## Common Strategies and Technologies for the Ecosafety Assessment and Design of Nanomaterials Entering the Marine Environment

Ilaria Corsi,<sup>\*,†</sup> Gary N. Cherr,<sup>‡</sup> Hunter S. Lenihan,<sup>§</sup> Jerome Labille,<sup>⊥</sup> Martin Hassellov,<sup>∥</sup> Laura Canesi,<sup>#</sup> Francesco Dondero,<sup>¬</sup> Giada Frenzilli,<sup>◆</sup> Danail Hristozov,<sup>×</sup> Victor Puntes,<sup>◇</sup> Camilla Della Torre,<sup>†</sup> Annalisa Pinsino,<sup>¶</sup> Giovanni Libralato,<sup>×,+</sup> Antonio Marcomini,<sup>×</sup> Enrico Sabbioni,<sup>+</sup> and Valeria Matranga<sup>\*,¶</sup>

<sup>†</sup>Department of Physical, Earth and Environmental Sciences, University of Siena, Siena 53100, Italy, <sup>‡</sup>Bodega Marine Laboratory and Department of Environmental Toxicology and Nutrition, University of California, Davis, Bodega Bay, California 94923, United States, <sup>§</sup>Bren School of the Environment, University of California, Santa Barbara, California 93106, United States, <sup>⊥</sup>Aix-Marseille Université, CNRS, IRD, CEREGE UM34, Aix en Provence cedex 04, France, <sup>II</sup> Department of Chemistry and Molecular Biology, University of Gothenburg, Gothenburg S-405 30, Sweden, <sup>#</sup>Department of Earth, Environment and Life Sciences, University of Genova, Genova 16142, Italy, <sup>¬</sup>Department of Earth, Environment and Life Sciences, University of Piemonte Orientale, x Alessandria 15100, Italy, <sup>♠</sup>Department of Clinical and Experimental Medicine, University of Pisa, Pisa 56126, Italy, <sup>×</sup>Department of Environmental Sciences, Informatics and Statistics, University Cà Foscari Venice, Venice 30123, Italy, <sup>◇</sup>Catalan Institute of Nanoscience and Nanotechnology (ICN2), Barcelona 08193, Spain, <sup>+</sup>ECSIN - European Center for Sustainable Impact of Nanotechnology, Veneto Nanotech S.C.p.A., Rovigo 45100, Italy, and <sup>¶</sup>CNR — Institute of Biomedicine and Molecular Immunology "Alberto Monroy", Palermo 90146, Italy

**ABSTRACT** The widespread use of engineered nanomaterials (ENMs) in a variety of technologies and consumer products inevitably causes their release into aquatic environments and final deposition into the oceans. In addition, a growing number of ENM products are being developed specifically for marine applications, such as antifouling coatings and environmental remediation systems, thus increasing the need to address any potential risks for marine organisms and ecosystems. To safeguard the marine environment, major scientific gaps related to assessing and designing ecosafe ENMs need to be filled. In this Nano Focus, we examine key issues related to the state-of-the-art models and analytical tools being developed to understand ecological risks and to design safeguards for marine organisms.

ngineered nanomaterials (ENMs) are used extensively in a variety of emerging technologies and commercial products, including biomedicine, pharmaceuticals and personal care, renewable energies, and electronic devices.<sup>1,2</sup> As these materials are used, disposed of, and degraded, they can release ENMs into the environment. Fate and transport models indicate that ENMs entering soil and waterways will eventually reach the marine environment as nanowaste that can cause human injuries as well as ecological impact with significant socioeconomic conseguences.<sup>3,4</sup> In addition to accidental releases and exposures, a suite of new marine nanotechnologies, including antifouling paints and pollution remediation systems, are also being developed with great uncertainty about their ecosafety and sustainability for the marine environment. An increasing number of short-term, well-controlled laboratory studies have tested ENM's toxicity on marine organisms and showed a wide variety of potential biological injuries.<sup>5,6</sup> Whether ENMs cause similar injuries in the dynamic natural marine environment is uncertain because the fate, transport, and behavior of many ENMs in seawater, and thus their biological risks, remain poorly understood.

The chemistry of ENMs plays a crucial role as their bioavailability, bioaccumulation, and toxicity are difficult to predict in seawater because the materials undergo complex interactions/transformations when exposed to elevated ionic concentrations. For example, metallic ENMs undergo aggregation, sedimentation, corrosion, and reprecipitation in seawater, but the rates at which these processes occur depend heavily on the inherent dissolution rate of the dominant metal. Therefore, developing tools to predict, to estimate, and to compare the long-term effects and risks of ENMs presents many challenges. Meeting these challenges is a fundamental objective of many marine

\* Address correspondence to ilaria.corsi@unisi.it, valeria.matranga@ibim.cnr.it.

Published online September 29, 2014 10.1021/nn504684k

© 2014 American Chemical Society

VOL.8 • NO.10 • 9694-9709 • 2014



nanoecosafety focused research programs, as there is little doubt that organisms living in marine waters and sediments will be exposed to ENMs in effluents from factories and households or from urban runoff. In this Nano Focus, we examine the state-of-the-art approaches used to predict and to measure the exposure and toxicity of ENMs in marine ecosystems. We focus on the influence of key environmental conditions, such as salinity, chronic low-level exposures that probably will dominate natural coastal seascapes, the effects of EMNs in concert with other multiple environmental stressors, and the varying effects of ENMs as they age under natural environmental regimes. Our goal is to provide a synopsis of available methods, models, and insights necessary to highlight the gaps that exist and need to be filled for the design and production of ecosafe ENMs to be used for marine ecosystem sustainability.

In this Nano Focus, we examine the state-ofthe-art approaches used to predict and to measure the exposure and toxicity of ENMs in marine ecosystems

Characterization and Behavior in Seawater: From Models to Analytical Tools. Predicting ENM exposure will lead to a better understanding of their fate and behavior in the marine ecosystem. Their dispersion stability is a key factor that determines their residence time in the water column, and thus their occurrence in the benthic or pelagic systems. Dispersion is influenced by a variety of parameters, including the intrinsic characteristics of the ENMs, the solution chemistry, and the interaction with surrounding components. $^{7-12}$ Ranging from the basic theoretical

prediction of the colloidal stability to the most relevant and holistic approach accounting for the system heterogeneity, a number of physicochemical mechanisms appear determinant in the fate of ENMs in the seawater column and sediments. In the seawater column, both hydrodynamic flow (river outlet, wave oscillations) and random diffusion (Brownian) favor the colloidal transport of particulate matter, competing with gravitational sedimentation (see Figure 1). The particle size and density thus determine the persistence in suspension. The size is that of the individual isolated nanoparticles (NP) or their homogeneous and heterogeneous aggregates (nanoobjects and their aggregates and agglomerates). The aggregation dynamics consist of a two-step process where the suspended particles first collide, and then may attach to each other.<sup>13</sup> The collision frequency and the attachment efficiency, respectively, drive these two steps, together determining the aggregation rate. The attachment efficiency results from the balance between repulsive and attractive interparticle forces, as predicted by DLVO theory and its numerous extensions.14-17 These interactions have been widely studied for synthetic systems with basic and homogeneous compositions, revealing how surface charge, which strongly depends on pH and salinity, plays a determining role in the electrostatic dispersion stability of particles, especially in those that have no steric protection against aggregation.<sup>18,19</sup> Critical salt concentrations have been determined for the counterion nature and valence and solution pH by measuring the induced kinetics of aggregation.<sup>12,20–23</sup> Bare NPs often display low dispersion stability in natural water, while functionalized NPs (e.g., using polyethylene glycol or a polyvinylpyrrolidone coating) may remain very stable at NaCl concentrations higher than that of seawater. Nevertheless, the marine environment is rarely homogeneous and of basic composition. Natural mineral

(clay, carbonates, etc.) and organic matter (algaes, exopolymers, etc.) reside suspended in the seawater column as geogenic and biogenic colloids,<sup>24</sup> potentially interacting with the ENMs. The affinity between the NPs and these naturally occurring colloids, a consequence of the NPs' high surface energy, drives the socalled heteroaggregation phenomenon. This has been studied under certain solution chemistries.<sup>10,21-23,25</sup> As an example, metal oxide and silver ENMs have been shown to undergo high aggregation and sedimentation rates in seawater of high ionic strength (IS) and low in natural organic matter (NOM) content while they remained more stable in freshwater (low IS, high NOM).26,27 Similarly, metal<sup>28</sup> and oxide ENMs<sup>29</sup> aggregating at high salt concentrations are actually stabilized by interactions with proteins. Moreover, the kinetics of ENMs removal appeared also to correlate with their concentrations.

In natural water, both homo- and heteroaggregation may occur simultaneously, their respective kinetics resulting directly from the corresponding collision frequencies and attachment efficiencies.<sup>25</sup> In the seawater column, the natural colloids mainly consist of dissolved and particulate organic carbon, among which large and amorphous organic aggregates displaying low settling velocities are distinct from denser particles with higher settling velocities, such as diatoms.<sup>30,31</sup> Their concentrations fall in the microgram per liter  $(\mu q/L)$  range. However, the release of ENMs in seawater is of terrestrial origin, implying their prior transport through surface water wherein suspended particulate matter is 3 orders of magnitude more abundant (mg/L), while the predicted ENMs concentrations range below microgram per liter ( $\mu$ g/L). The most widely used ENMs, based on TiO<sub>2</sub> and Ag NPs, are expected at 10<sup>-2</sup>-10 and 10<sup>-5</sup>-1 mg/L, respectively.<sup>32</sup> These concentrations are likely to favor the heteroaggregation and colloidal transport of persistent ENMs to the sea. Estuaries

VOL. 8 • NO. 10 • 9694-9709 • 2014







and coastal marine environments are thus areas of high interest, where stable colloids brought by river encounter increasing salinity vgradients that may cause their large agglomeration and sedimentation. This situation typically forms siltation areas, where ENMs and other pollutants bound to those colloids may bury and accumulate within sediments or persist in the water column.<sup>31,33–35</sup> The NOM composition also plays a role in this balance. Seasonal variations influence phytoplankton and microbial activity that produces exudates made of fibrillar and acidic polysaccharide compounds. These biopolymers can remain dispersed or assemble together vas gel networks.<sup>36–40</sup> Their structure and abundance determine their potential roles in the fate of the suspended particulate matter, and thus of ENMs. Electrostatic or steric stabilization is often observed when the particle surface is saturated with adsorbed molecules.41,42 On the contrary, lower surface coverage added to large molecular weight may favor bridging flocculation<sup>43–45</sup> and formation of large aggregates such as marine snow. Moreover, because of a given NP's

size ratio, the resulting heteroaggregates may sometimes consist of biopolymers decorated with adsorbed NPs, as pearls on a necklace, or in other cases, of biogel incorporating the NP in their network.46,47 The unique term heteroaggregation accounts for a very wide panel of colloidal dynamics, ranging from flocculation to colloidal stabilization. Very contrasting final states may be reached, depending on the respective size and concentration of the natural colloids with regard to the ENM.<sup>45,48–50</sup> It is the final size and density of the formed heteroaggregates that determine the persistence of ENMs in the seawater column. Those remaining in the colloidal size range, typically below 1  $\mu$ m in size,<sup>24</sup> promote further transport of ENMs and their exposure to pelagic organisms, while the formation and sedimentation of larger flocs instead concerns the benthic ecosystem.

The State of the Art on Marine ENMs Ecotoxicology. Many harmful effects of ENMs have been reported for marine organisms, but factors including physicochemical properties of ENMs, seawater parameters, interactions with both physical and chemical factors (*i.e.*, ultraviolet and marine pollutants), and organisms' physiology and ecology make ecotoxicological assessment difficult.<sup>4</sup>

Selected invertebrate organisms have been recognized as potential biological targets of ENMs exposure, including biofilm, phytoplankton, bivalves, and bottom grazers (reviewed in ref 6 for inorganic NPs). Little data are available on higher trophic levels such as fish and marine mammals.<sup>51–54</sup> Biological responses and end points of toxicity have been investigated as (1) mechanisms of uptake and of translocation inside the body, (2) organelles/ compartments/cells/tissues as targets of toxicity and/or retention, and (3) cellular pathways/mechanisms of toxicity.<sup>6</sup> With few exceptions, there is a lack of epidemiological studies focusing on reproduction and development (e.g., early life stages, embryos, and larvae).55-58

Increasing evidence supports the hypothesis that the immune system of marine bivalves represents a significant target of ENMs. The blue mussel *Mytilus* has been the species most utilized so far for marine ecotoxicological studies on the effects and mechanisms of action of ENMs on innate immunity.<sup>59,60</sup> *In vitro* studies showed that different NPs

VOL.8 • NO.10 • 9694-9709 • 2014



Many harmful effects of ENMs have been reported for marine organisms, but factors including physicochemical properties of ENMs, seawater parameters, interactions with both physical and chemical factors, and organisms' physiology and ecology make ecotoxicological assessment difficult.

are rapidly taken up by mussel hemocytes, affecting a large number of functional parameters, from lysosomal function to phagocytic activity and oxyradical production, and also inducing pro-apoptotic processes; the effects of NPs were mediated by stress-activated mitogen-activated protein kinase signaling, as in mammalian phagocytes.61-63 In vivo exposure to different NPs and, in particular, to nano-TiO<sub>2</sub> chosen as a model NP type<sup>64,65</sup> enabled formulation of a hypothesis on the possible pathways leading to nanoinduced immunomodulation<sup>60</sup> (see Figure 2). Due to the physiological mechanisms involved in the feeding process, nano-TiO<sub>2</sub> agglomerates/ aggregates formed in seawater are taken up by the gills and partly directed to the digestive gland, where intracellular uptake of NPs induces lysosomal perturbations and changes in the expression of antioxidant and immune-related genes. Nanoparticles can then potentially be translocated from the digestive system to the hemolymph and to circulating hemocytes, where nano-TiO<sub>2</sub> induced changes both in functional parameters (lysosomal



Figure 2. Possible routes of engineered nanomaterial (ENM) uptake and disposal in the model marine bivalve Mytilus leading to immunomodulation.

integrity, phagocytosis, reactive oxygen species, ROS, and NO production, induction of preapoptotic processes) and in transcription of antimicrobial peptides. Interestingly, the in vivo effects of nano-TiO<sub>2</sub> on mussel immune parameters were observed at concentrations  $(1-10 \mu g/L)$  much lower than those usually utilized in ecotoxicity tests on aquatic species, and closer to predicted environmental concentrations.<sup>65</sup> Recently, the rapidly expanding application of DNA microarrays and next generation sequencing (NGS) technologies offer new and broader research perspectives, from the whole transcriptome coverage to the Mytilus genome sequencing, leading to the identification of an increasing number of immune-related genes that could be targeted by different ENMs.66,67 Likely candidates are the members of the Toll receptor family recently identified in mussels;68 among these, transcription of the TRL-i isoform has been shown to be down-regulated in mussel hemocytes by in vivo exposure to nano-TiO<sub>2</sub>.<sup>69</sup>

Among invertebrates, the sea urchin is a successful marine model, globally distributed in almost all depths, latitudes, temperatures, and environments in the sea. The key to its successful survival is its

potent immune system, which provides protection, robustness, and molecular plasticity.<sup>70</sup> Thus, sea urchins represent an excellent model to uncover molecular and regulatory mechanisms promoting roles of the immune system on survival. Sea urchin immune cells have been demonstrated to activate their immune response machinery in response to different kinds of physical and chemical stressors, such as temperature shocks; pH decreases; exposure to UV-B radiation and heavy metals; and exposure to tin  $(SnO_2)$ . cerium (CeO<sub>2</sub>), iron (Fe<sub>3</sub>O<sub>4</sub>), and TiO<sub>2</sub> NPs.<sup>71–76</sup> The utility of the heat shock protein HSCP70/HSC70 as a general stress response marker to be used for monitoring both acute and chronic stresses has been demonstrated, with the only exception being NP exposure.<sup>76</sup> In contrast, it is expected that specific pathways and biomarkers are selectively elicited in response to NPs, as already demonstrated in human immune cells.77,78 The availability of the sea urchin genome, which has been shown to be closely phylogenetically related to the human genome,<sup>79</sup> offers the possibility to analyze its complex and sophisticated immune system and to compare the biological effects observed in sea urchins and human immune cells. Analysis of

VOL. 8 • NO. 10 • 9694-9709 • 2014

JAI



Figure 3. Sea urchin model to study immunity and development for the safe use of nanomaterials.

the amazingly large repertoire of innate pathogen recognition proteins, such as the 20-fold expansion and diversification of the Toll-like receptors relative to human genes, would probably identify specific target genes for different NPs or ENMs. Complementary to the use of adults for studies on immune modulation, embryos have emerged as valid tools for studies on developmental and molecular perturbations induced by environmentally relevant classical and emerging contaminants, including metal NPs.<sup>80–89</sup> As an example, the in vivo exposure to nano-Ag causes dose-dependent

developmental defects as well as alterations in swimming patterns in sea urchin embryos.<sup>56</sup> However, the cause—effect relationship remains to be demonstrated. In conclusion, the value of the sea urchin model is twofold: first, it is proposed as a proxy to humans for the analysis of the effects of ENMs on the immune system, and second, it can be used as an alternative model for ecotoxicological studies (see Figure 3).

Engineered nanomaterials could give rise to genotoxic effects in exposed marine organisms; the loss of DNA integrity, if not properly repaired, may lead to mutations, birth

defects, and long-term effects, such as cancer in vertebrates.<sup>90,91</sup> The mechanisms of ENMs genotoxicity are still not well understood<sup>92</sup> and it is often unclear if an effect on DNA is nanospecific. DNA strand breaks are one of the major types of oxidative damage to DNA via oxidative stress, which is generally assessed by the Comet assay,<sup>92,93</sup> a technique widely used to evaluate the genotoxic effects of contaminants in bivalves' hemocytes, which are a potential target for genotoxicity.59,64,94,95 Oxidative stress after long-term exposure to nano-Ag was evidenced in mussel gills and digestive glands.<sup>96</sup>

VOL.8 • NO.10 • 9694-9709 • 2014



www.acsnano.org

A loss of DNA integrity was found in marine mussel cells after in vitro exposure to nano-iron<sup>95</sup> and in blue mussel after in vitro exposure to nano-Ag<sub>2</sub>S and CdS quantum dots.<sup>97</sup> Concerning marine species, the effects of nano-TiO<sub>2</sub> have been investigated with respect to invertebrates,<sup>98-101</sup> and two articles have reported the susceptibility to micrometric rutile and nanometric anatase TiO<sub>2</sub> genotoxic potential of the top predator bottle-nose dolphin leukocytes<sup>51</sup> and fibroblasts.<sup>54</sup> Interactive genotoxic effects of C<sub>60</sub> fullerenes and fluoranthene were studied in marine mussels,<sup>102</sup> highlighting that both fluoranthene and  $C_{60}$  on their own caused concentration-dependent increases in DNA strand breaks, while combined exposure to  $C_{60}$ and fluoranthene additively enhanced the levels of DNA strand breaks, likely related to oxidative defense impairment. Recent evidence also showed that nano-TiO<sub>2</sub> and 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) can exert synergistic or antagonistic effects depending on the experimental condition, cell/tissue, and type of measured response, as evaluated by a battery of biomarkers including DNA primary damage (Comet assay), genome instability (RAPD assay), and chromosomal damage (Micronucleus test).<sup>103</sup>

Biotransformation/detoxification may play a significant role in ENM toxicity, including genotoxicity, by affecting ENMs localization/ disposition inside the cell. The accumulation and consequent toxicity of a compound is strongly influenced by the organism's detoxifying/clearance capabilities. Biotransformation of ENMs might alter their life-cycle metabolism, but also might interfere with that of other compounds, as occurs in coexposure scenarios in the natural environment. Therefore, it is crucial to investigate the interactions of ENMs with the so-called "cell defensome" (i.e., the cellular machinery activated in response to chemical stressors) in marine organisms that are naturally exposed to mixtures of compounds/toxicants

present in the environment.4,104 Among actors on the first line of defense, P-glycoprotein (P-gp) is the most well-characterized membrane transport protein; it is also involved in the efflux of xenobiotics generally known as multidrug resistance (MDR, previously known as MXR), i.e., phase 0 and III of biotransformation.<sup>105,106</sup> Several ENMbased approaches have recently been proposed to reverse/overcome the efflux-mediated resistance controlled by P-gp in human cell lines to favor cytotoxic drugs accumulation in tumor cells. Of course, such complex medical ENMs are transformed inside the body before expulsion.<sup>107–109</sup> Little attention has been paid to marine organisms where MDR is actively involved in xenobiotic biotransformation and a similar interaction may occur. One example is the reported increase in the efflux of P-gp substrate Rhodamine B in the gills of the marine mussel Mytilus galloprovincialis caused by nano-TiO<sub>2</sub>.<sup>103</sup> In parallel, an increase of TCDD bioconcentration in the whole body was observed in co-exposure conditions. Possible involvement of nano-TiO<sub>2</sub> on efflux functionality that, in turn, affects the disposition/cellular retention of other toxic compounds can be hypothesized.<sup>103</sup> Concerning heavy metal biotransformation, metallothioneins (MTs) are ubiquitous, low-molecular-weight soluble proteins with high heavy-metal-ionbinding capacity. Their role in essential trace elements  $(Zn^{2+} and Cu^{+})$ homeostasis as well as detoxification from noxious ions such as Hg<sup>2+</sup> and Cd<sup>2+</sup> has been widely demonstrated in eukaryotic cells, including those of marine organisms.<sup>110</sup> The increase of MT in response to a variety of oxidebased and metallic NPs has been reported in cells and/or tissues of several marine organisms. Nano-Ag increased MT gene expression in embryo and adult hepatopancreas cells of Crassostrea virginica55 in the nanomolar range. Conversely, no increase in MT-like proteins was reported in tissues of the ragworm

Hediste diversicolor exposed to nano-Ag in sediments, whereas soluble Ag caused MT induction.<sup>111</sup> Gold NPs (0.1 mg/L) were effective in tissues of the endobenthic clam Scrobicularia plana.<sup>112</sup> Among nano oxides, despite the number of studies on the effects of ZnO and nano-TiO<sub>2</sub>, only reports on nano-TiO<sub>2</sub> are available, demonstrating no effects on MT transcription in the tissues of in M. galloprovincialis.<sup>69,101</sup> However, several studies have focused on the effects of nano-CuO, for which an increase of MT proteins has been documented in at least two molluscan species, M. galloprovincialis and S. plana.<sup>113–116</sup> It is worth noting that in the latter studies, environmental Cu dissolution was very low or virtually absent, therefore MT induction had three possible explanations: (1) intracellular Cu dissolution to ionic forms; (2) oxyl-radical formation; or, most likely, (3) a combination of both, since metal and oxidative stress show overlapping features, cellular outcomes, and biochemical responses. It is widely accepted that MT genes are transcriptionally activated by different heavy metal ions penetrating into cells through a mechanism involving a zinc-sensitive transcription factor (MTF-1) and zinc displacement from a physiological pool of MTs by more electrophilic cations. In addition, oxyl radicals and, in general, oxidative stress are promoters of MT induction.<sup>117</sup> Authors did not report MT modulation along with effects to the antioxidant enzymatic system and/or the occurrence of oxidative insults in the aforementioned works (except in ref 111, where no NP dissolution was observed). Assaying both MT heavy metal content and oxidation state and its concentration in marine organisms' tissue thus represents a powerful tool to provide mechanistic information on the mode of action of metallic and metal-oxide NPs.

The Need To Develop Models. A major need in marine ecotoxicology for ENMs is the development of quantitative approaches for integrating more realistic ENM exposure scenarios

VOL. 8 • NO. 10 • 9694-9709 • 2014



www.acsnano.org



Figure 4. Process using estuarine fate and transport modeling to predict realistic exposure scenarios to design focused fate and transport (F&T), high-content screening assays (HCS), and microcosm experiments with individuals and populations of estuarine fauna. Results of the experiments generate predictions for impacts under realistic estuarine environmental scenarios in mesocosms. Results from F&T, HCS, microcosm, and mesocosms are then synthesized in multivariate machine-learning statistical analyses, the results of which update the estuarine F&T model.

and predicting ecosystem impacts, thus enhancing the ability to design ecosafe ENMs. This can be accomplished through modeling exposure scenarios specific to estuaries and marine environments, testing those predictions with environmental fate and transport (F&T) experiments, and testing toxicity with organisms that could be exposed in both pelagic and benthic marine habitats (see Figure 4). This integration requires specifically: (1) generating predictive F&T models for specific ENMs and environments, such as multimedia environmental distribution (MendNano) models for coastal or estuarine ecosystems-similar models have been produced for terrestrial environments<sup>118</sup> and allow flows of ENMs within and between environmental and biological media; (2) testing the predictions of the MendNano models through a combination of F&T experiments, high-content-screening (HCS) assays, microcosms, and computerregulated mesocosm experiments at the level of individuals, populations, and/or communities; (3) analyzing the results of the multiple experiments for correlations using machine learning

analysis and statistical modeling framework;<sup>119</sup> and (4) using the correlation output to parametrize and update the MendNano F&T model. A full array of ENMs of different shapes and sizes, such as Cu, CuO, core-shell, and safe design materials as they are developed, can be studied when they are exposed to water, sediment, and biological matrices, and as they are transformed by physical (e.g., temperature, salinity, and dissolved and particulate organic material) and biological processes associated with estuarine environments and food webs composed of bacteria, plants, invertebrates, and fish. Both the benthic and pelagic communities, from sedimentdwelling invertebrates to fish and important estuarine fauna, represent potential targets of ENMs. The microbial community is crucial in determining ENMs' F&T and effects; impacts to phytoplankton, for example, which are the primary producers in estuaries, probably have manifold ecological effects. The objectives of this approach are structured to assess the environmental implications of ENMs through integration models that generate theory, experiments that test

those theories, and subsequent improvement of the models to generate better theory. Marine ecotoxicology efforts use this approach by linking ENMs' chemical properties, exposure, and biochemical responses of injury with the ecological and physical processes that ultimately regulate marine ecosystem-level impacts and ecosystem services (see Figure 4). The general observations produced from this approach so far are the following:

(1) Engineered nanomaterials can be toxic to bacteria;<sup>5</sup> in response to external aggression, algae and bacteria can release extracellular polymeric substances (EPS)<sup>120</sup> that will potentially bind to metal ENMs, perhaps reducing their bioavailability and toxicity.<sup>121,122</sup> The role of flocculation of EPS ("marine snow") in the fate of ENMs remains vastly unappreciated.

(2) Oysters are estuarine filter feeders that provide many important ecosystem services.<sup>123</sup> As filter feeders, they process large volumes of water and are exposed to estuarine ENMs through ventilation for respiration, thus acting as potential ENM bioaccumulators from the

VOL.8 NO.10 9694-9709 2014



pelagic habitat. Benthic infaunal invertebrates, especially polychaete worms, crustaceans, echinoderms, and deposit-feeding clams, are probably exposed to ENMs deposited onto sediments, detritus, and biofilms.<sup>116,120</sup> Together, these organisms, which are important in estuarine food webs, may store ENMs or their dissolution products in soft or hard tissue, and may also, through excretion, enhance ENM bioavailability. This may or may not represent an important pathway for human exposure, as well as for the trophic transfer of ENMs or their dissolution products. Fish are top predators in coastal ecosystems and can be exposed to metal ENMs directly through water column ingestion or gill tissue exposure. However, many fish species are also predators on numerous sediment-dwelling invertebrates that will have consumed ENMs, so this trophic transfer may represent an important route of exposure.

(3) Dynamic energy budget (DEB) theory<sup>124</sup> focuses on the individual organism, with differential equations describing the rates at which an organism assimilates and utilizes energy and materials from food for maintenance, growth, reproduction, and development. Engineered nanomaterials may disrupt energy utilization, as do many other anthropogenic contaminants. Dynamic energy budget models use theory to provide estimates of dosespecific responses of ENM concentrations on individual growth, survival, and reproductive capacity.<sup>125,126</sup>

(4) The outputs of DEB models can be used in population dynamic model simulations to predict the potential impact of ENM injuries on populations. For small organisms such as phytoplankton, for which population dynamics can be measured in microcosms or mesocoms, the population dynamic models can be used as predictions and then tested empirically with follow-up experiments. For larger organisms, for which population level manipulations are impractical, the models can provide theoretical impacts. A major need in marine ecotoxicology for engineered nanomaterials (ENMs) is the development of quantitative approaches for integrating more realistic ENM exposure scenarios and predicting ecosystem impacts, thus enhancing the ability to design ecosafe ENMs.

Risk Assessment, Including Nanotechnology Applications in Marine Waters. The increasing exposure of aquatic organisms to ENMs may lead to extended ecological risks. Considerable research efforts in Europe have focused on evaluating the risks from ENMs in the aquatic environment (see Table 1). In the United States, the Nanotechnology Environmental and Health Implications Working Group of the National Nanotechnology Initiative coordinates the research that aims to develop protocols, standards, instruments, models, and validated data for ecological risk assessment (RA) of ENMs.127

Despite the significant research efforts, a key issue in both the European Union and the United States that most research has not sufficiently addressed is that pristine ENMs undergo aging and transformation reactions during incorporation into products, and weathering and aging when released into the environment.<sup>121</sup> Thus, at each step of the supply chain (lifecycle), there is a potential risk of exposure to ENMs with different physicochemical properties, which would affect their toxicity. Some of the few results for textiles, paints, and nanocomposites<sup>32</sup> suggest that the

released particles undergo significant aging<sup>128</sup> and exhibit different environmental behavior and effects compared to the pristine ENMs.45 Aged and pristine Cu ENMs showed different fate and toxicity in an aquatic compartment under the same environmental conditions.129 In addition, aged nano-TiO<sub>2</sub> from sunscreen caused lower mortality in Daphnia magna compared to bare nano-TiO<sub>2</sub>.<sup>130</sup> These studies have confirmed that aged ENMs can exhibit differences in solubility, aggregation, and reactivity that can affect their aquatic mobility and toxicity. Moreover, it is hypothesized that many ENMs are (designed to be) persistent, which might lead to long-term exposure of aquatic and sediment species. However, longerterm ecological effects have not yet been comprehensively researched, especially for ENMs used in real products. Experiments with single species have provided evidence of changes being transferred across generations, causing epigenetic, mutational, or reproductive effects.<sup>131</sup> Therefore, testing of longer-term genotoxicity on multiple species should be developed. Quantitative ecological risk analyses of ENMs would typically involve deterministic modeling of exposure-doseresponse relationships. However, this would be affected by severe uncertainty and data variability. Therefore, it is recommended that the ecological risk modeling of ENMs is addressed in a probabilistic manner using stochastic approaches such as the Monte Carlo and the Latin hypercube simulations.<sup>132</sup> In this case, distributions of hazard estimates would be derived instead of single values, which could be plotted against distributions of exposure estimates in order to identify central tendencies of expected risk and associated high-end probability of exposure. With this approach, predicted no effect concentrations (PNEC) can be estimated based on species sensitivity distribution (SSD) methodologies,<sup>32</sup> which are well suited to deal with high uncertainty



TABLE 1. Overview of European Union Research Projects Providing Data and Methods for Ecological Risk Assessment of Engineered Nanomaterials (ENMs) in the Aquatic Environment<sup>a</sup>

Project Acronym	eNanoMapper	FutureNanoNeeds	GuideNano	MARINA	MembraneNanoPart	ModNanoTox	NanoDefine	NanoDetector	NanoFATE	NanoHeter	NanoMiCex	NanoMILE	NanoPolyTox	NanoPuzzles	NANOREG	NanoRISK	NanoSolutions	nanoSTAIR	NanoSustain	NanoTransKinetics	NanoValid	NanOxiMet	PreNanoTox	QualityNano	SIINN	SIRENA	SUN
Start year	2014	2014	2013	2011	2013	2011	2013	2012	2010	2013	2012	2013	2010	2013	2013	2013	2013	2012	2010	2011	2011	2013	2013	2011	2013	2013	2013
End year	2017	2017	2016	2015	2015	2013	2017	2015	2014	2016	2014	2017	2013	2015	2016	2016	2017	2014	2013	2014	2014	2016	2016	2015	2016	2015	2017
Measurement		x		х			х	x	х				x				х		x		х						
Physico-chemical																											
properties		X	X	х	х		х	x	х	x	х	X	X		X	x	х		x		х	х	х	X	х		X
Analysis of "next																											
generation" nanomaterials																											
(2nd, 3rd or 4th generation)		X	X				Х					<u> </u>				X								<u> </u>	Х		X
Exposure assessment for		X	X	х					Х	X	X		X			X			X		х						X
Develop & validate																											
exposure measurement																											
and modelling methods		x	x	х		x			x	x	x		x	x	x	x				x	x				х	х	x
Environmental Exposure																											1
Assessment		X	X	х		х			х	х	х		X		X	x			X		х				х	х	X
Interaction of NM with																											
biological systems		X		Х					Х				X				Х		Х		Х						
Interaction with																											
physiological mechanisms		X		Х	Х				Х				X	Х	Х				Х	X	Х	Х	Х	X			
Toxicokinetics				х	Х				Х		Х		X	X	X				X		х						
variability									Х				X	X	Х									X			
Predictive models				х	Х	Х			Х		Х		X	X	X		х		X	X	х	Х					X
Long term monitoring and																											x
assessment									X				X		X									<u> </u>			
Ecotoxicology		X	X	х					X			X		X			X		X		X						
Develop testing and		~	1 v	~					~		~			×	~		~		×		~						~
assessment strategy		^	^	~					~		~	^		· ^	~		~		~		~			<u> </u>			-
Apply testing and				v		~			v						~		~				~						
Proliminary handling				^		<u>^</u>						<u> </u>					^				^			<u> </u>			<u> </u>
guidelines				x										x		x	x		×		x						×
Collect available and				~													~				~						
ongoing approaches				x										x	x	x	x		x		x					x	×
Evaluation and further																											
development		x		х										x	x	x	х		x		x					х	x
Information transfer		X	X						Х								х				х						
Database generation	х	X	X	х		Х			Х	Х		X	X	X	Х		х		х	X	х	х	х	X		Х	X
Public dialogue			X	Х					Х	Х				X		Х					Х						
Information to and training																											
		X	X		<u> </u>	<u> </u>	<u> </u>	<u> </u>	X	<u> </u>	Х	X	X	<u> </u>	<u> </u>	X			X	<u> </u>	Х	Х		<u> </u>		Х	X
National and international				~					~								~	~			~						
collaboration	~	- V	- N	X				×	X	X		-	X	X	×	×	X	X	X		X	×	~		~	~	
Development	X	X	X	X	v			X	X	X	v		X	X	X	X	X		X		X	X	X	X	X	X	×
l esting	×	-		~	Ň		v	~	×	×	~					^	×			V	×	~	×		×	×	
Validation	×		×	~	^		Ŷ		Ŷ	×				Ŷ	Ŷ		^	×	-	×	×	×	×	$\frac{1}{2}$	~	×	÷
Assessment activities	^		Y	Ŷ			^		Ŷ	Ŷ	x		Ŷ	Ŷ	Ŷ		×	^	×	Ŷ	Ŷ	^	Ŷ	$\hat{\mathbf{x}}$		^	x
Assessment activities		1	ΙŇ	^							_ ^	1				1	~		_ ∧		~		~	_ ∧			

<sup>a</sup> Reproduced unchanged with permission from NanoSafety Cluster; European NanoSafety Cluster Compendium: Compendium of Projects in the European NanoSafety Cluster, 2014 Edition; Lynch, I., Ed.; Karolinska Institutet on behalf of WG8 of the NanoSafety Cluster, access at http://www.nanosafetyduster.eu/uploads/files/pdf/2014\_NSC\_Compendium. pdf, under the Creatives Commons Attribution license 3.0 (http://creativecommons.org/licenses/by/3.0/). Copyright 2014 EU NanoSafety Cluster.

and variability. Such SSD modeling procedures produce the biological responses of each species of a target environmental system and combine them into a cumulative SSD for the aquatic compartment. Probabilistic estimation of ecological risks can then be conducted by linking this SSD to the probability of critical predicted environmental concentrations (PEC). Such modeling approaches for ecological risk estimation and the resulting risk mitigation measures can provide a strong basis for guidance in industrial and regulatory contexts.

**ENMs for Environmental Applications:** Smart Nanoparticles for Sea Water Remediation. Most of the remediation

Nanomaterials have a number of physicochemical properties that make them particularly attractive for water purification and environmental remediation.

technologies available today are based on the adsorption, ion

exchange, amalgamation, chemical degradation or precipitation, and bioremediation. While effective in many cases, these methods are often costly and time-consuming, particularly pump-and-treat methods that require expensive investments and procedures. In this context, the high adsorption capacity of ENMs for certain pollutants has been demonstrated in many cases. There are a variety of examples in the literature that exploit the use of NPs for the removal of heavy metals from aqueous solutions, such as using  $\beta$ -FeO(OH) nanocrystals to remove As(V),133 SiO2 NPs for removal of Cr(III),134 as well as AI2O3 NPs135

VOL. 8 • NO. 10 • 9694-9709 • 2014



www.acsnano.org

and supported Au NPs on alumina<sup>136</sup> and as colloids,<sup>137</sup> for removal of Hg(II). Although the ability of inorganic NPs to trap pollutants is best known for cations of elements such as the previous and Ni(II) and Cd(II), they can also be functionalized to trap organic molecules specifically and/or to act as photocatalysts to promote the complete degradation of toxic organic matter. A comprehensive overview of the different manufactured nanomaterials along with the pollutants they could potentially remediate has been reviewed elsewhere.<sup>2,138</sup> The toxic species once absorbed onto the NPs can be removed by applying mild (and affordable) gravitational (centrifugation) or magnetic (in the case of magnetic NPs) field gradients. Other strategy when dealing with organic compounds is to promote their reduction (e.g., to transform perchlorates to Cl<sup>-</sup>) with sacrificial electrodes, as iron NPs that will reduce halogenated compounds during their oxidation. Moreover, nanoremediation can present complementary properties to bioremediation, not just by decreasing pollutants, but also by dispersing the pollutants, and by synergetic interactions with biota. The majority of these works have been performed in the lab with synthetic water due to complexity and restrictions to disperse NPs in the natural environment.

All this research interest is because nanomaterials have a number of physicochemical properties that make them particularly attractive for water purification and environmental remediation. Their capacity to be dispersed in water allows them to travel farther than larger, macrosized particles, thus achieving wider distribution and permitting the whole volume to be quickly scanned with a relatively small amount of material.<sup>2,139,140</sup> These unique properties can be employed to degrade and to scavenge pollutants (see Table 2).

Nanoremediation may address different levels of pollution with specific targeted technologies, which

may be used in a stepwise strategy (see Figure 6), e.g., (i) cleaning very polluted sites toward less polluted soil, after which biota may be able to remediate pollutants further; (ii) cleaning less polluted sites with existing microbiota by developing synergies between the NPs and the involved biota to increase the potential for natural attenuation; and finally, (iii) cleaning low but hazardous concentrations of pollutants (microcontaminants). Very polluted areas, where all signs of life have disappeared, and therefore precluding the option of bioremediation, are a big challenge for remediation and a wonderful opportunity to test the reactivity, catalytic behavior, and large surface area per unit mass of NPs. In such cases, NPs are needed so they can induce/catalyze the initial chemical breakdown or adsorb and then slowly release the contaminants in order to achieve a level of contamination where microbial processes can take place, enhancing the sequential synergy of nanotechnology and bioremediation. Ideally, the ENM would degrade into harmless substances once its job is finalized. In addition, while cleaning very polluted water into less polluted water is feasible, cleaning low pollution levels in water to safe water (especially in the case of microcontaminants) is also a challenge that could be overcome by nanoremediation. However, the intrinsic nature of the inorganic NPs may also cause damage to the environment if dispersed uncontrollably.

Marine pollution by petroleum products (*i.e.*, oil spills) often produces disastrous effects with negative impacts, affecting both the environment and socioeconomic development. In this context, the use of ENMs for *in situ* remediation represents a promising and cutting-edge solution, by ensuring a quick and efficient removal of pollutants.<sup>2</sup> However, the use of such materials should not pose any additional risk to the marine ecosystem in which they are released.<sup>2,141</sup>

Developing safe and effective remediation technologies for petroleum products based on the design and synthesis of new ecofriendly ENMs might be a challenging solution for promoting in situ nanoremediation for the marine environment. Such innovative ecofriendly ENMs should meet the highest standards of environmental safety and effectiveness, supporting economic development in terms of industrial competitiveness and innovation. Their application will contribute to the management, in more efficient and sustainable ways, of several systematic sources of petroleum pollution in various productive sectors, such as maritime transport, refining, mining, and liquefied natural gas terminals in marine waters. Finally, the use of ecofriendly ENMs, which provide rapid and effective removal of petroleum products without compromising the ecosystem, is a priority for the safeguard of the oceans.

## FUTURE PERSPECTIVES AND FORESEEN CHALLENGES

As mentioned, the deliberate or accidental release of ENMs into the oceans will impact marine organisms, especially if solutions for their safe manufacturing are not properly considered. To this purpose, two main organizations, the University of California Center for Environmental Implications of Nanotechnology (UC CEIN) and the Marine Focus Group in the framework of Hazard Working Group of the NanoSafety Cluster of the European Commission, focused their mission on studying the impacts of ENMs on the marine environment. The UC CEIN was established in 2008 with funding from the U.S. National Science Foundation and the U.S. Environmental Protection Agency, with the aim of studying the impact of nanotechnology on the environment, including the identification of hazard and exposure scenarios that take into consideration the novel physicochemical properties of ENMs. The UC CEIN has made great progress in

VOL. 8 • NO. 10 • 9694-9709 • 2014



TABLE 2. Remediation Mechanisms of Actions of Different Engineered Nanomaterials							
mechanism of action							
CATALYST	Au, Pt, Ag, Pd, Fe <sub>3</sub> O <sub>4</sub> , CoO, CeO <sub>2</sub>						
oxidation and reduction							
e <sup>-</sup> SOURCES	Fe, Cu, Co, Ni, $Fe_3O_4$ , CeO <sub>2</sub>						
Known Active Ion PROVIDERS	Fe <sub>3</sub> O <sub>4</sub> , Ag, Pt, CeO <sub>2</sub> , ZnO						
ABSORBERS	Fe <sub>3</sub> O <sub>4</sub> , Fe <sub>2</sub> O <sub>3</sub> , TiO <sub>2</sub> , SiO <sub>2</sub> (porous, partially methylated), functionalized Layered Double Hydroxides (LDH)						
HYBRIDES I	Au—CeO <sub>2</sub> , Pd/Cu—CeO <sub>2</sub> , Au—TiO <sub>2</sub> , Au—Fe <sub>3</sub> O <sub>4</sub> , CeO <sub>2</sub> —Fe <sub>3</sub> O <sub>4</sub>						
Heterodimer							
HYBRIDES II	Au-SiO <sub>2</sub> , $Fe_3O_4$ —SiO <sub>2</sub> , $Fe_7e_3O_4$						
Core—shell							
Tautomeric species ("oxygen/electron sponges")	$CeO_2$ , $Fe_3O_4$ , $FeMnO_x$ (perovsquites), $MnO_x$						

## Marine and freshwater ecosystems impact and ecotoxicology



and high-content screening (HCS) assays, and injury responses across different levels of biological organization. Dynamic

energy budget (DEB) models enable the linkage of individual responses to population and community levels.

Figure 5. Integration of engineered nanomaterials (EMN) chemistry, exposure modeling, high-throughput screening (HTS)

assembling a multidisciplinary team to develop the scientific underpinnings, research, knowledge, education, and outreach that is required for assessing the safety of nanotechnology in the environment. The approach includes high-throughput/ high content screening (HTS and HCS) studies to develop structure-activity relationships (QSARs) that can be used to predict the impact of ENMs on organisms in freshwater, marine, and terrestrial environments.<sup>142,143</sup> A major goal of UC CEIN is to integrate research goals and themes to better estimate realistic nanomaterial exposure scenarios (F&T), predict ecosystem impacts, and design safer materials. UC CEIN's strategies for accomplishing this goal include

iterative learning between environmental modeling, chemistry, toxicology, ecosystem impacts, and sociological research efforts. Marine studies have included species of phytoplankton as primary producers, and copepods and mussels as primary consumers linking photosynthesis as well as ENM transfer to higher trophic levels. The effect of ENMs on embryo development is also a focus of the Center. The UC CEIN has been renewed (2013–2018) and new directions are planned to address the integrated marine nanotoxicology effort. A central role is to identify which ENMs have important environmental impacts versus those that are safe. UC CEIN's marine ecotoxicology effort uses this approach

by linking ENM chemical properties, exposure, and biochemical responses of injury with the ecological and physical processes that ultimately regulate ecosystem-level impacts and ecosystem services.

Within the NanoSafety Cluster of the European Commission, the Marine Ecotox Focus Group in the framework of the Hazard Working Group (http://www.nanosafetycluster.eu/working-groups/2-hazard-wg/ marine-ecotox.html) was established in 2011. Its mission is based on the following assumptions: (1) the marine environment is likely to be a sink of ENMs as it is for most manmade pollutants, and it is therefore possible that certain ENMs may bioaccumulate and biomagnify along the

VOL.8 • NO.10 • 9694-9709 • 2014 A







Figure 6. Combining nano- and bioremediation.

marine trophic chain, thus potentially affecting marine biological resources (wild and farmed); (2) ENMs may be transferred to humans through diet by consumption of contaminated seafood products; (3) ENMs may lead to a deterioration of marine environmental quality (coastal areas including natural and recreational interests) with social and economic repercussions; (4) some ENMs can, however, be used to reduce marine pollution, through selected applications, such as nanoremediation by binding and removing specific contaminants. The progress in the development of nanotechnologies and nanoenabled products is vast and continuously progressing. The aim of the Focus group is to offer a platform for linking nanotechnologists with ecotoxicologists, environmental scientists, analytical chemists, biochemists, molecular biologists, industries, and end users (public) to provide the proper scenario suitable for an overall risk assessment of ENMs in the marine environment. By doing this, the focus group will provide appropriate support for decision and policy makers, including an understanding of the risks that may occur for ENMs that fall outside the definition, guidance, further development of measurement techniques, and help dealing with changes during the life cycle

in agreement with recent work.<sup>144</sup> Again, at the EU level, the Quality-Nano pan-European infrastructure for quality in nanomaterials safety testing is providing support for a full assessment of potential impacts of past and next generation of ENMs at all stages of their lifecycle including interactions with environmental matrices and biota (http://www.qualitynano.eu/).

At the national level, the Italian Marine Nano Ecotoxicology working group was formally announced in November 2012 and further enlarged at the International level during the first Marine NanoEcoToxicolgy Workshop (MANET) held in Palermo in 2012 and fully dedicated to addressing the ecotoxicology of ENMs in the marine environment. Currently, it groups more than 50 scientists from 23 countries including France, Germany, Sweden, the United Kingdom, and the United States, involving both universities and research centers. Its mission is to encourage applied and basic research as a support for a safe and sustainable use of ENMs, fostering interdisciplinary approaches for research and training in the field of nanoecotoxicology and promoting informed discussion on ethical and relevant social issues as well. In this context, the Italian Marine Nano Ecotoxicology working group acts as a reference group for the

evaluation of the health and environmental impacts of ENMs with regard to scientific research, industrial innovation, and regulatory issues. It has the potential to act as a growing network integrating multidisciplinary skills and acquire the necessary expertise to fill the existing prioritized gaps.

*Conflict of Interest:* The authors declare no competing financial interest.

## **REFERENCES AND NOTES**

- Fabrega, J.; Luoma, S. N.; Tyler, C. R.; Galloway, T. S.; Lead, J. R. Silver Nanoparticles:Behavior and Effects in the Aquatic Environment. *Envir*on. Int. **2011**, *37*, 517–531.
- Karn, B.; Kuiken, T.; Otto, M. Nanotechnology and *in Situ* Remediation: A Review of the Benefits and Potential Risks. *Environ. Health Persp.* 2009, *117*, 1823–1831.
- Quigg, A.; Chin, W.-C.; Chen, C.-S.; Zhang, S.; Jiang, Y.; Miao, A.-J.; Schwehr, K. A.; Xu, C.; Santchi, P. H. Direct and Indirect Toxic Effects of Engineered Nanoparticles on Algae: Role of Natural Organic Matter. ACS Sustainable Chem. Eng. 2013, 1, 685–702.
- Matranga, V.; Corsi, I. Toxic Effects of Engineered Nanoparticles in the Marine Environment: Model Organisms and Molecular Approaches. *Mar. Environ. Res.* 2012, 76, 32–40.
- Holden, P. A.; Nisbet, R. M.; Lenihan, H. S.; Miller, R. J.; Cherr, G. N.; Schimel, J. P.; Gardea-Torresdey, J. L. Ecological Nanotoxicology: Nanomaterial Hazard Considerations at the Subcellular, Population, Community, and Ecosystems Levels. Acc. Chem. Res. 2013, 46, 813–822.

VOL.8 • NO.10 • 9694-9709 • 2014



- Baker, T. J.; Tyler, C. R.; Galloway, T. S. Impacts of Metal and Metal Oxide Nanoparticles on Marine Organisms. *Environ. Pollut.* 2014, 186, 257–271.
- Angelini, G.; De Maria, P.; Fontana, A.; Pierini, M.; Maggini, M.; Gasparrini, F.; Zappia, G. Study of the Aggregation Properties of a Novel Amphilic C<sub>60</sub> Fullerene Derivative. *Langmuir* **2001**, *17*, 6404– 6407.
- Kallay, N.; Zalac, S. Stability of Nanodispersions: A Model for Kinetics of Aggregation of Nanoparticles. J. Colloid Interface Sci. 2002, 253, 70–77.
- Brant, J.; Lecoanet, H.; Wiesner, M. R. Aggregation and Deposition Characteristics of Fullerene Nanoparticles in Aqueous Systems. J. Nanopart. Res. 2005, 7, 545–553.
- Chen, K. L.; Mylon, S. E.; Elimelech, M. Aggregation Kinetics of Alginate-Coated Hematite Nanoparticles in Monovalent and Divalent Electrolytes. *Environ. Sci. Technol.* 2006, 40, 1516–1523.
- Guzman, K. A. D.; Finnegan, M. P.; Banfield, J. F. Influence of Surface Potential on Aggregation and Transport of Titania Nanoparticles. *Environ. Sci. Technol.* 2006, 40, 7688–7693.
- French, R. A.; Jacobson, A. R.; Kim, B.; Isley, S. L.; Penn, R. L.; Baveye, P. C. Influence of Ionic Strength, pH, and Cation Valence on Aggregation Kinetics of Titanium Dioxide Nanoparticles. *Environ. Sci. Technol.* 2009, 43, 1354–1359.
- Von Smoluchowski, M. Versuch Einer Mathematischen Theorie der Koagulation. Z. Phys. Chem. 1917, 92, 129–155.
- Derjaguin, B.; Landau, L. Theory of the Stability of Strongly Charged Lyophilic Sols and of the Adhesion of Strongly Charged Particles in Solutions of Electrolytes. *Acta Phys. Chim.* **1941**, *14*, 633–662.
- Verwey, E. J. W.; Overbeek, J. T. G. Theory of the Stability of Lyophobic Colloids. J. Phys. Chem. 1947, 51, 631–636.
- Wu, W.; Giese, R. F.; van Oss, C. J. Stability versus Flocculation of Particle Suspensions in Water-Correlation with the Extended DLVO Approach for Aqueous systems, Compared with Classical DLVO Theory. Colloid Suff. B 1999, 14, 47–55.
- van Oss, C. J. Long-Range and Short-Range Mechanisms of Hydrophobic Attraction and Hydrophilic Repulsion in Specific and Aspecific Interactions. J. Mol. Recognit. 2003, 16, 177–190.
- Casals, E.; Vazquez-Campos, S.; Bastús, N.; Puntes, V. Distribution and Potential Toxicity of Engineered Inorganic Nanoparticles and Carbon Nanostructures in Biological Systems. *Trends Anal. Chem.* 2008, 27, 672–683.

- Labille, J.; Brant, J. Stability of Nanoparticles in Water. *Nanomedicine* 2010, *5*, 985–998.
- Chen, K. L.; Elimelech, M. Aggregation and Deposition Kinetics of Fullerene (C-60) Nanoparticles. *Langmuir* 2006, *22*, 10994–11001.
- Chen, K. L.; Elimelech, M. Influence of Humic Acid on the Aggregation Kinetics of Fullerene (C<sub>60</sub>) Nanoparticles in Monovalent and Divalent Electrolyte Solutions. *J. Colloid Interface Sci.* 2007, 309, 126–134.
- Domingos, R. F.; Tufenkji, N.; Wilkinson, K. J. Aggregation of Titanium Dioxide Nanoparticles: Role of a Fulvic Acid. *Environ. Sci. Technol.* 2009, 43, 1282–1286.
- Saleh, N. B.; Pfefferle, L. D.; Elimelech, M. Influence of Biomacromolecules and Humic Acid on the Aggregation Kinetics of Single-Walled Carbon Nanotubes. *Environ. Sci. Technol.* 2010, 44, 2412–2418.
- Wells, M. L. Marine Colloids—A Neglected Dimension. *Nature* 1998, 391, 530–531.
- Findlay, A. D.; Thompson, D. W.; Tipping, E. The Aggregation of Silica and Haematite Particles Dispersed in Natural Water Samples. *Colloid Surface A* 1996, 118, 97–105.
- Keller, A. A.; Wang, H. T.; Zhou, D. X.; Lenihan, H. S.; Cherr, G.; Cardinale, B. J.; Miller, R.; Ji, Z. X. Stability and Aggregation of Metal Oxide Nanoparticles in Natural Aqueous Matrices. *Environ. Sci. Technol.* 2010, 44, 1962–1967.
- Chinnaponge, S. L.; MacCuspie, R. I.; Hackley, V. A. Persistence of Singly Dispersed Silver Nanoparticles in Natural Freshwaters, Synthetic Seawater, and Simulated Estuarine Waters. *Sci. Total Environ.* 2011, 409, 2443–2450.
- Casals, E.; Pfaller, T.; Duschl, A.; Oostingh, G. J.; Puntes, V. Time Evolution of the Nanoparticle Protein Corona. ACS Nano 2010, 4, 3623–3632.
- Casals, E.; Pfaller, T.; Duschl, A.; Oostingh, G. J.; Puntes, V. F. Hardening of the Nanoparticle-Protein Corona in Metal (Au, Ag) and Oxide (Fe<sub>3</sub>O<sub>4</sub>, CoO, and CeO<sub>2</sub>) Nanoparticles. Small **2011**, 7, 3479–3486.
- Wells, M. L.; Goldberg, E. D. Marine Submicron Particles. *Mar. Chem.* 1992, 40, 5–18.
- Gustafsson, O.; Duker, A.; Larsson, J.; Andersson, P.; Ingri, J. Functional Separation of Colloids and Gravitoids in Surface Waters Based on Differential Settling Velocity: Coupled Cross-Flow Filtration-Split Flow Thin-Cell Fractionation (CFF-SPLITT). Limnol. Oceanogr. 2000, 45, 1731–1742.
- Gottschalk, F.; Sun, T. Y.; Nowack, B. Environmental Concentrations of Engineered Nanomaterials: Review of Modeling and Analytical Studies. Environ. Pollut. 2013, 181, 287–300.

- Guo, L. D.; Santschi, P. H.; Warnken, K. W. Dynamics of Dissolved Organic Carbon (DOC) in Oceanic Environments. *Limnol. Oceanogr.* 1995, 40, 1392–1403.
- Stolpe, B.; Hassellov, M. Nanofibrils and other Colloidal Biopolymers Binding Trace Elements in Coastal Seawater: Significance for Variations in Element Size Distributions. *Limnol. Oceanogr.* 2010, 55, 187– 202.
- Stolpe, B.; Hassellov, M. Changes in Size Distribution of Fresh Water Nanoscale Colloidal Matter and Associated Elements on Mixing with Seawater. *Geochim. Cosmochim. Acta* 2007, 71, 32923301.
- Chin, W. C.; Orellana, M. V.; Verdugo, P.; Verdugo, P. Spontaneous Assembly of Marine Dissolved Organic Matter into Polymer Gels. *Nature* 1998, 391, 568–572.
- Santschi, P. H.; Balnois, E.; Wilkinson, K. J.; Zhang, J. W.; Buffle, J.; Guo, L. D. Fibrillar Polysaccharides in Marine Macromolecular Organic Matter as Imaged by Atomic Force Microscopy and Transmission Electron Microscopy. *Limnol. Oceanogr.* 1998, 43, 896–908.
- Verdugo, P.; Alldredge, A. L.; Azam, F.; Kirchman, D. L.; Passow, U.; Santschi, P. H. The Oceanic Gel Phase: a Bridge in the DOM-POM Continuum. *Mar. Chem.* 2004, *92*, 67–85.
- Ding, Y. X.; Hung, C. C.; Santschi, P. H.; Verdugo, P.; Chin, W. C. Spontaneous Assembly of Exopolymers from Phytoplankton. *Terr. Atmos. Oceanic Sci.* 2009, 20, 741–747.
- 40. Stolpe, B.; Guo, L. D.; Shiller, A. M.; Hassellov, M. Size and Composition of Colloidal Organic Matter and Trace Elements in the Mississippi River, Pearl River and the Northern Gulf of Mexico, as Characterized by Flow Field-Flow Fractionation. *Mar. Chem.* **2010**, *118*, 119–128.
- Hesselin, F. F.; Vrij, A.; Overbeek, J. T. Theory of Stabilization of Dispersions by Adsorbed Macromolecules Interaction Between Two Flat Particles. J. Phys. Chem. **1971**, 75, 2094–2103.
- Napper, D. H. Steric Stabilization. J. Colloid Interface Sci. 1977, 58, 390–407.
- Fleer, G. J.; Scheutjens, J. M. H. M.; Stuart, M. A. C. Theoretical Progress in Polymer Adsorption, Steric Stabilization and Flocculation. *Colloids Surf.* **1988**, *31*, 1–29.
- Labille, J.; Thomas, F.; Bihannic, I.; Santaella, C. Destabilization of Montmorillonite Suspension by Ca<sup>2+</sup> and Succinoglycan. *Clay Miner.* 2003, *38*, 173–185.
- Labille, J.; Feng, J. H.; Botta, C.; Borschneck, D.; Sammut, M.; Cabie, M.; Auffan, M.; Rose, J.; Bottero, J. Y. Aging of TiO<sub>2</sub> Nanocomposites Used in Sunscreen. Dispersion and Fate of the Degradation Products



in Aqueous Environment. *Environ. Pollut.* **2010**, *158*, 3482–3489.

- Labille, J.; Fatin-Rouge, N.; Buffle, J. Local and Average Diffusion of Nanosolutes in Agarose Gel: The Effect of the Gel/Solution Interface Structure. *Langmuir* 2007, 23, 2083–2090.
- Pletikapic, G.; Zutic, V.; Vrcek, I. V.; Svetlicic, V. Atomic Force Microscopy Characterization of Silver Nanoparticles Interactions with Marine Diatom Cells and Extracel-Iular Polymeric Substance. J. Mol. Recognit. 2012, 25, 309–317.
- Hyung, H.; Fortner, J. D.; Hughes, J. B.; Kim, J. H. Natural Organic Matter Stabilizes Carbon Nanotubes in the Aqueous Phase. *Environ. Sci. Technol.* 2007, 41, 179184.
- Dutta, N.; Green, D. Nanoparticle Stability in Semidilute and Concentrated Polymer Solutions. *Langmuir* 2008, 24, 5260–5269.
- Louie, S. M.; Tilton, R. D.; Lowry, G. V. Effects of Molecular Weight Distribution and Chemical Properties of Natural Organic Matter on Gold Nanoparticle Aggregation. *Environ. Sci. Technol.* 2013, *47*, 4245–4254.
- Bernardeschi, M.; Guidi, P.; Scarcelli, V.; Frenzilli, G.; Nigro, M. Genotoxic Potential of TiO<sub>2</sub> on Bottlenose Dolphin Leukocytes. Anal. Bioanal. Chem. **2010**, 396, 619–623.
- Griffitt, R. J.; Brown-Peterson, N.-J.; Savin, D. A.; Mannings, C. S.; Boube, I.; Ryan, R. A.; Brouwer, M. Effects of Chronic Nanoparticulate Silver Exposure to Adult and Juvenile Sheepshead Minnows (*Cyprinodon variegatus*). *Environ. Toxicol. Chem.* 2012, *31*, 160–167.
- Wang, J.; Wang, W. X. Low Bioavailability of Silver Nanoparticles Presents Trophic Toxicity to Marine Medaka (*Oryzias melastigma*). Environ. Sci. Technol. 2014, 48, 8152– 8161.
- 54. Frenzilli, G.; Bernardeschi, M.; Guidi, P.; Scarcelli, V.; Lucchesi, P.; Marsili, L.; Fossi, M. C.; Brunelli, A.; Pojana, G.; Marcomini, A.; Nigro, M. Effects of *in Vitro* Exposure to Titanium Dioxide on DNA Integrity of Bottlenose Dolphin (*Tursiops truncatus*) Fibroblasts and Leukocytes. Mar. Environ. Res. **2014**, 100, 66–73.
- Ringwood, A. H.; McCarthy, M.; Bates, T. C.; Carroll, D. L. The Effects of Silver Nanoparticles on Oyster Embryos. *Mar. Environ. Res.* 2009, *69*, 549–551.
- Siller, L.; Lemloh, M. L.; Piticharoenphun, S.; Mendis, B. G.; Horrocks, B. R.; Brümmer, F.; Medaković, D. Silver Nanoparticle Toxicity in Sea Urchin Paracentrotus lividus. Environ. Pollut. 2013, 178, 498–502.
- 57. Manzo, S.; Miglietta, M. L.; Rametta, G.; Buono, S.; Di Francia, G. Embryotoxicity and Spermiotoxicity of

Nanosized ZnO for Mediterranean Sea Urchin *Paracentrotus lividus. J. Haz. Mater.* **2013**, *254*, 1–9.

- Libralato, G.; Minetto, D.; Totano, S.; Mičetić, I.; Pigozzo, A.; Sabbioni, E.; Marcomini, A.; Volpi Ghirardini, A. Embryotoxicity of TiO<sub>2</sub> Nanoparticles to *Mytilus galloprovincialis* (Lmk). *Mar. Environ. Res.* **2013**, *92*, 71–78.
- Canesi, L.; Ciacci, C.; Fabbri, R.; Marcomini, A.; Pojana, G.; Gallo, G. Bivalve Molluscs as a Unique Target Group for Nanoparticle Toxicity. *Mar. Environ. Res.* **2012**, *76*, 16–21.
- Canesi, L.; Prochàzovà, P. The Invertebrate Immune System as a Model for Investigating the Environmental Impact of Nanoparticles. In Nanoparticles and the Immune System; Boraschi, D, Duschl, A, Eds.; Academy Press: Oxford, 2013; pp 91–112.
- Canesi, L.; Ciacci, C.; Betti, M.; Fabbri, R.; Canonico, B.; Fantinati, A.; Marcomini, A.; Pojana, G. Immunotoxicity of Carbon Black Nanoparticles to Blue Mussel Hemocytes. *Environ. Int.* **2008**, *34*, 1114– 1119.
- Canesi, L.; Ciacci, C.; Vallotto, D.; Gallo, G. *In Vitro* Effects of Suspensions of Selected Nanoparticles (C<sub>60</sub> Fullerene, TiO<sub>2</sub>, SiO<sub>2</sub>) on *Mytilus* Emocytes. *Aquat. Toxicol.* **2010**, *96*, 151–158.
- Ciacci, C.; Canonico, B.; Bilanicova, D.; Fabbri, R.; Cortese, K.; Gallo, G.; Marcomini, A.; Pojana, G.; Canesi, L. Immunomodulation by Different Types of N-Oxides in the Hemocytes of the Marine Bivalve *Mytilus galloprovincialis. PLoS One* **2012**, 7, e36937.
- Canesi, L.; Fabbri, R.; Gallo, G.; Vallotto, D.; Marcomini, A.; Pojana, G. Biomarkers in *Mytilus galloprovincialis* Exposed to Suspensions of Selected Nanoparticles (Nano Carbon Black, C<sub>60</sub> Fullerene, Nano-TiO<sub>2</sub>, Nano-SiO<sub>2</sub>). *Aquat. Toxicol.* **2010**, *100*, 168–177.
- Barmo, C.; Ciacci, C.; Canonico, B.; Fabbri, R.; Cortese, K.; Balbi, T.; Marcomini, A.; Pojana, G.; Gallo, G.; Canesi, L. *In vivo* Effects of n-TiO<sub>2</sub> on Digestive Gland and Immune Function of the Marine Bivalve *Mytilus galloprovincialis*. *Aquat. Toxicol.* **2013**, *132–133*, 9–18.
- Venier, P.; Varotto, L.; Rosani, U.; Millino, C.; Celegato, B.; Bernante, F.; Lanfranchi, G.; Novoa, B.; Roch, P.; Figueras, A.; *et al.* Insights into the Innate Immunity of the Mediterranean Mussel Mytilus galloprovincialis. BMC Genomics **2011**, *12*, 69.
- Philipp, E. E.; Kraemer, L.; Melzner, F.; Poustka, A. J.; Thieme, S.; Findeisen, U.; Schreiber, S.; Rosenstiel, P. Massively Parallel RNA Sequencing Identifies a Complex Immune Gene Repertoire in the Lophotrochozoan *Mytilus edulis. PLoS One* **2012**, *7*, e33091.

- Toubiana, M.; Gerdol, M.; Rosani, U.; Pallavicini, A.; Venier, P.; Roch, P. Toll-Like Receptors and MyD88 Adaptors in *Mytilus*: Complete Cds and Gene Expression Levels. *Dev. Comp. Immunol.* 2013, 40, 158–166.
- Balbi, T.; Smerilli, A.; Fabbri, R.; Ciacci, C.; Montagna, M.; Grasselli, E.; Brunelli, A.; Pojana, G.; Marcomini, A.; Gallo, G.; *et al.* Co-Exposure to n-TiO<sub>2</sub> and Cd<sup>2+</sup> Results in Interactive Effects on Biomarker Responses but Not in Increased Toxicity in the Marine Bivalve Mytilus galloprovincialis. Sci. Total Environ. **2014**, 493C, 355–364.
- Rast, J. P.; Smith, L. C.; Loza-Coll, M.; Hibino, T.; Litman, G. W. Genomic Insights into the Immune System of the Sea Urchin. *Science* 2006, 314, 952–956.
- Matranga, V.; Toia, G.; Bonaventura, R.; Muller, W. E. G. Cellular and Biochemical Responses to Environmental and Experimentally Induced Stress in Sea Urchin Coelomocytes. *Cell Stress Chaperones* 2000, *5*, 158–165.
- Matranga, V.; Pinsino, A.; Celi, M.; Natoli, A.; Bonaventura, R.; Schröder, H. C.; Müller, W. E. G. Monitoring Chemical and Physical Stress Using Sea Urchin Immune Cells. *Prog. Mol. Subcell. Biol.* 2005, 39, 85–110.
- 73. Matranga, V.; Pinsino, A.; Celi, M.; Di Bella, G.; Natoli, A. Impacts of UVB Radiation on Short Term Cultures of Sea Urchin Coelomocytes. *Mar. Biol.* **2006**, *149*, 25–34.
- 74. Pinsino, A.; Della Torre, C.; Sammarini, V.; Bonaventura, R.; Amato, E.; Matranga, V. Sea Urchin Coelomocytes as a Novel Cellular Biosensor of Environmental Stress: A Field Study in the Tremiti Island Marine Protected Area, Southern Adriatic Sea, Italy. *Cell Biol. Toxicol.* 2008, 24, 541–552.
- Smith, L. C.; Ghosh, J.; Buckley, K. M.; Clow, L. A.; Dheilly, N. M.; Haug, T.; Henson, J. H.; Li, C.; Lun, C. M.; Majeske, A. J.; *et al.* Echinoderm Immunity. *Adv. Exp. Med. Biol.* **2010**, *708*, 260–301.
- Falugi, C.; Aluigi, M. G.; Chiantore, M. C.; Privitera, D.; Ramoino, P.; Gatti, M. A.; Fabrizi, A.; Pinsino, A.; Matranga, V. Toxicity of Metal Oxide Nanoparticles in Immune Cells of the Sea Urchin. *Mar. Envir*on. Res. **2012**, *76*, 114–121.
- Kodali, V.; Littke, M. H.; Tilton, S. C.; Teeguarden, J. G.; Shi, L.; Frevert, C. W.; Wang, W.; Pounds, J. G.; Thrall, B. D. Dysregulation of Macrophage Activation Profiles by Engineered Nanoparticles. ACS Nano 2013, 7, 6997–7010.
- Boraschi, D.; Costantino, L.; Italiani, P. Interaction of Nanoparticles with Immunocompetent Cells: Nanosafety Considerations. *Nanomedicine* 2012, 7, 121–31.



9707

- Sea Urchin Genome Sequencing Consortium; *et al.* The Genome of the Sea Urchin Strongylocentrotus purpuratus. Science **2006**, 314, 941–952.
- Russo, R.; Bonaventura, R.; Zito, F.; Schröder, H. C.; Müller, I.; Müller, W. E. G.; Matranga, V. Stress to Cadmium Monitored by Metallothionein Gene Induction in *Paracentrotus lividus* Embryos. *Cell Stress Chaperones* **2003**, *8*, 232–241.
- Russo, R.; Zito, F.; Costa, C.; Bonaventura, R.; Matranga, V. Transcriptional Increase and Misexpression of 14-3-3 Epsilon in Sea Urchin Embryos Exposed to UV-B. *Cell Stress Chaperones* **2010**, *15*, 993– 1001.
- Russo, R.; Bonaventura, R.; Matranga, V. Time- and Dose-Dependent Gene Expression in Sea Urchin Embryos Exposed to UVB. *Mar. Environ. Res.* 2014, 93, 85–92.
- Bonaventura, R.; Poma, V.; Costa, C.; Matranga, V. UVB Radiation Prevents Skeleton Growth and Stimulates the Expression of Stress Markers in Sea Urchin Embryos. *Biochem. Biophys. Res. Commun.* 2005, 328, 150–157.
- Bonaventura, R.; Poma, V.; Russo, R.; Zito, F.; Matranga, V. Effects of UV-B Radiation on the Development and Hsp 70 Expression in Sea Urchin Cleavage Embryos. *Mar. Biol.* 2006, 149, 79–86.
- Bonaventura, R.; Zito, F.; Costa, C.; Giarrusso, S.; Celi, F.; Matranga, V. Stress Response Gene Activation Protects Sea Urchin Embryos Exposed to X-rays. *Cell Stress Chaperones* 2011, *16*, 681–687.
- Matranga, V.; Zito, F.; Costa, C.; Bonaventura, R.; Giarrusso, S.; Celi, F. Embryonic Development and Skeletogenic Gene Expression Affected by X-rays in the Mediterranean Sea Urchin *Paracentrotus lividus. Ecotoxicology* **2010**, *19*, 530–537.
- Pinsino, A.; Matranga, V.; Trinchella, F.; Roccheri, M. C. Sea Urchin Embryos as an *in Vivo* Model for the Assessment of Manganese Toxicity: Developmental and Stress Response Effects. *Ecotoxicology* 2010, 19, 555–562.
- Pinsino, A.; Roccheri, M. C.; Costa, C.; Matranga, V. Manganese Interferes with Calcium, Perturbs ERK Signaling, and Produces Embryos with no Skeleton. *Toxicol. Sci.* 2011, 123, 217–230.
- Pinsino, A.; Roccheri, M. C.; Matranga, V. Manganese Overload Affects p38 MAPK Phosphorylation and Metalloproteinase Activity during Sea Urchin Embryonic Development. *Mar. Environ. Res.* **2014**, *93*, 64–69.
- Handy, R. D.; Owen, R.; Valsami-Jones, E. The Ecotoxicology of Nanoparticles and Nanomaterials: Current Status, Knowledge Gaps,

Challenges, and Future Needs. *Ecotoxicology* **2008**, *17*, 315–325.

- Moore, M. N. Do Nanoparticles Present Ecotoxicological Risks for the Health of the Aquatic Environment? *Environ. Int.* 2006, *32*, 967– 976.
- Magdolenova, Z.; Collins, A.; Kumar, A.; Dhawan, A.; Stone, V.; Dusinska, M. Mechanisms of Genotoxicity. A Review of *in Vitro* and *in Vivo* Studies with Engineered Nanoparticles. *Nanotoxicology* 2014, 8, 233–278.
- Frenzilli, G.; Nigro, M.; Lyons, B. P. The Comet Assay for the Evaluation of Genotoxic Impact in Aquatic Environments. *Mutat. Res.* 2009, 681, 80–92.
- 94. Gagné, F.; Auclair, J.; Turcotte, P.; Fournier, M.; Gagnon, C.; Sauvé, S.; Blaise, C. Ecotoxicity of CdTe Quantum Dots to Freshwater Mussels: Impacts on Immune System, Oxidative Stress and Genotoxicity. Aquat. Toxicol. **2008**, *86*, 333–340.
- Kadar, E.; Tarran, G. A.; Jha, A. N.; Al-Subiai, S. N. Stabilization of Engineered Zero-Valent Nanoiron with Na-Acrylic Copolymer Enhances Spermiotoxicity. *Environ. Sci. Technol.* 2011, 45, 3245–3251.
- Gomes, T.; Araújo, O.; Pereira, R.; Almeida, A. C.; Cravo, A.; Bebianno, M. J. Genotoxicity of Copper Oxide and Silver Nanoparticles in the Mussel Mytilus galloprovincialis. Mar. Environ. Res. 2013, 84, 51–59.
- Munari, M.; Sturve, J.; Frenzilli, G.; Sanders, M. B.; Christian, P.; Nigro, M.; Lyons, B. P. Genotoxic Effects of Ag<sub>2</sub>S and CdS Nanoparticles in Blue Mussel (*Mytilus edulis*) Haemocytes. *Chem. Ecol.* 2014, 10.1080/02757540.2014.894989.
- Lovern, S. B.; Klaper, R. Daphnia magna Mortality when Exposed to Titanium Dioxide and Fullerene (C60) Nanoparticles. Environ. Toxicol. Chem. 2006, 2, 1132–1137.
- Galloway, T.; Lewis, C.; Dolciotti, I.; Johnston, B. D.; Moger, J.; Regoli, F. Sublethal Toxicity of Nano-Titanium Dioxide and Carbon Nanotubes in a Sediment Dwelling Marine Polychaete. *Environ. Pollut.* 2010, *158*, 1748–1755.
- Zhu, X.; Zhou, J.; Cai, Z. The Toxicity and Oxidative Stress of TiO<sub>2</sub> Nanoparticles in Marine Abalone (*Haliotis diversicolor supertexta*). Mar. Pollut. Bull. 2011, 63, 334–338.
- D'Agata, A.; Fasulo, S.; Dallas, L. J.; Fisher, A. S.; Maisano, M.; Readman, J. W.; Jha, A. N. Enhanced Toxicity of 'Bulk' Titanium Dioxide Compared to 'Fresh' and 'Aged' Nano-TiO<sub>2</sub> in Marine Mussels (*Mytilus galloprovincialis*). Nanotoxicology **2014**, 8, 549–558.
- Al-Subiai, S. N.; Arlt, V. M.; Frickers, P. E.; Readman, J. W.; Stolpe, B.; Lead, J. R.; Moody, A. J.; Jha, A. N. Merging Nano-Genotoxicology with Eco-Enotoxicology: An Integrated

Approach to Determine Interactive Genotoxic and Sub-Lethal Toxic Effects of C(60) Fullerenes and Fluoranthene in Marine Mussels *Mytilus* sp. *Mutat. Res.* **2012**, *745*, 92–103.

- Canesi, L.; Frenzilli, G.; Balbi, T.; Bernardeschi, M.; Ciacci, C.; Corsolini, S.; Della Torre, C.; Fabbri, R.; Faleri, C.; Focardi, S.; et al. Interactive Effects of n-TiO<sub>2</sub> and 2,3,7,8-TCDD on the Marine Bivalve Mytilus galloprovincialis. Aquat. Toxicol. **2014**, 153, 53–65.
- 104. Goldstone, J. V.; Hamdoun, A.; Cole, B. J.; Howard-Asbhy, M.; Nebert, D. W.; Scally, M.; Dean, M.; Epel, D.; Hahn, M. E.; Stegeman, J. J. The Chemical Defensome: Environmental Sensing and Response Genes in the *Strongylocentrotus purpuratus* Genome. *Dev. Biol.* **2006**, 300, 366–384.
- 105. Smital, T.; Luckenbach, T.; Sauerborn, R.; Hamdoun, A.; Vega, R.; Epel, D. Emerging Contaminants, PPCPs, Pesticides and Microbial Degradation Products as Inhibitors of the Multixenobiotic Defense in Aquatic Organisms. *Mut. Res.* 2004, 552, 101–117.
- Epel, D. Use of Multidrug Transporters as First Line of Defense Against Toxins in Aquatic Organisms. Comp. Biochem. Physiol. A 1998, 120, 23–28.
- 107. Chen, J.; He, Q.; Gao, Y.; Shi, J.; Li, Y. Mesoporous Silica Nanoparticles Loading Doxorubicin Reverse Multidrug Resistance: Performance and Mechanism. *Nanoscale* **2011**, 3, 4314–4322.
- Koyuco, J. C.; Liu, Y.; Tan, B.-J.; Ciu, G. N. C. Functionalyzed Carbon Nanomaterials: Exploring the Interactions with Caco-2 Cells for Potential Oral Drug Delivery. *Int. J. Nanomed.* 2011, 6, 2253–2263.
- Kirtane, A. R.; Kalscheuer, S. M.; Panyam, J. Exploiting Nanotechnology to Overcome Tumor Drug Resistance: Challenges and Opportunities. Adv. Drug Delivery Rev. 2013, 65, 1731–1747.
- 110. Amiard, J. C.; Amiard-Triquet, C.; Barka, S.; Pellerin, J.; Rainbow, P. S. Metallothioneins in Aquatic Invertebrates: Their Role in Metal Detoxification and their Use as Biomarkers. Aquat. Toxicol. 2006, 76, 160–202.
- 111. García-Alonso, J.; Khan, F. R.; Misra, S. K.; Turmaine, M.; Smith, B. D.; Rainbow, P. S.; Luoma, S. N.; Valsami-Jones, E. Cellular Internalization of Silver Nanoparticles in Gut Epithelia of the Estuarine Polychaete Nereis diversicolor. Environ. Sci. Technol. 2011, 45, 4630–4636.
- 112. Pan, J.; Buffet, P.; Poirier, L.; Amiard-Triquet, C.; Gilliland, D.; Joubert, Y.; Pilet, P.; Guibbolini, M.; Risso de Faverney, C.; Roméo, M.; et al. Size Dependent Bioaccumulation and Ecotoxicity of Gold Nanoparticles in an Endobenthic Invertebrate: The Tellinid Clam Scrobicularia plana. Env. Pollut. **2012**, *168*, 37–43.



- Gomes, T.; Pinheiro, J. P.; Cancio, I.; Pereira, C. G.; Cardoso, C.; Bebianno, M. J. Effects of Copper Nanoparticles Exposure in the Mussel *Mytilus* galloprovincialis. Environ. Sci. Technol. **2011**, 45, 9356–9362.
- Gomes, T.; Pereira, C. G.; Cardoso, C.; Pinheiro, J. P.; Cancio, I.; Bebianno, M. J. Accumulation and Toxicity of Copper Oxide Nanoparticles in the Digestive Gland of *Mytilus galloprovincialis. Aquat. Toxicol.* 2012, 118–119, 72–79.
- 115. Buffet, P. E.; Tankoua, O. F.; Pan, J. F.; Berhanu, D.; Herrenknecht, C.; Poirier, L.; Amiard-Triquet, C.; Amiard, J. C.; Bérard, J. B.; Risso, C.; et al. Behavioural and Biochemical Responses of Two Marine Invertebrates Scrobicularia plana and Hediste diversicolor to Copper Oxide Nanoparticles. Chemosphere 2011, 84, 166–174.
- 116. Buffet, P. E.; Richard, M.; Caupos, F.; Vergnoux, A.; Perrein-Ettajani, H.; Luna-Acosta, A.; Mouneyrac, C. A Mesocosm Study of Fate and Effects of CuO Nanoparticles on Endobenthic Species (*Scrobicularia plana*, *Hediste diversicolor*). Environ. *Sci. Technol.* **2013**, *47*, 1620–1628.
- 117. Günther, V.; Lindert, U.; Schaffner, W. The Taste of Heavy Metals: Gene Regulation by MTF-1. *Biochim. Biophys. Acta* **2012**, *1823*, 1416–1425.
- Liu, H. H.; Cohen, Y. Multimedia Environmental Distribution of Engineered Nanomaterials. *Environ. Sci. Technol.* 2014, 48, 3281–3292.
- Cohen, Y.; Rallo, R.; Liu, R.; Liu, H. H. In Silico Analysis of Nanomaterials Hazard and Risk. Acc. Chem. Res. 2012, 46, 802–812.
- Decho, A. W. Microbial Exopolymer Secretions in Ocean Environments: Their Role(s) in Food Webs and Marine Processes. Ocean. Mar. Biol. Annu. Rev. 1990, 28, 73–153.
- Nowack, B.; Bucheli, T. D. Occurrence, Behavior, and Effects of Nanoparticles in the Environment. *Environ. Pollut.* 2007, 150, 5–22.
- Miao, A.-J.; Schwehr, K. A.; Xu, C.; Zhang, S.-J.; Luo, Z.; Quigg, A.; Santschi, P. H. The Algal Toxicity of Silver Engineered Nanoparticles and Detoxification by Exopolymeric Substances. *Environ. Pollut.* **2009**, 157, 3034–3041.
- Lenihan, H. S.; Peterson, C. H.; Byers, J. E.; Grabowski, J. H.; Thayer, G. W.; Colby, D. R. Cascading of Habitat Degradation: Oyster Reefs Invaded by Refugee Fishes Escaping Stress. *Ecol. Appl.* 2001, *11*, 764–782.
- 124. Miller, R. J.; Lenihan, H. S.; Muller, E. B.; Tseng, N.; Keller, A. A. Impacts of Metal Oxide Nanoparticles on Marine Phytoplankton. *Environ. Sci. Technol.* **2010**, *44*, 7329–7334.
- Hanna, S. K.; Miller, R. J.; Muller, E. B.; Nisbet, R. M.; Lenihan, H. S. Impact of Zinc Oxide Nanoparticles on the Individual Performance of *Mytilus*

galloprovincialis. PLoS One **2014**, 10.1371/journal.pone.0061800.

- Muller, E. B.; Hanna, S. K.; Miller, R. J.; Lenihan, H. S.; Nisbet, R. M. Impact of Engineered Zinc Oxide Nanoparticles on the Energy Budgets of *Mytilus galloprovincialis. J. Sea Res.* 2014, 10.1016/j.seares.2013.12.013.
- 127. Nel, A. E.; Nasser, E.; Godwin, H.; Avery, D.; Bahadori, T.; Bergeson, L.; Beryt, E.; Bonner, J. C.; Boverhof, D.; Carter, J.; Castranova, V.; DeShazo, J. R.; Hussain, S. M.; Kane, A. B.; Klaessig, F. F.; Kuempel, E.; Lanfranconi, M.; Landsiedel, R.; Malloy, T.; Miller, M. B.; et al. A Multi-Stakeholder Perspective on the Use of Alternative Test Strategies for Nanomaterial Safety Assessment. ACS Nano 2013, 7, 6422–6433.
- 128. Elimelech, M. N.; Steinfeldt, M.; von Gleich, A.; Petschow, U.; Haum, R. Hazards and Resource Efficiency: A Three-Tiered Approach to Assessing the Implications of Nanotechnology and Influencing its Development. J. Ind. Ecol. 2008, 12, 493–493.
- Mudunkotuwa, I. A.; Pettibone, J. M.; Grassian, V. H. Environmental Implications of Nanoparticle Aging in the Processing and Fate of Copper-Based Nanomaterials. *Environ. Sci. Technol.* 2012, *46*, 7001–7010.
- Fouqueray, M.; Dufils, B.; Vollat, B.; Chaurand, P.; Botta, C.; Abacci, K.; Labille, J.; Rose, J.; Garric, J. Effects of Aged TiO<sub>2</sub> Nanomaterial from Sunscreen on *Daphnia magna* Exposed by Dietary Route. *Environ. Pollut.* **2012**, *163*, 55–61.
- 131. Pluskota, A.; Horzowski, E.; Bossinger, O.; von Mikecz, A. In *Caenorhabditis elegans* Nanoparticle-Bio-Interactions Become Transparent: Silica-Nanoparticles Induce Reproductive Senescence. *PLoS One* **2009**, *4*, e6622.
- 132. Hristozov, D. R.; Gottardo, S.; Critto, A.; Marcomini, A. Risk Assessment of Engineered Nanomaterials: A Review of Available Data and Approaches from a Regulatory Perspective. Nanotoxicology 2012, 6, 880–898.
- Deliyanni, E. A.; Bakoyannakis, D. N.; Zouboulis, A. I.; Matis, K. A. Sorption of As(V) lons by Akaganeite-Type Nanocrystals. *Chemosphere* 2003, 50, 155–163.
- Lopez, X.; Castaño, V. M. Chromium Removal from Industrial Water Through Functionalized Nanoparticles. J. Nanosci. Nanotechnol. 2009, 8, 5733–5738.
- Pacheco, S.; Medina, M.; Valencia, F.; Tapia, J. Removal of Inorganic Mercury from Polluted Water Using Structured Nanoparticles. J. Environ. Eng. 2006, 132, 342–349.
- Lisha, K. P.; Anshup, P. T. Towards a Practical Solution for Removing Inorganic Mercury from Drinking Water Using Gold Nanoparticles. *Gold Bull.* 2009, 42, 144–152.

- 137. Ojea-Jimenez, I.; Lopez, X.; Arbiol, J.; Puntes, V. Citrate-Coated Gold Nanoparticles as Smart Scavengers for Mercury(II) Removal from Polluted Waters. *ACS Nano* **2012**, *6*, 2253–2260.
- Sanchez, A.; Recillas, S.; Font, X.; Casals, E.; Gonzalez, E.; Puntes, V. Ecotoxicity of, and Remediation with, Engineered Inorganic Nanoparticles in the Environment. *TrAC*, *Trends Anal. Chem.* **2011**, *30*, 507– 516.
- 139. Diallo, M. S.; Savage, N. Nanoparticles and Water Quality. *J. Nanoparticle Res.* **2005**, *7*, 325–330.
- Savage, N.; Diallo, M. S. Nanomaterials and Water Purification: Opportunities and Challenges. J. Nanoparticle Res. 2005, 7, 331–342.
- Stuart, J. E.; Rees, N. V.; Cullenb, J. T.; Compton, R. G. Direct Electrochemical Detection and Sizing of Silver Nanoparticles in Seawater Media. *Nanoscale* **2013**, *5*, 174–177.
- 142. Thomas, C. R.; George, S.; Horst, A.; Ji, Z.; Miller, R. J.; Peralta-Videa, J. R.; Xia, T.; Porkhel, S.; Madler, L.; Gardea-Torresdey, J. L.; Holden, P.; Keller, A. A.; Lenihan, H. S.; Nel, A. E.; Zink, J. I. Nanomaterials in the Environment: From Materials to High-Throughtput Screening to Organisms. ACS Nano 2011, 5, 13–20.
- 143. Holden, P. R.; Nisbet, R. M.; Lenihan, H. S.; Miller, R. J.; Cherr, G.; Schimel, J.; Gardea-Torresdey, J. Ecological Nanotoxicology: Nanomaterial Hazard Considerations at the Subcellular, Population, Community, and Ecosystem levels. Acc. Chem. Res. 2013, 46, 812–822.
- 144. Bleeker, E. A.; de Jong, W. H.; Geertsma, R. E.; Groenewold, M.; Heugens, E. H.; Koers-Jacquemijns, M.; van de Meent, D.; Popma, J. R.; Rietveld, A. G.; Wijnhoven, S. W. Considerations on The EU Definition of a Nanomaterial: Science to Support Policy Making. *Regul. Toxicol. Pharmacol.* **2013**, *65*, 119–125.

