

SHORT COMMUNICATION

IDENTIFICATION AND FUNCTIONAL INFERENCE ON THE MLO FAMILY IN VIRIDIPLANTAE

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SUMMARY

Powdery mildew (PM) is a widespread plant disease of temperate climates caused by ascomycete fungi of the order Erysiphales. PM is an important agricultural issue since it can cause significant economic losses. Specific members of the *MLO* gene family act as susceptibility factors towards the PM disease. A step towards the stability of crop productions would be thus the characterization of *MLO* genes at the genomic level. We carried out a genome-wide characterization of the *MLO* gene family in twenty-three plant and two algal genomes providing manual curated *MLO* protein catalogues. In total, 180 novel proteins containing the *MLO* domain were identified. Evolutionary history and phylogenetic relationships were studied through maximum likelihood analysis. This highlighted eight different clades, including a new monocot-specific clade (VIII) identified for the first time. In addition, 15 and 67 putative PM susceptibility genes, clustering in clade IV and V, respectively, were identified. Results of this work may help to address further biological questions concerning *MLO*s involved in PM susceptibility. In follow-up studies, it could be investigated whether the silencing or loss-of-function mutations in one or more of these candidate genes may lead to PM resistance.

Keywords: *MLO*, Powdery mildew, Functional inference, Gene annotation.

Powdery mildew (PM) is a widespread plant disease of temperate climates caused by ascomycete fungi of the order Erysiphales (Glawe, 2008). It is an important threat for many crops and can cause significant losses in cereal crops such as wheat and barley, vegetable crops such as

tomato and cucumber (Dean *et al.*, 2012), tree species such as those of the family of Rosaceae (Pessina *et al.*, 2014) and ornamental plants such as roses (Linde *et al.*, 2006). Accordingly, the development of varieties that exhibit robust immunity to this disease is of great economic interest. The *Mildew resistance Locus O* (*MLO*) gene family encodes proteins with seven transmembrane domains and a calmodulin-binding site (Büschges *et al.*, 1997; Kim *et al.*, 2002). Specific homologs of the *MLO* gene family act as susceptibility factors towards PM fungi. Indeed, plants carrying recessively inherited loss-of-function mutations of these genes show durable broad-spectrum PM resistance, referred to as *mlo* resistance (Jørgensen *et al.*, 1992; Büschges *et al.*, 1997; Consonni *et al.*, 2006). The *mlo* resistance mechanism involves actin cytoskeleton, proteins involved in cell exocytosis, such as PEN1, PEN2 and PEN3, and possibly Receptor-Like Kinases (RLKs) (Collins *et al.*, 2003; Lipka *et al.*, 2005; Stein *et al.*, 2006; Humphry *et al.*, 2011; Feechan *et al.*, 2013). It was hypothesized that strong resistance of *mlo* mutants is either based on perturbed MAMP responses (Lorek *et al.*, 2013) or on missed defence suppression by PM fungi (Kim *et al.*, 2002). In the latter scenario, wild-type functional *MLO* proteins should be targeted by fungal effector proteins to interfere with the plant immune system. The availability of complete plant genomes allowed the identification of *MLO* family members in several plant species (e.g. Appiano *et al.*, 2015; Devoto *et al.*, 2003; Feechan *et al.*, 2009; Iovieno *et al.*, 2015; Liu *et al.*, 2008; Pessina *et al.*, 2014; Zheng *et al.*, 2016). Moreover, the expression of *MLO* genes has been shown to occur in different plant organs, tissues and cell types, and to be affected by multiple biotic or abiotic stresses (Piffanelli *et al.*, 2002; Chen *et al.*, 2006). In this study, we investigated the *MLO*-family within twenty-three and two genomes of plants and algae, respectively. Furthermore, we conducted a phylogenetic analysis in order to address questions about evolutionary history of *MLO* proteins and to provide a list of candidates for breeding purpose.

Putative *MLO* amino acid sequences of twenty-five different species (*Volvox carterii*, *Chlamydomonas reinhardtii*, *Arabidopsis thaliana*, *Brassica rapa*, *Capsella rubella*, *Citrullus lanatus*, *Cucumis melo*, *Cucumis sativus*,

Table 1. MLO sequences annotated in 25 green plant genomes.

| Family | Specie | Novel MLOs | Previously annotated MLOs | Total MLOs | References of previously annotated MLOs |
|---------------------------|------------------------|------------|---------------------------|------------|---|
| Volvocaceae | <i>V. carteri</i> | 3 | – | 3 | – |
| Chlamydomonadaceae | <i>C. reinhardtii</i> | 4 | – | 4 | – |
| Brassicaceae | <i>A. thaliana</i> | – | 15 (15) ^a | 15 | Consonni <i>et al.</i> , 2006 |
| | <i>B. rapa</i> | 23 | – | 23 | – |
| | <i>C. rubella</i> | 16 | – | 16 | – |
| Cucurbitaceae | <i>C. lanatus</i> | – | 14 (14) | 14 | Iovieno <i>et al.</i> , 2015 |
| | <i>C. melo</i> | – | 16 (16) | 16 | Iovieno <i>et al.</i> , 2015 |
| | <i>C. sativus</i> | – | 13 (13) | 13 | Schouten <i>et al.</i> , 2014 |
| Euphorbiaceae | <i>M. esculenta</i> | 19 | – | 19 | – |
| Fabaceae | <i>G. max</i> | 2 | 39 (39) | 41 | Deshmukh <i>et al.</i> , 2014 |
| | <i>M. truncatula</i> | 16 | – | 16 | – |
| | <i>P. vulgaris</i> | 20 | – | 20 | – |
| Poaceae | <i>B. distachyon</i> | 2 | 11 (11) | 13 | Ablazov and Tombuloglu, 2015 |
| | <i>O. sativa</i> | – | 12 (12) | 12 | Liu <i>et al.</i> , 2007 |
| | <i>S. bicolor</i> | 2 | 13 (13) | 15 | Singh <i>et al.</i> , 2012 |
| | <i>T. aestivum</i> | 47 | 8 (8) | 55 | Elliot, 2005 |
| Rosaceae | <i>F. vesca</i> | – | 17 (17) | 17 | Pessina <i>et al.</i> , 2014 |
| | <i>M. domestica</i> | – | 21 (21) | 21 | Pessina <i>et al.</i> , 2014 |
| | <i>P. persica</i> | – | 19 (19) | 19 | Pessina <i>et al.</i> , 2014 |
| Solanaceae | <i>S. lycopersicum</i> | – | 17 (17) | 17 | Chen <i>et al.</i> , 2014 |
| | <i>N. tabacum</i> | – | 14 (14) | 14 | Appiano <i>et al.</i> , 2015 |
| | <i>C. annuum</i> | 14 | 1 (1) | 15 | Kim <i>et al.</i> , 2012 |
| | <i>S. melongena</i> | 17 | 1 (1) | 18 | Appiano <i>et al.</i> , 2015 |
| | <i>S. tuberosum</i> | – | 12 (12) | 12 | Appiano <i>et al.</i> , 2015 |
| Vitaceae | <i>V. vinifera</i> | 2 | 17 (17) | 19 | Feechan <i>et al.</i> , 2009 |
| Total | | 187 | 260 | 447 | |

^aIn brackets the number of previously annotated MLO genes that were confirmed in this work.

Manihot esculenta, *Glycine max*, *Medicago truncatula*, *Phaseolus vulgaris*, *Brachypodium distachyon*, *Oryza sativa*, *Sorghum bicolor*, *Triticum aestivum*, *Fragaria vesca*, *Malus domestica*, *Prunus persica*, *Solanum lycopersicum*, *Nicotiana tabacum*, *Capsicum annuum*, *Solanum melongena*, *Solanum tuberosum* and *Vitis vinifera* were retrieved from the portal Phytozome v10 (<https://phytozome.jgi.doe.gov/>). A BLASTp (E-value < 1e-6) search in twenty-four proteomes, using the Arabidopsis MLO protein dataset reported by Consonni *et al.* (2006) was performed. The domain composition of proteins was assessed through a domain detection analysis using InterProScan (Zdobnov and Apweiler, 2001), and a total of 447 proteins containing the MLO domain (Pfam ID: PF03094) was identified (Supplementary Table 1, <http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>). Sequences were examined manually to verify completeness and correctness of characteristic protein regions (extracellular N-terminal, transmembrane, intracellular/extracellular loops and intracellular C-terminus), using Geneious v6 (Kearse *et al.*, 2012).

As shown in Table 1, with respect to previous scientific literature our analysis allowed the identification of two novel MLO members in *G. max* (Deshmukh *et al.*, 2014), two in *B. distachyon* (Ablazov and Tombuloglu, 2015), two in *S. bicolor* (Singh *et al.*, 2012), 47 in *T. aestivum* (Konishi *et al.*, 2010) and two in *V. vinifera* (Feechan *et al.*, 2009)

and provided manual curated MLO protein catalogues. Among the twenty-five MLO families used in this study, a vast difference was found in terms of member numbers (ranging from three in *V. carteri* to fifty-five in *T. aestivum*) (Fig. 1). Differently from previous unsuccessful search in the *Chlorophyta* phylum, three and four MLO proteins were identified for the first time in *V. carteri* and *C. reinhardtii*, respectively. This result is plausibly obtained due to an increasing number of available ESTs (Devoto *et al.*, 2003). Normalization of the total number of MLOs identified in each species with respect to the number of genes/haploid genome revealed that the proportion of each MLO family varies considerably (Fig. 1). In detail, the highest k-values were found in *P. vulgaris*, *V. vinifera*, *P. persica* and *G. max* (> 7e-4) whereas the lowest was found in *N. tabacum* genome (1.6e-4). However, the MLO family is poorly represented in algae (k-value *ca.* 2e-4). At the family level, the highest expansion of MLO proteins was observed in Vitaceae with average k-values of 7.2e-4. In contrast, in Solanaceae genome this gene family seems contracted (average k-value *ca.* 4e-4). In the genome of Brassicaceae and Poaceae, the MLO protein family size seems to be expanded since it shows an average k-value of *ca.* 6e-4. Our analysis displayed a slight reduction of the number of MLO genes in function of Rosaceae, Fabaceae and in Poaceae size genomes (average k-value of *ca.* 4e-4).

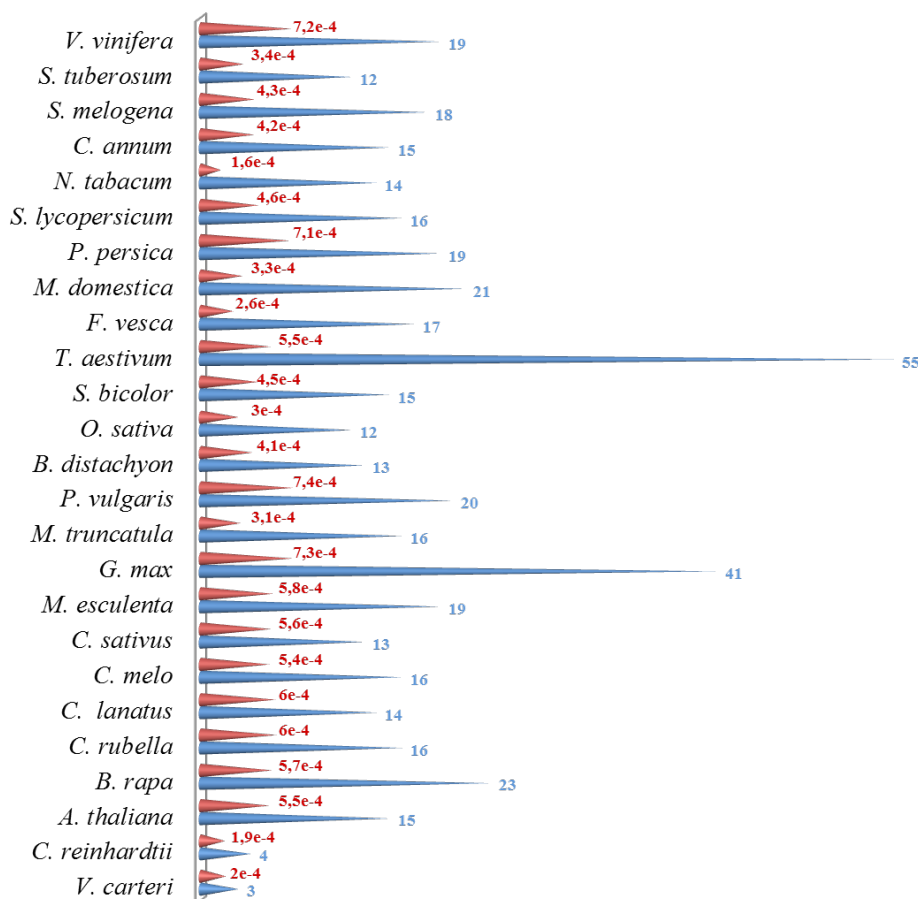


Fig. 1. Composition of twenty-five MLO gene families and corresponding normalized profiles on genome size. The dataset analysed include twenty-three plant (*A. thaliana*, *B. distachyon*, *B. rapa*, *C. lanatus*, *C. melo*, *C. rubella*, *C. sativus*, *F. vesca*, *G. max*, *S. lycopersicum*, *M. esculenta*, *M. domestica*, *M. truncatula*, *N. tabacum*, *O. sativa*, *C. annuum*, *P. persica*, *P. vulgaris*, *S. melongena*, *S. bicolor*, *S. tuberosum*, *T. aestivum*, *V. vinifera*) and two green algae (*V. carteri* and *C. reinhardtii*) genomes. *k*-value: ratio between number of MLO members for each species and the number of genes/haploid genome for each species.

In order to address questions about evolutionary diversification of MLO gene families, predicted protein sequences of twenty-five different genomes were aligned by MUSCLE v3.6 (Edgar *et al.*, 2004). MLO sequences with less than 50% of the full-length MLO domain were not included in the analysis. The corresponding MLO domain, predicted from DB-Pfam (ID: PF03094), of a final dataset of 396 MLO proteins was used for a multi alignment (Fig. 2). Evolutionary analyses were conducted using MEGA6 (Tamura *et al.*, 2011). A consensus phylogenetic tree was obtained using the Maximum Likelihood method and setting the bootstrap value as 100. We considered significant clades those having a bootstrap value support ≥ 69 . These were designated with the Roman numerals from I to VIII (Fig. 2). In order to simplify this complex phylogenetic analysis, a subsequently partitioning of clades in subgroups was performed. This highlighted subgroups indicated as “subclades”, shown in the Supplementary Fig. 1-8 (<http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>). Seven out of the eight clades included homologs in accordance with those characterized by previous phylogenetic studies carried out in monocots

and dicots (Devoto *et al.*, 2003). In addition, we identified a new clade, indicated as clade VIII. The algal MLO homolog group is separated from the rest of sequences (Fig. 2). The presence of this ‘outgroup’ is probably due to a real low similarity with the protein dataset or to the incompleteness of the retrieved amino acid sequences (Fig. 2).

Clade I is characterized by the presence of two Arabidopsis homologs, AtMLO4 (AT1G110000) and AtMLO11 (AT5G53760), involved in root responses to mechanical stimuli (thigmomorphogenesis) (Jaffe, 1973; Chen *et al.*, 2009). In our tree this clade contains 75 members that can be partitioned into three subclades (Ia, Ib, Ic) (Supplementary Fig. 1, <http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>). Thus, homologs of this clade could be further investigated for their role in thigmomorphogenesis. Other roles of clade I homologs are also possible since AtMLO14 (AT1G26700), not involved in root thigmomorphogenesis, is also included in clade I. Interestingly, the subclade Ib differs from the others for not containing members of the Brassicaceae family. Evolutionarily, clade I is

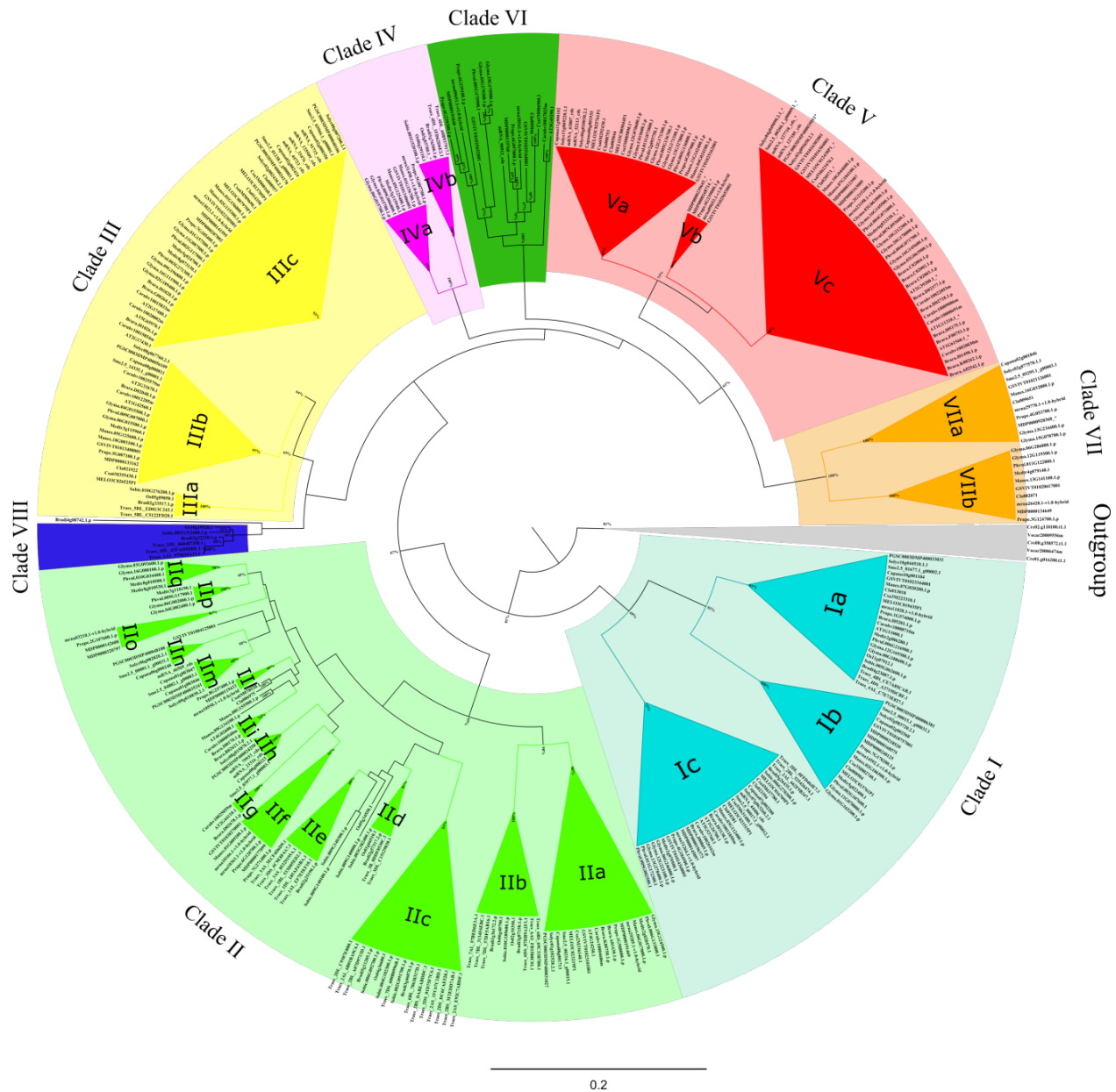


Fig. 2. Maximum likelihood analysis of twenty-five Viridiplantae MLO-families. Bootstrap values > 50% are indicated above branches. The tree is drawn to scale, with branch lengths proportional to the number of substitutions per site. Identified clades are indicated by Roman numbers and underlined by coloured boxes (clades I-VIII). To facilitate the tree description, the clades were split in “subclades” (subgroups described in more detail).

reported by Jiwan *et al.* (2013) to be the most ancient clade since they hypothesized that this clade was present before the separation of vascular and nonvascular plants.

Clade II, exhibiting 113 members, is the most numerous MLO proteins group characterized in this study. It is divided in 15 subclades (IIa-IIq), most of which seems to be class or even family-specific (Supplementary Fig. 2, <http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>). In more detail, the subclades IIb, IIc, IId, IIe (with 44 members in total) are exclusive for monocots; the subclades IIg and IIi include 7 members of the Brassicaceae family; IIh, IIm and IIn include 16 Solanaceae-specific members; III and IIo contain 7 homologs

specific of Rosaceae; IIp and IIq group 9 homologs of the Fabaceae family.

Clade III includes the *Arabidopsis* homolog AtMLO7 (AT2G17430), previously shown to be involved in pollen tube reception from the synergid cells during fertilization (Kessler *et al.*, 2010). In this study, it grouped 69 members divided in three subclades (Supplementary Fig. 3, <http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>). Interestingly, the subclade IIIa is specific for monocots.

Clade IV is primarily but not exclusively represented by monocot MLO proteins (Acevedo-Garcia *et al.*, 2014; Várallyay *et al.*, 2012) and contains all the

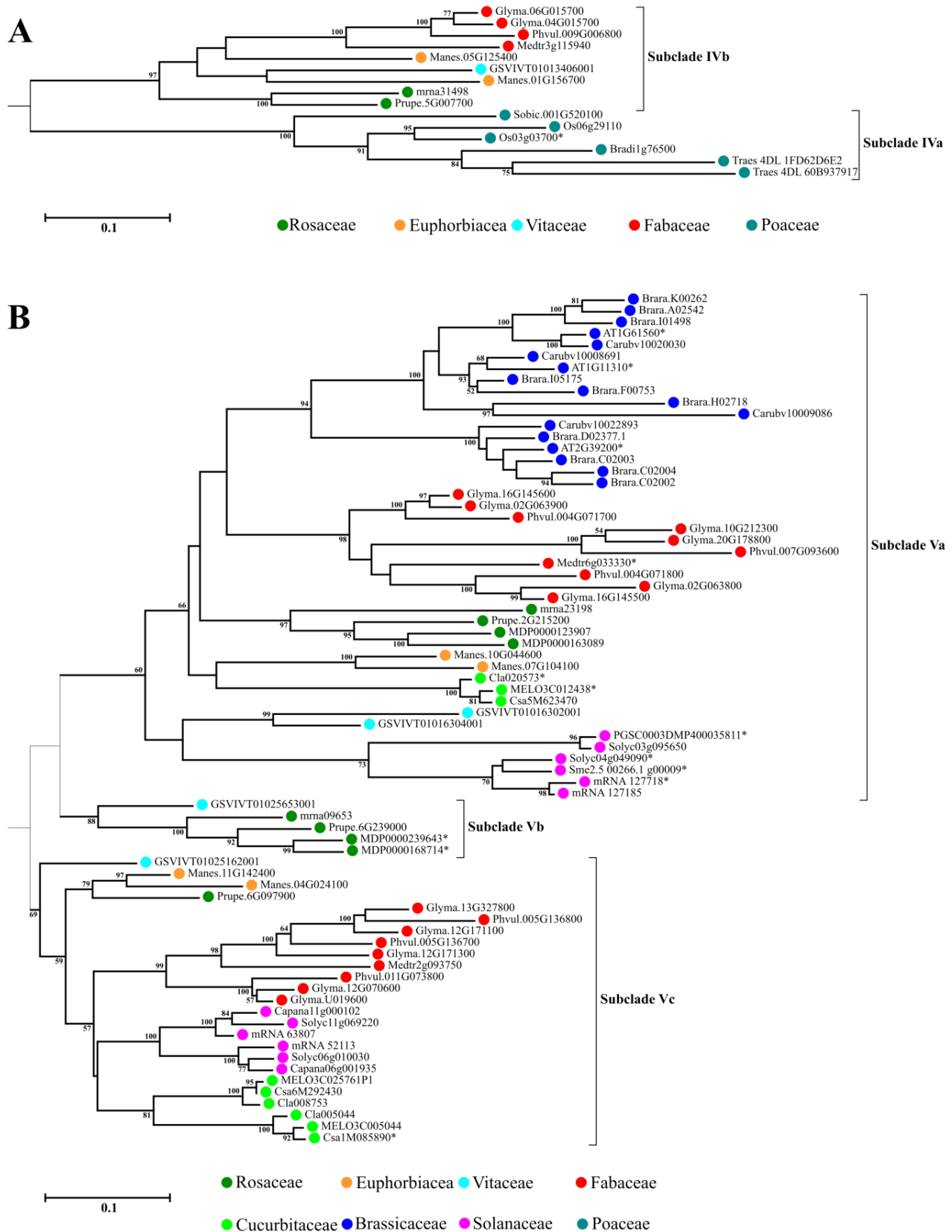


Fig. 3. Close-up view on IV and V phylogenetic clades. Botanic families corresponding to each homolog are indicated by coloured spots. Gene-IDs of MLO-homologs functionally involved in powdery mildew susceptibility are marked with an asterisk (*). A) Clade IV known to be the clade of monocot MLO homologs acting as susceptibility factors for Powdery Mildew. Subgroups IVa are exclusively composed by monocots members and IVb by dicots ones. B) Clade V including some of the homologues functionally characterized for their involvement in PM interaction.

monocot homologs involved in the interaction with PM fungi (namely barley HvMLO, rice OsMLO3 and wheat TaMLO_A1 and TaMLO_B1) (Fig. 3A). For the first time, we divided the clade IV in the two subclades IVa and IVb, which group monocot and dicot members, respectively. In further detail, subclade IVa is composed by 6 members, whereas subclade IVb contains 9 genes of the Eurosids I Cladus (Fig. 3A). Many eudicot species seem to have lost the clade IV, as we found that 13 out of 19 of the dicot species considered in this study do not show any MLO homolog in clade IV. Probably, this MLO group plays a minor role in these species.

All the *MLO* susceptibility genes functionally characterized in dicots, namely Arabidopsis AtMLO2, AtMLO6, AtMLO12 (Consonni *et al.*, 2006), tomato SIMLO1 (Bai *et al.*, 2008), grapevine VvMLO3 and VvMLO4 (Feechan *et al.*, 2013), tobacco NtMLO1 (Appiano *et al.*, 2015), pepper CaMLO2 (Zheng *et al.*, 2013) and barrel clover MtMLO1 (Humphry *et al.*, 2011) group in clade V. In the present study, 74 putative homologs clustered in clade V and were further separated into three subclades, named Va, Vb and Vc (Fig. 3B). The Va subclade presents 6 homologs belonging to Solanaceae, 2 to Vitaceae, 3 to Cucurbitaceae, 2 to Euphorbiaceae, 4 to Rosaceae, 10 for Fabaceae and 17 for Brassicaceae species, the latter possibly as the result of a recent duplication event. The subclade Vb, except for a *Vitis vinifera* homolog, seems to be typical of the Rosaceae family. The subclade Vc includes 6 members belonging to Solanaceae, 6 to Cucurbitaceae, 9 to Fabaceae, 5 to Rosaceae, 2 to Euphorbiaceae and 1 to Vitaceae. We identified 19 and 5 putative novel PM susceptibility factors in subclades Va and Vc, respectively. In more detail, the subclade Va presents 10, 4, 2 and 3 novel *MLO* candidate genes identified in *B. rapa*, *C. rubella*, *M. esculenta* and *P. vulgaris* genomes, respectively (Fig. 3B). The subclade Vc presents 2 and 3 PM susceptibility candidates in *M. esculenta* and *P. vulgaris*, respectively. This phylogenetic analysis confirmed that clade V, emerging after the divergence of monocots from dicots (Jiwan *et al.*, 2013), likely replaced the function of clade IV. Indeed, Appiano *et al.* (2015) demonstrated that, despite the phylogenetic distance of monocot and dicot MLO proteins involved in the interaction with PM fungi are functionally conserved.

As shown in supplementary Fig. 4 (<http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>), clade VI is composed of 17 dicot members and does not contain homologs of the Euphorbiaceae family. It presents 3 homologs of Fabaceae, 7 of Rosaceae, 2 of Cucurbitaceae, 2 of Vitaceae and 2 of Brassicaceae.

Our study expanded the clade VII to 21 dicot members, including the tomato SIMLO2 (Solyc02g77570) as previously proposed (Acevedo-Garcia *et al.*, 2014) (Supplementary Fig. 5, <http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>). The subclade VIIa includes 4 homologs of Fabaceae, 1 of Euphorbiaceae, 1 of Vitaceae, 1 of Cucurbitaceae and 3 of Rosaceae. The

subclade VIIb includes 11 MLO proteins from the same species of the subclade VIIa and Solanaceae species. Differently from the findings of Acevedo-Garcia *et al.* (2014), there are no members of *C. sativus* in this clade since we used full-length sequence genes (Schouten *et al.*, 2014) from a new genome version.

Moreover, three MLO genes (VvMLO6, CIMLO1 and CIMLO8), previously reported in the Clade V (Iovieno *et al.*, 2015; Feechan *et al.*, 2008), were grouped into the phylogenetic clade VII. Interestingly, in this study we identified a clade (VIII) including six monocot-specific members (Supplementary Fig. 6, <http://sipav.org/main/jpp/index.php/jpp/article/downloadSuppFile/3699/5>).

In conclusion, in this study we present the identification of 183 novel *MLO* genes in several plant genomes and confirm 264 *MLO* homologs previously reported. In addition, we identified *MLO* genes in two algal species for the first time. We report a detailed investigation on phylogenetic reconstruction of the *MLO* gene family in twenty-five genomes. Furthermore, we refine previous phylogenetic work reporting two new clades in which no Arabidopsis members are present. We confirm that *MLO* homologues functionally characterized for their involvement in plant-PM interactions belong to clade IV and clade V, and we identified new putative PM susceptibility factors. The information here retrieved significantly increases our understanding of *MLO* gene family organization and diversification and is a basis for further functional investigations on the role of *MLO* genes. The functional characterization of the putative PM susceptibility factors and the identification or generation of *mlo* mutant (using genome editing technologies, for instance) could be used as a breeding strategy to introduce PM resistance across cultivated species.

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REFERENCES

- Ablazov A., Tombuloglu H., 2015. Genome-wide identification of the *Mildew resistance Locus O* (*MLO*) gene family in novel cereal model species *Brachypodium distachyon*. *European Journal of Plant Pathology* **145**: 239-253.
- Acevedo-Garcia J., Kusch S., Panstruga R., 2014. Magical mystery tour: MLO proteins in plant immunity and beyond. *The New Phytologist* **204**: 273-81.

- Appiano M., Pavan S., Catalano D., Zheng Z., Bracuto V., Lotti C., Visser R.G.F., Ricciardi L., Bai Y., 2015. Identification of candidate *MLO* powdery mildew susceptibility genes in cultivated Solanaceae and functional characterization of tobacco *NtMLO1*. *Transgenic Research* **24**: 847-858.
- Bai Y., Pavan S., Zheng Z., Zappel N.F., Reinstädler A., Lotti C., De Giovanni C., Ricciardi L., Lindhout P., Visser R., Theres K., Panstruga R., 2008. Naturally occurring broad-spectrum powdery mildew resistance in a central american tomato accession is caused by loss of *mlo* function. *Molecular Plant-Microbe Interactions* **21**: 30-39.
- Büsches R., Hollricher K., Panstruga R., Simons G., Wolter M., Frijters A., Daelen R., Lee T., Diergaard P., Groenendijk J., Vos P., Salamini F., Schulze-Lefert P., 1997. The Barley *Mlo* gene: a novel control element of plant pathogen resistance. *Cell* **88**: 695-705.
- Chen Z., Hartmann H., Wu M., Friedman E., Chen J., Pulley M., Schulze-Lefert P., Panstruga R., Jones A., 2006. Expression analysis of the *AtMLO* gene family encoding plant-specific seven-transmembrane domain proteins. *Plant Molecular Biology* **60**: 583-597.
- Chen Z., Noir S., Kwaaitaal M., Hartmann H.A., Wu M.J., Mudgil Y., Sukumar P., Muday G., Panstruga R., Jones A.M., 2009. Two Seven-Transmembrane Domain MILDEW RESISTANCE LOCUS O Proteins Cofunction in Arabidopsis Root Thigmomorphogenesis. *The Plant Cell* **21**: 1972-1991.
- Chen Y., Wang Y., Zhang H., 2014. Genome-wide analysis of the mildew resistance locus o (*MLO*) gene family in tomato (*Solanum lycopersicum* L.). *Plant Omics Journal* **7**: 87-93.
- Collins N.C., Thordal-Christensen H., Lipka V., 2003. SNARE-protein-mediated disease resistance at the plant cell wall. *Letters to Nature* **312**: 310-312.
- Consonni C., Humphry M.E., Hartmann H.A., Livaja M., Westphal L., Vogel J., Lipka V., Kemmerling B., Schulze-Lefert P., Somerville S.C., Panstruga R., 2006. Conserved requirement for a plant host cell protein in powdery mildew pathogenesis. *Nature Genetics* **38**: 716-720.
- Dean R., Van Kan J., Pretorius Z., Kosack K., Di Pietro A., Spanu P., Rudd J., Dickman M., Kahmann R., Ellis J., Foster G., 2012. The Top 10 fungal pathogens in molecular plant pathology. *Molecular Plant Pathology* **13**: 414-430.
- Deshmukh R., Singh V.K., Singh B.D., 2014. Comparative phylogenetic analysis of genome-wide *MLO* gene family members from *Glycine max* and *Arabidopsis thaliana*. *Molecular Genetics and Genomics* **289**: 345-359.
- Devoto A., Hartmann H.A., Piffanelli P., Elliott C., Simmons C., Taramino G., Goh C., Cohen F.E., Emerson B.C., Schulze-Lefert P., Panstruga R., 2003. Molecular phylogeny and evolution of the plant-specific seven-transmembrane *MLO* family. *Journal Molecular Evolution* **56**: 77-88.
- Edgar R.C., Drive R.M., Valley M., 2004. MUSCLE : multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* **32**: 1792-1797.
- Elliott C., Uller J.M., Miklis M., Bhat R.A., Schulze-Lefert P., Panstruga R., 2005. Conserved extracellular cysteine residues and cytoplasmic loop-loop interplay are required for functionality of the heptahelical *MLO* protein. *Biochemical Journal* **254**: 243-254.
- Feechan A., Jermakow A.M., Dry I.B., 2009. Grapevine *MLO* candidates required for powdery mildew pathogenicity? *Plant Signaling & Behavior* **4**: 522-523.
- Feechan A., Jermakow A.M., Ivancevic A., Godfrey D., Pak H., Panstruga R., Dry I.B., 2013. Host cell entry of powdery mildew is correlated with endosomal transport of antagonistically acting VvPEN1 and VvMLO to the papilla. *Molecular Plant-Microbe Interactions* **26**: 1138-1150.
- Glawe D.A., 2008. The powdery mildews : a review of the world's most familiar (yet poorly known) plant pathogens. *Annual Reviews of Phytopathology* **46**: 27-51.
- Humphry M., Reinstädler A., Ivanov S., Bisseling T., Pastruga R., 2011. Durable broad-spectrum powdery mildew resistance in pea *erl* plants is conferred by natural loss-of-function mutations in *PsMLO1*. *Molecular Plant Pathology* **12**: 866-878.
- Iovieno P., Andolfo G., Schiavulli A., Catalano D., Ricciardi L., Frusciant L., Ercolano M.R., Pavan S., 2015. Structure, evolution and functional inference on the *Mildew Locus O* (*MLO*) gene family in three cultivated Cucurbitaceae. *BMC Genomics* **16**: 1-13.
- Jaffe M.J., Forbes S., 1993. Thigmomorphogenesis : the effect of mechanical perturbation on plants. *Plant Growth Regulation* **12**: 313-324.
- Jiwan D., Roalson E.H., Main D., 2013. Antisense expression of peach mildew resistance locus O (*PpMlo1*) gene confers cross-species resistance to powdery mildew in *Fragaria x ananassa*. *Transgenic Research* **12**: 1119-1131.
- Jørgensen J.H., 1992. Discovery, characterization and exploitation of *Mlo* powdery mildew resistance in barley. *Euphytica* **66**: 141-152.
- Kearse M., Moir R., Wilson A., Stones-Havas S., Cheung M., Sturrock S., Buxton S., Cooper A., Markowitz S., Duran C., Thierer T., Ashton B., Mentjies P., Drummond, A., 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* **28**: 1647-1649.
- Kessler S.A., Shimosato-Asano H., Keinath N.F., Wuest S.E., Ingram G., Panstruga R., Grossniklaus U., 2010. Conserved molecular components for pollen tube reception and fungal invasion. *Science* **330**: 968-971.
- Kim M.C., Panstruga R., Elliott C., Mu J., Devoto A., Yoon H.W., Park H.C., Cho M.J., 2002. Calmodulin interacts with *MLO* protein to regulate defence against mildew in barley. *Letters to Nature* **416**: 447-450.
- Konishi S., Sasakuma T., Sasanuma T., 2010. Identification of novel *Mlo* family members in wheat and their genetic characterization. *Genes & Genetic Systems* **85**: 167-175.
- Linde M., Hattendorf A., Kaufmann H., Debener T., 2006. Powdery mildew resistance in roses: QTL mapping in different environments using selective genotyping. *Theoretical and Applied Genetics* **113**: 1081-1092.
- Lipka V., Dittgen J., Bednarek P., Bhat R., Wiermer M., Stein M., Brandt W., Rosahl S., Scheel D., Llorente F., Molina A., Parker J., Somerville S., Schulze-Lefert P., 2005. Pre- and postinvasion defenses both contribute to nonhost resistance in Arabidopsis. *Science* **310**: 1180-1183.
- Liu Q., Zhu H., 2008. Molecular evolution of the *MLO* gene

- family in *Oryza sativa* and their functional divergence. *Gene* **409**: 1-10.
- Lorek J., Griebel T., Jones AM., Kuhn H., Panstruga R., 2013. The Role of Arabidopsis heterotrimeric G-protein subunits in MLO2 function and MAMP-triggered immunity. *Molecular Plant-Microbe Interactions Journal* **26**: 991-1003.
- Pessina S., Pavan S., Catalano D., Gallotta A., Visser R.G., Bai Y., Malnoy M., Schouten H.J., 2014. Characterization of the MLO gene family in Rosaceae and gene expression analysis in *Malus domestica*. *BMC genomics* **15**: 618.
- Piffanelli P., Zhou F., Casais C., Orme J., Jarosch B., Schaffrath U., Collins N.C., Panstruga R., Schulze-Lefert P., 2002. The barley MLO modulator of defense and cell death is responsive to biotic and abiotic stress stimuli. *Plant Physiology* **129**:1076-1085.
- Singh V.K., Singh K., Chand R., Singh B.D., 2012. Genome wide analysis of disease resistance MLO gene family in Sorghum [*Sorghum bicolor* (L) Moench]. *International Journal of Plant Pathology* **2**: 18-27.
- Schouten H., Krauskopf J., Visser R., Bai Y., 2014. Identification of candidate genes required for susceptibility to powdery or downy mildew in cucumber. *Euphytica* **200**: 475-486.
- Stein M., Dittgen J., Sánchez-Rodríguez C., Hou B.H., Molina A., Schulze-Lefert P., Lipka V., Somerville S., 2006. Arabidopsis PEN3/ PDR8, an ATP binding cassette transporter, contributes to nonhost resistance to inappropriate pathogens that enter by direct penetration. *Plant Cell* **18**: 731-746.
- Tamura K., Peterson D., Peterson N., Stecher G., Nei M., Kumar S., 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution* **28**: 2731-2739.
- Várallyay E., Gabor G., Burgyan J., 2012. Virus-induced gene silencing of Mlo genes induces powdery mildew resistance in *Triticum aestivum*. *Archives of Virology* **7**: 1345-1350.
- Zdobnov E.M., Apweiler R., 2001. InterProScan - an integration platform for the signature-recognition methods in InterPro. *Bioinformatics* **17**: 847-848.
- Zheng Z., Nonomura T., Appiano M., Pavan S., Matsuda Y., Toyoda H., Wolters A., Visser R.G.F., Bai Y., 2013. Loss of function in Mlo orthologs reduces susceptibility of pepper and tomato to powdery mildew disease caused by *Leveillula taurica*. *PloS one* **8**: e70723.
- Zheng Z., Appiano M., Pavan S., Bracuto V., Ricciardi L., Visser R.G.F., Wolters A.A., Bai Y. 2016. Genome-wide study of the tomato SIMLO gene family and its functional characterization in response to the powdery mildew fungus *Oidium neolycopersici*. *Frontiers in Plant Science* **7**: 380.

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