Patients present nephritic-range  
 proteinuria, microscopic hematuria and renal insufficiency.  
 Renal biopsy demonstrates proliferative glomerulonephritis  
 with immune complex deposition [47]. Whether this different  
 phenotype is due to the presence of residual protein function  
 or to the presence in some patients of more efficient FOXP3-

531 independent compensatory mechanisms of tolerance remains  
532 to be clarified.

533 **Considerations on the Diagnostic Approach**  
534 **and Molecular Analysis for CDD Patients**

535 Diarrhea is relatively rare in the first weeks of life, but its early  
536 onset may be predictive of CDDs and requires hospitalization  
537 for an accurate diagnostic work up and therapeutic manage-  
538 ment. Early diagnosis is of paramount importance for the out-  
539 come of most CDDs. The diagnostic approach to CDDs is a  
540 multistep process based on different tools that include anam-  
541 nestic and clinical data, laboratory and instrumental diagnostic  
542 tools, and pathology and molecular analysis. However, con-  
543 sidering the complexity of the CDDs, many cases could have  
544 an atypical presentation that limited the application of a sys-  
545 tematic approach. Positive familiar history of early-onset  
546 chronic diarrhea, polyhydramnios, and/or dilated bowel loops  
547 at ultrasound examination during pregnancy is highly sugges-  
548 tive of CDDs. In the vast majority of cases, the main clinical  
549 manifestation is chronic diarrhea [1••]. In the approach to a  
550 newborn or infant with suspected CDD, it is important to  
551 remember that also at this particular age, infections and food  
552 allergy are frequent causes of chronic diarrhea [48] and that  
553 these conditions together with malformations of gastrointesti-  
554 nal tract should be considered as primary hypothesis. In the  
555 last 10 years, many genes responsible for most CDDs have  
556 been identified (Table 1). The availability of DNA sequencing  
557 techniques has greatly ameliorated the diagnostic approach to  
558 these diseases. Molecular genetics has become helpful to ob-  
559 tain early and unequivocal diagnoses, allowing a rapid and  
560 targeted therapeutic strategies (Table 2) and reducing repeti-  
561 tive invasive and expensive procedures [1••]. Moreover, the  
562 identification of disease-causing mutations in the affected

proband can help to reveal asymptomatic carriers and to offer 563  
counseling and future prenatal diagnosis [49]. 564

Whether the type of mutation(s) can help about the severity 565  
of the clinical phenotype is still under discussion. In specific 566  
CDDs, such as congenital chloride diarrhea, the clinical ex- 567  
pression is poorly related to the genotype and the presence of 568  
modifier genes might contribute to modulate the phenotype. 569  
However, genotype might predict response to therapy. In fact, 570  
it has been demonstrated that specific SLC26A3 mutations 571  
that retain at least some activity of the protein could predict 572  
a more remarkable effect of oral butyrate therapy [50]. More 573  
complex in vitro functional studies are needed to explain the 574  
effect of mutations of uncertain clinical significance, but such 575  
studies are rarely performed in a routine setting. 576

**Conclusions** 577

In recent years, much progress has been made on the 578  
understanding of the pathogenesis of these conditions, 579  
thanks to the development of 3D models derived from 580  
human stem cells, providing a new research perspective 581  
[1••]. The molecular diagnosis has further changed the 582  
scenario of the CDDs, opening the way for new therapeutic 583  
strategies such as the transplantation of hematopoietic 584  
stem cells [1••] and gene therapy endo-nucleases, includ- 585  
ing Talens or CRISPR/Cas9 [1••]. Long-term studies are 586  
necessary to provide other information about the prognos- 587  
is of these conditions. Given the number of the CDDs, 588  
the complexity of the genotype–phenotype relationship 589  
and the need for a multidisciplinary counseling for family 590  
members are essential close collaboration between clinical 591  
and laboratory as part of an international network. Some 592  
examples are the Registry of patients with MVID [1••], 593  
the website Diarrheal Congenital Disorders [1••], and the 594

t2.1 **Table 2** Main therapeutic  
t2.2 strategies for congenital diarrheal  
disorders

t2.1	Defects in absorption and transport of nutrients and electrolytes	<ul style="list-style-type: none"> <li>• Exclusion diet</li> <li>• Substitutive therapy</li> <li>• Parenteral nutrition</li> </ul>
t2.2	Defects in enterocyte structure	<ul style="list-style-type: none"> <li>• Total parenteral nutrition</li> <li>• Antisecretory drugs</li> <li>• Intestinal transplantation</li> </ul>
t2.3	Defects in enteroendocrine cell differentiation	<ul style="list-style-type: none"> <li>• Total parenteral nutrition</li> <li>• Hormone therapy</li> <li>• Intestinal transplantation</li> </ul>
t2.4	Defects in intestinal immune-related homeostasis	<ul style="list-style-type: none"> <li>• Total parenteral nutrition</li> <li>• Hormone therapy</li> <li>• Immunosuppressive, immunomodulator drugs, biologics (corticosteroids, cyclosporine, azathioprine, 6-mercaptopurine, tacrolimus mycophenolate mofetil, sirolimus, infliximab, rituximab)</li> <li>• Bone marrow transplantation</li> </ul>
t2.5		

595 consortium IPEX syndrome [1••], in order to provide  
 596 quick access to analysis and other molecular diagnostic  
 597 procedures for patients suspected of CDDs.

598 **Compliance with Ethical Standards**

599 **Conflict of Interest** Vincenza Pezzella, Giusi Grimaldi, Mariateresa  
 600 Russo, Serena Mazza, Domenica Francesca Mariniello, Lorella Paparo,  
 601 Ausilia Elce, Giuseppe Castaldo, and Roberto Berni Canani each declare  
 602 no potential conflicts of interest.

603 **Human and Animal Rights and Informed Consent** This article does  
 604 not contain any studies with human or animal subjects performed by any  
 605 of the authors.

606 **References**

607 Papers of particular interest, published recently, have been  
 608 highlighted as:

- 609 • Of importance
- 610 •• Of major importance

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