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# Mitochondria: the gatekeepers between metabolism and immunity

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Metabolism and immunity are crucial monitors of the whole-body homeodynamics. All cells require energy to perform their basic functions. One of the most important metabolic skills of the cell is the ability to optimally adapt metabolism according to demand or availability, known as metabolic flexibility. The immune cells, first line of host defense that circulate in the body and migrate between tissues, need to function also in environments in which nutrients are not always available. The resilience of immune cells consists precisely in their high adaptive capacity, a challenge that arises especially in the framework of sustained immune responses. Pubmed and Scopus databases were consulted to construct the extensive background explored in this review, from the Kennedy and Lehninger studies on mitochondrial biochemistry of the 1950s to the most recent findings on immunometabolism. In detail, we first focus on how metabolic reconfiguration influences the action steps of the immune system and modulates immune cell fate and function. Then, we highlighted the evidence for considering mitochondria, besides conventional cellular energy suppliers, as the powerhouses of immunometabolism. Finally, we explored the main immunometabolic hubs in the organism emphasizing in them the reciprocal impact between metabolic and immune components in both physiological and pathological conditions.

#### KEYWORDS

immunometabolism, metabolic flexibility, mitochondrial function, mitochondrial dynamics, metabolic reprogramming

**Abbreviations:** ATP, Adenosine triphosphate; PGC1, peroxisome proliferator-activated receptor gamma coactivator 1 alpha; STAT6, signal transducer and activator of transcription 6; OXPHOS, oxidative phosphorylation; TCA, tricarboxylic acid; mTOR, mechanistic target of rapamycin; SREBPs, sterol regulatory binding element proteins; HIF-1 $\alpha$ , hypoxia inducible factor-1 $\alpha$ ; FAO, fatty acid oxidation; DAMPs, damage-associated molecular patterns; TFAM, mitochondrial transcription factor A; mtROS, mitochondrial ROS; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; MAVS, mitochondrial antiviral signaling protein; MFNs, mitofusins; OPA1, Optic-atrophy-1; Drp1, Dynamin-related protein-1; NET, neutrophil extracellular traps; ETC, electron transport chain; FFAs, free fatty acids; LSECs, liver sinusoidal endothelial cells; IFN-I, I interferon; PBMCs, Peripheral blood mononuclear cells; SCFAs, short chain fatty acids.



# **1** Introduction

The rules of human survival teach us that: i) we cannot be unarmed against the pathogen attack and ii) nothing survives without nourishment. The "immunometabolism" defines the portal between immunology and metabolism, two trials which our organism trusts to maintain a state of wellness. These processes are inextricably linked and the interfaces between the immune and metabolic systems mediate the whole-body homeodynamics. The crosstalk between these two major balancers of the body health has multiple facets (1). The immune system continually perceives and reacts to pathogenic or environmental dangers with secretion of cytokines, chemokines, and inflammatory mediators by the innate immune cells, and with the proliferation of adaptive immune cells. These processes are bioenergetically expensive and need an accurate control of cellular metabolic pathways (2). The immune response requires the reallocation of nutrients within immune cells in order to: provide the substrates for ATP production serving to sustain the functions of activated immune cells; and build blocks for the production of necessary macromolecules for the proliferation of immune cells. The cellular metabolic reprogramming, that help to regulate specific immune cell functions, is an aspect of immunometabolic research which has already been to some extent explored (3). Indeed, the concept that metabolism influences cellular functions and fate may seem obvious, but taking this step backwards is the appropriate approach for a detailed understanding of the immunometabolic mechanisms, consequently useful to design effective strategies to ensure the health of the organism. The feature of this review is the establishment of a thread which examines the several facets of the immunometabolism, starting from the intersection mechanisms of

the mitochondrial metabolism with the functionality of the immune cells. Then, we highlighted the mitochondrial dynamics processes in the activation phases of immune cells, and the immunometabolic regulation in different organs and tissue in both physiological and pathological conditions.

# 2 Metabolic adaptations of immune cells

The cells of the immune system are arguably the most dynamic components of our organism, they need to function in different contexts, including those where the availability of nutrients is restricted or compromised (4). Immune cells possess a broad set of skills ranging from being sleeping sentinels to inducing clonal expansion, modulating surface receptor expression and secreting large amounts of effector molecules (5, 6). The performance of these distinct functional activities is tightly dependent by the metabolic flexibility of these cells (7). Indeed, recent findings have demonstrated that peripheral immune cells can adapt to environmental shifts by metabolizing alternative non- glucose substrates, such as amino acids or fatty acids (8-15). A system in which nutritional and energy inputs are properly processed and substrate utilization is properly regulated is defined as a metabolically sensitive and flexible system. The mitochondrial machinery is responsible for switches in the oxidation of substrates, and the choices are orchestrated by an intricate network of cell signaling events. This metabolic flexibility enables peripheral immune cells to perform a multitude of functions in disrupted environments where the availability of carbon sources varies (16).

In the 20<sup>th</sup> century, Warburg was the first to launch the immunometabolic research describing the metabolic changes and aerobic glycolysis in cancer cells (17). Recent research has highlighted that each subset of immune cells has a different metabolic control and nutrient utilization. Vats et al. highlighted the molecular pathway that directly links mitochondrial oxidative metabolism to the anti-inflammatory program of macrophage activation, identifying peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC1a) and signal transducer and activator of transcription 6 (STAT6) as metabolic regulators that can control macrophage activation (18). To date, increasingly detailed experiments have shown that naïve T and B cells dynamically switch their metabolic programs upon activation. Several studies highlighted signaling and transcriptional networks that regulate metabolic state to tune the management of T cells fate (19). The results converge on the increase of the mitochondrial metabolism and oxidative phosphorylation (OXPHOS) (20-22) and on the induction of anabolic metabolism in T cells, specifically, evidencing that this occurs during the period of quiescence exit in parallel with higher glucose uptake and lactate production (23). Glutaminolysis is an important pathway in these cells, given that glutamine replenishes tricarboxylic acid (TCA) cycle intermediates as they are picked up for biosynthesis (15). These events are mediated by the mechanistic target of rapamycin (mTOR)associated signaling, in part, by upregulating transcriptional programs mediated by MYC (24), sterol regulatory binding element proteins (SREBPs) (25), and hypoxia inducible factor- $1\alpha$ (HIF-1 $\alpha$ ) (26–28). T<sub>reg</sub> cells and memory T cells have a dissimilar metabolism from their activated effector T cells counterparts, indeed, for their development and persistence depend on mitochondrial fatty acid oxidation (FAO) rather than aerobic glycolysis (29).

The interaction between nutrients, metabolic programs and signaling pathways has recently well described also in naïve B cell activation, differentiation and fueling of the antibody secretion machinery. Several studies have found that glucose uptake is increased upon B cells activation (30-33). It was also demonstrated that the initial B cell activation induces metabolic reprogramming, with increased glucose uptake without accumulation of glycolytic metabolites, suggesting that glucose is fluxing through the glycolytic pathway and is probably directed to alternative metabolic pathways in activated B cells. Indeed, stimulated B cells increase programs for OXPHOS, the TCA cycle, and nucleotide biosynthesis (34). The signaling and nutrient sensitive mechanisms that mediate B cell activation and differentiation and the function of energetic and biosynthetic pathways were widely reviewed (35). Several experiments have shown that the functional destination (tolerance, effector or regulatory activities) of B lymphocytes dictates the choice of their metabolic programming, also depending on the receptors and the co-activation molecules stimulated (36). Indeed, glucose restriction did not affect B cell functions, whereas OXPHOS inhibition or glutamine restriction significantly impaired B cell growth and differentiation (34). Metabolic restrictions (such as low ATP reserve and mitochondrial mass, or transcriptional repression of glucose transport and limited activity of the pentose phosphate pathway) provide a safeguard against autoreactive or premalignant B cells (37). This may happen through hyperactivation-induced metabolic stress, such as ATP deprivation and oxidative damage. Meanwhile, prolonged exposure to nutrient oversupply subverts B cell gatekeeper functions, promotes malignant B cell transformation and progression of autoimmune disease (38).

Notable, the metabolic availability influences immune cell fate decisions also through its impact on the epigenome (39, 40). Indeed, several metabolic checkpoints exist to limit epigenetic instability and restrain B cell development (7). Compelling evidence demonstrated that metabolic intermediates are ideal signaling mediators. Their levels are in dynamic equilibrium with systemic, microenvironmental and cell-intrinsic cues, whereby fluctuations inform cell fate decisions (41). They can both inform the fitness of extracellular conditions and integrate this sensing into the epigenome, serving as cofactors for chromatin remodeling enzymes (42). Metabolic-dependent epigenetic reprogramming might explain why changes in cellular metabolism are crucial for multi-stage B cell specification. Lastly, the metabolic state influences immune cell function not only through epigenetic remodeling but also through a restructuration of intracellular architecture, this will be discussed in more detail below in a dedicated section.

# 3 Immunometabolic skills of the mitochondria

In recent years, the scientific research has produced new striking knowledge of mitochondrial function in metabolism, leading to consider the mitochondria as targets for the development of new therapeutic approaches. Alongside the paradigm widely described in biochemistry textbooks, which defines these organelles as 'energy powerhouses' of the cell, a new image has emerged of the mitochondria as a 'Pandora's box', an intracellular 'container' crucial not only for the life but also for the cell death (43, 44). For this reason, it is essential to consider their expertise in immunometabolic management.

Mitochondria represent the most ancient endomembrane system in eukaryotic cells. They arose around two thousand million years ago and, over the years, mitochondria have continuously demonstrated their autonomy and ability (45). In 1907, they were defined as cellular organelles responsible for the functions of respiration and energy production (46). Around the 1950s, Kennedy and Lehninger discovered that the TCA, FAO and OXPHOS take place in the mitochondria (47). In 1967, Margulis revived the long-forgotten endosymbiont theory on the origin of organelles (48). Since the 1970s, the mechanism of mitochondrial biogenesis has been elucidated and it is recognized that mitochondria are semiautonomous organelles, capable of synthesizing 5% of the proteins they require and importing the rest from cytoplasmic synthesis (49). Morphologically, like their bacterial ancestor, mitochondria consist of two separate and functionally distinct outer and inner membranes that enclose the intermembrane space and matrix compartments. The architecture of mitochondria is essential for their proper functioning and also for the containment of immunogenic molecules derived from mitochondria (50, 51). They also contain the mitochondrial

DNA (mtDNA), a circular genome, which has been reduced in the course of evolution through gene transfer to the nucleus. The bacterial-like characteristics of mitochondria also reinforce the idea that they are hubs of immunity (52). The proteins present in mitochondria are structurally similar to those in bacteria and allow them to be recognized by the same receptors as the immune system (53). To date, as already hypothesized by Altmann in 1890, the main role of mitochondria is to provide metabolic energy in all eukaryotic cells (54). However, these organelles orchestrate mechanisms which directly impact cell fate and fitness, so to consider them trivially 'powerhouses of cells' would be limiting, to say the least. Indeed, it is well known that the metabolic functions of mitochondria reach far beyond bioenergetics. Additionally to their exclusive ability to carry out the OXPHOS, these organelles participate in intermediary metabolism, regulate programmed cell death, calcium homeostasis, and control the production of reactive oxygen species (ROS) (55-57).

A proper mitochondrial functionality is fundamental for the cellular homeostasis. In this regard, several molecules extruded from mitochondria alert neighboring cells, the immune system (58), and the producing cell itself about mitochondrial dysfunction (59). Several studies demonstrated that mitochondrial ROS also contribute to adaptive stress signaling pathways, such as hypoxia and control cell proliferation and differentiation (60, 61). Likewise, the levels of nitric oxide (NO), another by-products of mitochondrial respiratory activity, act as initiators through which mitochondria modulate signal transduction pathways implicated in the induction of cellular defense mechanisms and adaptive responses (62). Mitochondria are also the source of molecules, including proteins, lipids, metabolites and mtDNA, collectively named damage-associated molecular patterns (DAMPs). These DAMPs are endogenous danger molecules that are released from damaged or dying cells and activate the innate immune system by interacting with pattern recognition receptors. The DAMPs, when imbalanced, employ immunogenic capacity in immune and nonimmune cells (63). ATP, succinate, cardiolipin, N-formyl peptides (NFPs), mtDNA and mitochondrial transcription factor A (TFAM), are examples of DAMPs that serve as danger flags for immunological signaling (63). The secretion of succinate triggers pro-inflammatory differentiation of T-lymphocytes (64) and have synergic effects with Toll-like receptors ligands in dendritic cells for the production of cytokines. The succinate is a regulator of inflammation, in M1 macrophages due to a break point of Krebs cycle it was observed its accumulation, and demonstrated a prominent proinflammatory activity and roles in immunity (65). High levels of extracellular ATP signals induces release of proinflammatory cytokines, inflammasome activation (66), neutrophils degranulation, apoptosis and ROS release through P2X receptors (67, 68). The exposition of cardiolipin to the extracellular media is associated with increased apoptosis and autophagy: the cardiolipin can bind directly to Nod-like receptor 3 (NLRP3) and activate inflammasome-mediated immune response (69). Moreover, cardiolipin externalization to the outer mitochondrial membrane acts as an elimination signal for mitophagy in mammalian cells (70), this process is facilitated by the activated Gasdermin D permeabilization of mitochondrial membranes that cause rapid, cardiolipin-dependent mitochondrial destruction (71). N-formyl

peptides are extruded by the mitochondria of damaged or dying cells, they act as chemoattractant of neutrophils via formyl-peptides receptors (72). Extra mitochondrial mtDNA has been widely shown to induce a proinflammatory state (73, 74), its binding to TLR9 induces proinflammatory cytokines production, chemotaxis and phagocytic activation via a MyD88-dependent signaling cascade (75). TFAM enhances the immunogenicity of mtDNA (76), TFAM is recognized by the receptor for advanced glycation end products, which guides TFAM-mtDNA complexes to the endosomal pathway, also, TFAM enhances cytokine secretion in combination with NFPs (77). mtDNA may also amplify the activation of NLRP3 by mitochondrial ROS (mtROS) (78). ROS are a consequence of mitochondrial disruption in both M1 macrophages (40) and effector T cells and control adaptive immune-cell activation. It is noteworthy that ROS are also guiding signals in the production of inflammatory cytokines. T and B cells require ROS production to trigger an adequate immune response. T cell activation induces a spike in mtROS production, and blockade this process neutralizes IL-2 production by T cells (79). B cell activation is also managed by mtROS. The activation of B cell-surface receptors stimulates in turn the calcium release into the cytoplasm, which promotes ROS production, this cooperative interaction acts in a feedback manner to amplify the early signal generated (80). An interesting research showed that isolated human monocytes exposure to mtDAMPs generated significantly less interleukins IL-1β, IL-6, IL-12-p70 and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) upon lipopolysaccharide challenge when compared to their untreated counterparts, leading to speculate to the induction of a transient state in which these cells are refractory to further endotoxin stimulation (81). Further studies will be crucial to understanding the role of this phenomenon, that could be the root of the onset of noncommunicable chronic diseases, typified by mitochondrial dysfunction and disruption of the immune system. Recently, the intermediate role of mitochondria in toll-like receptor-mediated innate immune responses and in the activation of the NLRP3 inflammasome complex has highlighted, supporting the striking functions of mitochondria in innate immunity (82). Indeed, besides being DAMPs producers, mitochondria are also linked to immunity through their role as innate immune platforms that host the mitochondrial antiviral signaling protein (MAVS) as a viral RNA sensor and the inflammasome NLRP3 as a multiple immunogenic receptor (63).

Understanding how non-immune cells respond to DAMPs released following mitochondrial harm and the mechanisms implicated in these responses are among the main targets of recent researches (63). Understanding the conditions under which damage to non-immune cells leads to chronic and systemic inflammatory responses is relevant. This is discussed in a later section.

# 4 Mitochondrial dynamics drive the immunometabolic pathways

The multifaceted contributions of mitochondria to cell metabolism as bioenergetic powerhouses, biosynthetic centers, ROS production managers and waste management hubs is undisputed (83). Unequivocally, mitochondria perform a plethora of cellular functions besides energy production (84). It is equally clear that the fate and function of innate and adaptive immune cells depends crucially on mitochondrial bioenergetics (58). Exciting evidence demonstrated that mitochondria constantly change their morphology depending on the cell's metabolic requirements, highlighting reciprocal crosstalk between mitochondrial dynamics and metabolism (85, 86).

Mitochondrial dynamics refers to the formation of a dynamic network involving a continuous alternation of fusion and fission/ division processes in order to maintain their cellular abundance, morphology, quality and function control (87). Mitochondrial fusion and division typically counterbalance each other. Three proteins that control mitochondrial fusion and division have identified: i) mitofusins (MFNs) (outer mitochondrial membrane fusion), ii) Optic-atrophy-1 (OPA1)/mitochondria genome maintenance 1 (inner mitochondrial membrane fusion), and iii) Dynamin-related protein-1 (DRP1)/Dynamin-1 (division of outer and inner mitochondrial membranes) (88). Mitochondrial fusion is the physical merging of the mitochondrial membranes of two originally distinct mitochondria. Mitochondrial division is the separation of a single organelle into two or more independent structures. These two active and combined effects originate the mitochondrial networks. Several studies performed in metabolic tissues, such as the liver, the skeletal muscle and the central nervous system demonstrated that the unbalance in mitochondrial fusion/fission dynamics cause cell and tissue dysfunction and altered metabolic homeodynamics (89-92). A decade ago, it was already proven that deletion of any of the dynamics machinery perturbs OXPHOS and glycolytic rates at baseline (93). Tissue-specific deletion of mitofusin-2 (Mfn2) in muscles of mice disrupts glucose homeostasis (94), and Drp1 ablation in the liver results in reduced adiposity and elevated whole-body energy expenditure, protecting mice from diet-induced obesity (95). The evidence that the alterations in fusion/fission machinery alter the mitochondrial function and with it the cell function holds true across various tissues, including the immune system. Indeed, mitochondrial dynamics are a critical control point also for immune cell function (96).

In several immune cells, including neutrophils, macrophages, mast cells, and T- and B-cells, mitochondria adapt specific mitochondrial morphologies according to the cellular activation state (78, 97, 98). In LPS-activated macrophages the inhibition of mitochondrial fission, through the Drp1 inhibitor Mdivi-1, reduce glycolytic reprogramming that these cells implement to achieve polarization into a proinflammatory M1 state (97). The mitochondrial dynamics impact also on the role of the mitochondrial membrane as a signaling platform. Indeed, the depletion of Mfn1/2 or Opa1 reduced MAVS-driven innate antiviral signaling in a mitochondrial membrane potentialdependent manner (99, 100). In neutrophils, the mitochondrial fusion is implicated in the formation of neutrophil extracellular traps (NET), the deletion of OPA lead to a decrease of ATP levels which is fundamental for microtubule network assembly and NET formation (101). Also in human mast cell immune response was investigated the role of mitochondrial dynamics revealing that degranulation processes and secretion of preformed TNF are regulated by Drp1 activation (98). The different roles of Tlymphocytes as T effectors (T<sub>e</sub>) and T memory (T<sub>m</sub>) cells impose them large changes in ATP demand and nutrient utilization. Te cells promote aerobic glycolysis to sustain anabolic pathways of metabolism, while T<sub>m</sub> cells engage catabolic pathways, like FAO, and these metabolic differences are reflected in mitochondrial morphology (96). Memory T cells have more fused mitochondrial networks suggesting a requirement for mitochondrial fusion in memory T cell metabolism and homeostasis. Indeed, the fusion protein OPA1 is required for tight cristae organization in T<sub>m</sub> cells, facilitating efficient electron transport chain (ETC) activity and favorable redox balance, its deletion caused defects in T<sub>m</sub> survival (97). While, more fragmented mitochondria (fission process, with low expression of the fusion proteins MFN2 and OPA1 and high levels of active DRP1) were observed in Te cells, leading to punctate mitochondria, cristae expansion and reduced ETC efficiency which promote aerobic glycolysis (97, 102). In T cells the processes of IL-2 production and immune synapse formation are dependent on mitochondrial fission (103, 104). Indeed, activated T cells show an increase in the production of mtROS, required in the activation of the transcription factor NF-kB, which transcribes IL-2. Inhibition of DRP1 by Mdivi-1 reduced IL-2 mRNA levels and T-cell proliferation (96).

Mitochondrial fission also occurs during B-cell activation, while naïve B cells have predominantly elongated mitochondria. Activated B cells increase glucose uptake, TCA cycle and OXPHOS and have fragmented mitochondria, while naïve B cells maintain a predominance of elongated mitochondria (30, 34). These findings are similar to those noted in T cells; however, it was found that naïve B cells have significantly fewer mitochondria in comparison to naïve T cells (96, 105). It seems that B cells use the mitochondrial remodeling as a key mechanism to control the optimal function of these few mitochondria to compensate this restriction in the number and volume of mitochondria and ATP reserves (106). B cells predominantly favor mitochondria fission and thus house smaller, less functional mitochondria with limited capacity for oxygen consumption and ATP production (34, 107). However, a recent study demonstrated that T cell-dependent activation of murine B cells not only temporarily increased metabolic activity (e.g., glucose uptake and glutamine consumption) but also increased mitochondria number through fission in the absence of mtDNA replication (34).

These exciting evidence highlight that mitochondria are tightly interlaced with metabolic and immune cell homeostasis, it follows that the proper function of mitochondria is crucial to ensuring the health of the organism. From now on, talking about mitochondrialdriven immunometabolic homeostasis would not be a hazard.

# 5 Immunometabolism in health and disease: the main immunometabolic hubs in the organism

Metabolic homeostasis and immune function are pivotal requirements at the root of systemic health monitoring. The crosstalk between immune and metabolic processes is coordinated by communication circuits between specialized tissue-resident cells and organs that include messenger molecules such as hormones, neurotrophic peptides, cytokines, and metabolites (108–112). The intervention of these components of systemic immunometabolism underlie the impact of the metabolism on systemic inflammation and vice versa, both in health and in disease (Figure 1).

Nutritional habits are key determinants of body composition and systemic metabolism (113). Meantime, it is well known that malnutrition leads to inflammation and influences systemic immune responses (114, 115). The unbalance between the body energetic management (metabolism) and the ability to defend itself against pathogens (immune response) has critical implications for the occurrence of a wide range of chronic noncommunicable diseases, including obesity, diabetes, cardiovascular pathologies and cancer (108). Indeed, it has been well established that chronic inflammation is the trigger of the above-mentioned diseases (108) and, recently, the immunological adaptations in response to nutritional status have been highlighted. Undernutrition impairs immunity, causing inefficient responses to infections and vaccinations. Conversely, the metabolic overload in obesity can affect immunometabolism favoring chronic activation of both innate and adaptive immune cells, with subsequent low-grade systemic inflammation and altered susceptibility to autoimmune diseases (116). A thorough understanding of the intracellular network and intercellular correlations that regulate immunometabolism systemically is quite complex. For this reason, below we attempt to trace the main immunometabolic hubs of the organism.

## 5.1 Adipose tissue

The adipose tissue was identified as an important immune cell niche during homeostasis and an important immune-metabolic communication hub in metabolic syndrome (108, 117–121). Indeed, a wide range of immune cells are accumulated in the adipose tissue in the course of diet induced-obesity influencing the systemic metabolism (122, 123). The macrophages accumulate within adipose tissue produce the inflammatory cytokines IL-6 and TNF- $\alpha$  (124, 125). Locally, IL-6 can induce lipolysis in neighboring adipocytes and impair lipoprotein lipase, decreasing adipocyte lipid depot. The high circulating levels of IL-6 and free fatty acids (FFAs)



The immunometabolic control has significant clinical relevance for the health status of the organism. The reciprocal impact between metabolic and immune components unveils in the main body's immunometabolic hubs in both physiological and pathological conditions.

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promote insulin resistance and immune cell function alterations affecting both local and distal tissue microenvironments. Moreover, the accumulation of FFAs promotes ROS production in macrophages, which in turn augments activation of the NLRP3-ASC inflammasome (126), and peripheral insulin resistance, mediated by the secretion of the IL-1 $\beta$ . When inflammation persists, like in obesity and in non-communicable diseases, prolonged disruption of metabolic homeostasis could lead to immune cell dysfunction and dysregulated systemic metabolism. In this context, we are faced with 'sterile metabolic inflammation' that consists of persistent inflammation in the absence of infection (127). Here, the bilateral crosstalk between aberrant metabolism and immune regulation is disrupted and results not only in disease progression but leads also to immune senescence (128). Buck et al. have extensively reviewed the metabolic instruction of immunity and attempted to shed light on how feeding behaviors can also affect host immune fitness, but the relationship between the variety of factors that influence the systemic metabolism and immune cell activity is only beginning to be explored (7).

Recently, several lines of evidence on why the metabolic overload from obesity blunt the immune system and increases the vulnerability to infectious and autoimmune diseases have been reported (116, 129). Firstly, a functional impairment of both the innate and adaptive immune system is attributable to chronic lowgrade inflammation that lead to impaired chemotaxis, dysregulated production of pro- and anti-inflammatory factors and altered macrophages differentiation (130). This disrupts the delicate balance of adipose tissue between its function of immunologically active adipocytokines-producing organ, and the action of the latter on affecting adipocyte homeostasis and metabolism (131, 132). Moreover, the altered immunometabolism in obesity could lead to autoimmunity. The immune cell differentiation may be impaired in obese people due to excessive stimulation of nutrient- and energy-sensing pathways (such as increased mTOR activity) in immune cells, with consequent increase of proinflammatory TH1 and TH17 cells and decreases Tregs, which increase the risk of selftolerance lack (116). Lastly, exciting research demonstrated that the increased adiposity observed in obese people increase also the bone marrow adiposity (133), resulting quickly in a profound hematopoietic insult with reduction of lymphocyte population and compromised immune function (134–136).

## 5.2 Gut

The gut can be considered the main immunometabolic interface of the organism representing a barrier surface, where a single layer of epithelial cells is the main mediator of crosstalk between gut microbes in the lumen and host cells, including immune cells in the lamina propria (DCs, macrophages, innate lymphoid cells, and T cells). Peyer's patches are scattered along the epithelium which house germinal centers that maturate IgA-secreting B cells with the help of follicular helper T-cell help. B cells augment glycolysis upon activation and depend on pyruvate import for longevity as longlived plasma cells (7). The epithelial cells and intraepithelial lymphocytes in the gut coordinate the tightly regulated immune responses, implemented for both avoiding detrimental responses to commensals or food antigens and adequately respond to pathogens (137). It is noteworthy the role played by the short-chain fatty acids (SCFAs), metabolites produced by commensal bacteria able to influence B cell metabolism and boost antibody responses in both mouse and human B cells, promoting Ig A secretion (138). The presence of SCFAs and vitamins support the maintenance of barrier function by promoting the development and survival of Tregs and innate lymphoid cells. Noteworthy, homeostatic immune-driven signals secreted by gut resident immune cells (e.g., IL-10) may also mediate their effects through metabolic modulation (7). Indeed, it was demonstrated that the deficiency of the pleotropic antiinflammatory cytokine IL-10 in macrophages is sufficient to recapitulate the onset of severe colitis in mice (139). Possibly, the metabolic shift towards to aerobic glycolysis during innate immune cell activation is the explanation of the anti-inflammatory activity of the IL-10.

## 5.3 Liver

The liver can be defined as the immunometabolic controller of the organism. The central role played by the liver in the immunemetabolic homeostasis being well recognized (140). The function of the liver as the main metabolic organ inevitably exposes it to newly produced neo-antigens, enhancing the risk of overactivation of components of the immune system with potentially harmful consequences for hepatic cell homeostasis (141). Several evidence point out the importance of the liver as "regulatory system" where different immune and non-immune cell populations work together in order to protect the host from antigenic overload of dietary components and drugs derived from the gut, facilitating tolerance rather than immunoreactivity (141, 142). Indeed, the immune cells coexist in a close symbiotic manner to support the hepatic metabolic functions (143). In the liver, naïve T cells recirculating within the sinusoids make direct contact with sinusoidal cells, such as liver sinusoidal endothelial cells (LSECs) or Kupffer cells (144). Gut-derived food antigens are picked up by Kupffer cells, LSECs, and liver dendritic cells and presented to naïve T cells, leading to immune tolerance of both CD8+ T cells and CD4+ T cells (145). In addition, compelling evidence demonstrated that virus-induced innate immune responses in hepatocytes are mediated by the antiviral cytokine type I interferon (IFN-I) that apart from inducing an antiviral state, rewires cellular metabolism of innate immune cells to boost the production of immune-modulatory metabolites (146-148) and modulates cellular redox homeostasis and central metabolic pathways in hepatocytes (113, 149-151). Moreover, the portal blood delivers to the liver numerous factors derived from gut and visceral adipose tissue (e.g. pro-inflammatory cytokines, lipids and bacteria-derived factors, such as endotoxins) (152) that seem to be critical in the systemic and central inflammation (153). These endocrine and immune mediators, in turn, can modulate the hepatic metabolism by influencing the bioenergetic regulation of hepatic mitochondria. Conversely,

inflammation-induced metabolic reprogramming of hepatocytes can influence systemic energy metabolism (154). These data highlight the important role of the liver as central modulator of systemic immunometabolism and strengthen the bidirectional cause-effect relationship between mitochondrial metabolic stress and immune regulation.

### 5.4 Peripheral blood mononuclear cells

PBMCs are immunometabolic sentinels of the organism. PBMCs are circulating cells able to sense and respond to systemic metabolic and inflammatory stressors. They circulate continuously throughout the body in the bloodstream, and are subject to changes in blood composition, including those related to fluctuations in nutrients, substrates and hormones (155, 156). PBMCs can be defined sentinel cells able to respond either to internal signals (such as hormones) or external ones (such as nutrients) and to reflect energy metabolism of internal tissues with which they interact, as well as their gene expression profile. In addition, these cells contain respiring mitochondria and, therefore, are a functional biomarker in translational bioenergetics (157). For this reason, PBMCs represent a suitable system to study changes in cell metabolism and to control the management of immune surveillance (156). Moreover, since several researches in animal models demonstrated that PBMCs can reflect the metabolic framework that cannot or can hardly be sampled in humans, such as liver and brain (158-160). These cells can be used as a surrogate tissue to monitor nutritional responses and provide predictive disease risk markers (161).

# 6 Discussion and conclusions

This review has highlighted that immunometabolic control has significant clinical relevance to the health status of the organism. Furthermore, it has left no doubt that mitochondria are the main players in this fine-tuning between metabolism and immune function, finding the metabolic flexibility of immune cells and the mitochondrial dynamics processes to be the secrets of appropriate immunometabolic homeostasis (see also graphical abstract). In particular, we underlined the importance of the metabolic flexibility of immune cells, exploring the role of their metabolic pathways and how this regulates the outcome of the immune response. On the one hand, it is well established that initial T and B cells activation, during the period of quiescence exit, lead to increased glucose uptake and promote aerobic glycolysis (23). Then, effector T and B cell subsets display differences in metabolic activities on the basis of their subsequent functional specialization (23). T<sub>reg</sub> cells and memory T cells revert to a catabolic state and rely mainly on mitochondrial fatty acid oxidation (162). Also metabolic programs of activated B lymphocytes change depending on their functional destination (tolerance, effector or regulatory activities) and on the receptors and co-activation molecules stimulated (36). In addition, we explored the ability of mitochondrial DAMPs to employ the immunogenic capacity in immune and non-immune cells, and the significant relevance of mitochondrial ROS production in the trigger of an adequate immune response (163, 164). We traced the thread that leads from malnutrition to metabolic inflexibility in immune and non-immune cells, with consequent systemic meta-inflammation and disruption of the immune system. At once, a proper metabolic regulation supports immune cell activities in physiological contexts, while dysregulated immunometabolism contributes to pathophysiology. In the last chapter of the review, we explored and discussed the intricate intracellular networks and intercellular correlations in the main immunometabolic hubs of the organism. We highlighted as the interference of mitochondrial dysfunction (unbalanced DAMPs and ROS production and metabolic inflexibility) influence also non-immune components and lead to chronic and systemic inflammatory responses, typical features of noncommunicable diseases.

For this reason, it is advisable to pursue a constant and in-depth exploration of immunometabolism, both in the detailed molecular pathways involved and with an interdisciplinary approach. The aim is to identify mitochondrial targets useful for the development of new intervention therapies that could help reduce the global burden of metabolic, inflammatory and autoimmune diseases.

## Author contributions

GT: Conceptualization, Investigation, Visualization, Writing – original draft, Writing – review & editing. FC: Investigation, Writing – review & editing, Writing – original draft, Visualization. AC: Writing – original draft. GC: Writing – original draft. MM: Conceptualization, Project administration, Validation, Writing – review & editing.

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# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

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# References

1. Hachiya R, Tanaka M, Itoh M, Suganami T. Molecular mechanism of crosstalk between immune and metabolic systems in metabolic syndrome. *Inflammation Regener*. (2022) 42:13. doi: 10.1186/s41232-022-00198-7

2. Ganeshan K, Chawla A. Metabolic regulation of immune responses. Annu Rev Immunol. (2014) 32:609–34. doi: 10.1146/annurev-immunol-032713-120236

3. Chi H. Immunometabolism at the intersection of metabolic signaling, cell fate, and systems immunology. *Cell Mol Immunol.* (2022) 19:299–302. doi: 10.1038/s41423-022-00840-x

4. Collins N, Belkaid Y. Control of immunity via nutritional interventions. Immunity. (2022) 55:210-23. doi: 10.1016/j.immuni.2022.01.004

5. Soerens AG, Künzli M, Quarnstrom CF, Scott MC, Swanson L, Locquiao Jj, et al. Functional T cells are capable of supernumerary cell division and longevity. *Nature*. (2023) 614:762–6. doi: 10.1038/s41586-022-05626-9

 Rastogi I, Jeon D, Moseman JE, Muralidhar A, Potluri HK, McNeel DG. Role of B cells as antigen presenting cells. *Front Immunol.* (2022) 13:954936. doi: 10.3389/ fimmu.2022.954936

7. Buck MD, Sowell RT, Kaech SM, Pearce EL. Metabolic instruction of immunity. *Cell.* (2017) 169:570–86. doi: 10.1016/j.cell.2017.04.004

8. Caputa G, Castoldi A, Pearce EJ. Metabolic adaptations of tissue-resident immune cells. *Nat Immunol.* (2019) 20:793–801. doi: 10.1038/s41590-019-0407-0

9. Wang A, Luan HH, Medzhitov R. An evolutionary perspective on immunometabolism. *Science*. (2019) 363:eaar3932. doi: 10.1126/science.aar3932

10. Buck MD, O'Sullivan D, Pearce EL. T cell metabolism drives immunity. J Exp Med. (2015) 212:1345-60. doi: 10.1084/jem.20151159

11. Fox CJ, Hammerman PS, Thompson CB. Fuel feeds function: energy metabolism and the T-cell response. *Nat Rev Immunol.* (2005) 5:844–52. doi: 10.1038/nri1710

12. Pearce EL, Poffenberger MC, Chang C-H, Jones RG. Fueling immunity: insights into metabolism and lymphocyte function. *Science*. (2013) 342:1242454. doi: 10.1126/ science.1242454

13. O'Neill LAJ, Pearce EJ. Immunometabolism governs dendritic cell and macrophage function. J Exp Med. (2016) 213:15–23. doi: 10.1084/jem.20151570

14. Van den Bossche J, O'Neill LA, Menon D. Macrophage immunometabolism: where are we (Going)? *Trends Immunol.* (2017) 38:395-406. doi: 10.1016/j.it.2017.03.001

15. Pearce EL, Pearce EJ. Metabolic pathways in immune cell activation and quiescence. *Immunity*. (2013) 38:633-43. doi: 10.1016/j.immuni.2013.04.005

16. Muoio DM. Metabolic inflexibility: when mitochondrial indecision leads to metabolic gridlock. *Cell.* (2014) 159:1253–62. doi: 10.1016/j.cell.2014.11.034

17. Warburg O. On the origin of cancer cells. *Science*. (1956) 123:309-14. doi: 10.1126/science.123.3191.309

18. Vats D, Mukundan L, Odegaard JI, Zhang L, Smith KL, Morel CR, et al. Oxidative metabolism and PGC-1 $\beta$  attenuate macrophage-mediated inflammation. *Cell Metab.* (2006) 4:13–24. doi: 10.1016/j.cmet.2006.05.011

19. Lee H, Jeon J-H, Kim ES. Mitochondrial dysfunctions in T cells: focus on inflammatory bowel disease. *Front Immunol.* (2023) 14:1219422. doi: 10.3389/fimmu.2023.1219422

20. Boothby M, Rickert RC. Metabolic regulation of the immune humoral response. *Immunity*. (2017) 46:743–55. doi: 10.1016/j.immuni.2017.04.009

21. Chapman K, Ivanovici A, Taleski M, Sturrock CJ, Ng JLP, Mohd-Radzman NA, et al. CEP receptor signalling controls root system architecture in Arabidopsis and Medicago. *New Phytol.* (2020) 226:1809–21. doi: 10.1111/nph.16483

22. Reina-Campos M, Scharping NE, Goldrath AW. CD8+ T cell metabolism in infection and cancer. *Nat Rev Immunol.* (2021) 21:718–38. doi: 10.1038/s41577-021-00537-8

23. Chapman NM, Chi H. Metabolic adaptation of lymphocytes in immunity and disease. *Immunity*. (2022) 55:14–30. doi: 10.1016/j.immuni.2021.12.012

24. Wang R, Dillon CP, Shi LZ, Milasta S, Carter R, Finkelstein D, et al. The transcription factor myc controls metabolic reprogramming upon T lymphocyte activation. *Immunity*. (2011) 35:871–82. doi: 10.1016/j.immuni.2011.09.021

25. Kidani Y, Elsaesser H, Hock MB, Vergnes L, Williams KJ, Argus JP, et al. Sterol regulatory element-binding proteins are essential for the metabolic programming of effector T cells and adaptive immunity. *Nat Immunol.* (2013) 14:489–99. doi: 10.1038/ni.2570

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26. Clever D, Roychoudhuri R, Constantinides MG, Askenase MH, Sukumar M, Klebanoff CA, et al. Oxygen sensing by T cells establishes an immunologically tolerant metastatic niche. *Cell*. (2016) 166:1117–1131.e14. doi: 10.1016/j.cell.2016.07.032

27. Shi LZ, Wang R, Huang G, Vogel P, Neale G, Green DR, et al. HIF1 $\alpha$ -dependent glycolytic pathway orchestrates a metabolic checkpoint for the differentiation of TH17 and Treg cells. *J Exp Med.* (2011) 208:1367–76. doi: 10.1084/jem.20110278

28. Makowski L, Chaib M, Rathmell JC. Immunometabolism: From basic mechanisms to translation. *Immunol Rev.* (2020) 295:5-14. doi: 10.1111/imr.12858

29. van der Windt GJ, Everts B, Chang C-H, Curtis JD, Freitas TC, Amiel E, et al. Mitochondrial respiratory capacity is a critical regulator of CD8+ T cell memory development. *Immunity.* (2012) 36:68–78. doi: 10.1016/j.immuni.2011.12.007

 Caro-Maldonado A, Wang R, Nichols AG, Kuraoka M, Milasta S, Sun LD, et al. Metabolic reprogramming is required for antibody production that is suppressed in anergic but exaggerated in chronically BAFF-exposed B cells. J Immunol. (2014) 192:3626–36. doi: 10.4049/jimmunol.1302062

31. Cho SH, Ahn AK, Bhargava P, Lee C-H, Eischen CM, McGuinness O, et al. Glycolytic rate and lymphomagenesis depend on PARP14, an ADP ribosyltransferase of the B aggressive lymphoma (BAL) family. *Proc Natl Acad Sci.* (2011) 108:15972–7. doi: 10.1073/pnas.1017082108

 Dufort FJ, Bleiman BF, Gumina MR, Blair D, Wagner DJ, Roberts MF, et al. Cutting edge: IL-4-mediated protection of primary B lymphocytes from apoptosis via stat6-dependent regulation of glycolytic metabolism. J Immunol. (2007) 179:4953–7. doi: 10.4049/jimmunol.179.8.4953

33. Doughty CA, Bleiman BF, Wagner DJ, Dufort FJ, Mataraza JM, Roberts MF, et al. Antigen receptor-mediated changes in glucose metabolism in B lymphocytes: role of phosphatidylinositol 3-kinase signaling in the glycolytic control of growth. *Blood.* (2006) 107:4458–65. doi: 10.1182/blood-2005-12-4788

34. Waters LR, Ahsan FM, Wolf DM, Shirihai O, Teitell MA. Initial B cell activation induces metabolic reprogramming and mitochondrial remodeling. *iScience*. (2018) 5:99–109. doi: 10.1016/j.isci.2018.07.005

35. Boothby MR, Brookens SK, Raybuck AL, Cho SH. Supplying the trip to antibody production—nutrients, signaling, and the programming of cellular metabolism in the mature B lineage. *Cell Mol Immunol.* (2022) 19:352–69. doi: 10.1038/s41423-021-00782-w

36. Iperi C, Bordron A, Dueymes M, Pers J-O, Jamin C. Metabolic program of regulatory B lymphocytes and influence in the control of Malignant and autoimmune situations. *Front Immunol.* (2021) 12:735463. doi: 10.3389/fimmu.2021.735463

37. Müschen M. Metabolic gatekeepers to safeguard against autoimmunity and oncogenic B cell transformation. *Nat Rev Immunol.* (2019) 19:337–48. doi: 10.1038/s41577-019-0154-3

38. Sadras T, Chan LN, Xiao G, Müschen M. Metabolic gatekeepers of pathological B cell activation. *Annu Rev Pathol: Mech Dis.* (2021) 16:323–49. doi: 10.1146/annurev-pathol-061020-050135

39. Reid MA, Dai Z, Locasale JW. The impact of cellular metabolism on chromatin dynamics and epigenetics. *Nat Cell Biol.* (2017) 19:1298–306. doi: 10.1038/ncb3629

40. Diskin C, Ryan TAJ, O'Neill LAJ. Modification of proteins by metabolites in immunity. *Immunity*. (2021) 54:19–31. doi: 10.1016/j.immuni.2020.09.014

41. Calciolari B, Scarpinello G, Tubi LQ, Piazza F, Carrer A. Metabolic control of epigenetic rearrangements in B cell pathophysiology. *Open Biol.* (2022) 12:220038. doi: 10.1098/rsob.220038

42. Lu C, Thompson CB. Metabolic regulation of epigenetics. Cell Metab. (2012) 16:9–17. doi: 10.1016/j.cmet.2012.06.001

43. Bock FJ, Tait SWG. Mitochondria as multifaceted regulators of cell death. Nat Rev Mol Cell Biol. (2020) 21:85–100. doi: 10.1038/s41580-019-0173-8

44. Vringer E, Tait SWG. Mitochondria and cell death-associated inflammation. *Cell Death Differ*. (2023) 30:304–12. doi: 10.1038/s41418-022-01094-w

45. Rossmann MP, Dubois SM, Agarwal S, Zon LI. Mitochondrial function in development and disease. *Dis Models Mech.* (2021) 14:dmm048912. doi: 10.1242/dmm.048912

46. Osellame LD, Blacker TS, Duchen MR. Cellular and molecular mechanisms of mitochondrial function. *Best Pract Res Clin Endocrinol Metab.* (2012) 26:711–23. doi: 10.1016/j.beem.2012.05.003

47. Kennedy EP, Lehninger AL. Oxidation of fatty acids and tricarboxylic acid cycle intermediates by isolated rat liver mitochondria. *J Biol Chem.* (1949) 179:957–72. doi: 10.1016/S0021-9258(19)51289-3

48. Sagan L. On the origin of mitosing cells. J Theor Biol. (1967) 14:225–IN6. doi: 10.1016/0022-5193(67)90079-3

49. Fox TD. Mitochondrial protein synthesis, import, and assembly. *Genetics*. (2012) 192:1203–34. doi: 10.1534/genetics.112.141267

50. Iwasaki Y, Takeshima Y, Fujio K. Basic mechanism of immune system activation by mitochondria. *Immunol Med.* (2020) 43:142-7. doi: 10.1080/25785826.2020.1756609

51. David L. The mitochondria of microorganisms. London: Academic Press (1974). 553 p.

52. Meyer A, Laverny G, Bernardi L, Charles AL, Alsaleh G, Pottecher J, et al. Mitochondria: an organelle of bacterial origin controlling inflammation. *Front Immunol.* (2018) 9:536. doi: 10.3389/fimmu.2018.00536

53. Pallen MJ. Time to recognise that mitochondria are bacteria? *Trends Microbiol.* (2011) 19:58–64. doi: 10.1016/j.tim.2010.11.001

54. O'Rourke B. From bioblasts to mitochondria: ever expanding roles of mitochondria in cell physiology. *Front Physiol.* (2010) 1:7. doi: 10.3389/ fphys.2010.00007

55. Nunnari J, Suomalainen A. Mitochondria: in sickness and in health. *Cell*. (2012) 148:1145–59. doi: 10.1016/j.cell.2012.02.035

56. Gunter TE, Buntinas L, Sparagna GC, Gunter KK. The Ca2+ transport mechanisms of mitochondria and Ca2+ uptake from physiological-type Ca2+ transients. *Biochim Biophys Acta (BBA) - Bioenerget.* (1998) 1366:5–15. doi: 10.1016/S0005-2728(98)00117-0

57. Hockenbery D, Nuñez G, Milliman C, Schreiber RD, Korsmeyer SJ. Bcl-2 is an inner mitochondrial membrane protein that blocks programmed cell death. *Nature*. (1990) 348:334–6. doi: 10.1038/348334a0

58. Angajala A, Lim S, Phillips JB, Kim J-H, Yates C, You Z, et al. Diverse roles of mitochondria in immune responses: novel insights into immuno-metabolism. *Front Immunol.* (2018) 9:1605. doi: 10.3389/fimmu.2018.01605

59. Marchi S, Guilbaud E, Tait SWG, Yamazaki T, Galluzzi L. Mitochondrial control of inflammation. *Nat Rev Immunol.* (2023) 23:159–73. doi: 10.1038/s41577-022-00760-x

60. Shadel GS, Horvath TL. Mitochondrial ROS signaling in organismal homeostasis. *Cell.* (2015) 163:560-9. doi: 10.1016/j.cell.2015.10.001

61. Hamanaka RB, Chandel NS. Mitochondrial reactive oxygen species regulate cellular signaling and dictate biological outcomes. *Trends Biochem Sci.* (2010) 35:505–13. doi: 10.1016/j.tibs.2010.04.002

62. Erusalimsky JD, Moncada S. Nitric oxide and mitochondrial signaling. Arteriosclerosis Thrombosis Vasc Biol. (2007) 27:2524-31. doi: 10.1161/ ATVBAHA.107.151167

63. Rodríguez-Nuevo A, Zorzano A. The sensing of mitochondrial DAMPs by nonimmune cells. *Cell Stress.* (2019) 3:195–207. doi: 10.15698/cst2019.06.190

64. Tannahill GM, Curtis AM, Adamik J, Palsson-McDermott EM, McGettrick AF, Goel G, et al. Succinate is an inflammatory signal that induces IL-1 $\beta$  through HIF-1 $\alpha$ . *Nature*. (2013) 496:238–42. doi: 10.1038/nature11986

65. Mills E, O'Neill LAJ. Succinate: a metabolic signal in inflammation. *Trends Cell Biol.* (2014) 24:313–20. doi: 10.1016/j.tcb.2013.11.008

66. Rathinam VAK, Fitzgerald KA. Inflammasome complexes: emerging mechanisms and effector functions. *Cell.* (2016) 165:792-800. doi: 10.1016/j.cell.2016.03.046

67. Deli T, Csernoch L. Extracellular ATP and cancer—An overview with special reference to P2 purinergic receptors. *Pathol Oncol Res.* (2008) 14:219–31. doi: 10.1007/s12253-008-9071-7

68. Bours MJL, Swennen ELR, Virgilio FD, Cronstein BN, Dagnelie PC. Adenosine 5'-triphosphate and adenosine as endogenous signaling molecules in immunity and inflammation. *Pharmacol Ther.* (2006) 112:358-404. doi: 10.1016/j.pharmthera.2005.04.013

69. Iyer SS, He Q, Janczy JR, Elliott EI, Zhong Z, Olivier AK, et al. Mitochondrial cardiolipin is required for nlrp3 inflammasome activation. *Immunity*. (2013) 39:311–23. doi: 10.1016/j.immuni.2013.08.001

70. Chu CT, Ji J, Dagda RK, Jiang JF, Tyurina YY, Kapralov AA, et al. Cardiolipin externalization to the outer mitochondrial membrane acts as an elimination signal for mitophagy in neuronal cells. *Nat Cell Biol.* (2013) 15:1197–205. doi: 10.1038/ncb2837

71. Miao R, Jiang C, Chang WY, Zhang H, An J, Ho F, et al. Gasdermin D permeabilization of mitochondrial inner and outer membranes accelerates and enhances pyroptosis. *Immunity*. (2023) 56:2523-2541.e8. doi: 10.1016/j.immuni.2023.10.004

72. Czapiga M, Gao J-L, Kirk A, Lekstrom-Himes J. Human platelets exhibit chemotaxis using functional N-formyl peptide receptors. *Exp Hematol.* (2005) 33:73–84. doi: 10.1016/j.exphem.2004.09.010

73. Oka T, Hikoso S, Yamaguchi O, Taneike M, Takeda T, Tamai T, et al. Mitochondrial DNA that escapes from autophagy causes inflammation and heart failure. *Nature*. (2012) 485:251–5. doi: 10.1038/nature10992

74. West AP, Khoury-Hanold W, Staron M, Tal MC, Pineda CM, Lang SM, et al. Mitochondrial DNA stress primes the antiviral innate immune response. *Nature*. (2015) 520:553–7. doi: 10.1038/nature14156

75. Tsuji N, Tsuji T, Ohashi N, Kato A, Fujigaki Y, Yasuda H. Role of mitochondrial DNA in septic AKI via toll-like receptor 9. J Am Soc Nephrol. (2016) 27:2009–20. doi: 10.1681/ASN.2015040376

76. Julian MW, Shao G, Bao S, Knoell DL, Papenfuss TL, VanGundy ZC, et al. Mitochondrial transcription factor A serves as a danger signal by augmenting plasmacytoid dendritic cell responses to DNA. *J Immunol.* (2012) 189:433–43. doi: 10.4049/jimmunol.1101375

77. Crouser ED, Shao G, Julian MW, Macre JE, Shadel GS, Tridandapani S, et al. Monocyte activation by necrotic cells is promoted by mitochondrial proteins and formyl peptide receptors. *Crit Care Med.* (2009) 37:2000–9. doi: 10.1097/CCM.0b013e3181a001ae

78. Zhou R, Yazdi AS, Menu P, Tschopp J. A role for mitochondria in NLRP3 inflammasome activation. *Nature*. (2011) 469:221–5. doi: 10.1038/nature09663

79. Sena LA, Li S, Jairaman A, Prakriya M, Ezponda T, Hildeman DA, et al. Mitochondria are required for antigen-specific T cell activation through reactive oxygen species signaling. *Immunity*. (2013) 38:225–36. doi: 10.1016/j.immuni.2012.10.020

80. Singh DK, Kumar D, Siddiqui Z, Basu SK, Kumar V, Rao KVS. The strength of receptor signaling is centrally controlled through a cooperative loop between ca2+ and an oxidant signal. *Cell.* (2005) 121:281–93. doi: 10.1016/j.cell.2005.02.036

81. Fernández-Ruiz I, Arnalich F, Cubillos-Zapata C, Hernández-Jiménez E, Moreno-González R, Toledano V, et al. Mitochondrial DAMPs induce endotoxin tolerance in human monocytes: an observation in patients with myocardial infarction. *PloS One.* (2014) 9:e95073. doi: 10.1371/journal.pone.0095073

82. Mohanty A, Tiwari-Pandey R, Pandey NR. Mitochondria: the indispensable players in innate immunity and guardians of the inflammatory response. J Cell Commun Signaling. (2019) 13:303–18. doi: 10.1007/s12079-019-00507-9

83. Spinelli JB, Haigis MC. The multifaceted contributions of mitochondria to cellular metabolism. *Nat Cell Biol.* (2018) 20:745–54. doi: 10.1038/s41556-018-0124-1

84. McBride HM, Neuspiel M, Wasiak S. Mitochondria: more than just a powerhouse. Curr Biol. (2006) 16:R551-60. doi: 10.1016/j.cub.2006.06.054

85. Galloway CA, Lee H, Yoon Y. Mitochondrial morphology—emerging role in bioenergetics. *Free Radical Biol Med.* (2012) 53:2218–28. doi: 10.1016/ j.freeradbiomed.2012.09.035

86. De Oliveira LG, Angelo YDS, Iglesias AH, Peron JPS. Unraveling the link between mitochondrial dynamics and neuroinflammation. *Front Immunol.* (2021) 12:624919. doi: 10.3389/fimmu.2021.624919

87. Merz S, Hammermeister M, Altmann K, Dürr M, Westermann B. Molecular machinery of mitochondrial dynamics in yeast. *bchm*. (2007) 388:917–26. doi: 10.1515/ BC.2007.110

88. Tilokani L, Nagashima S, Paupe V, Prudent J. Mitochondrial dynamics: overview of molecular mechanisms. *Essays Biochem*. (2018) 62:341-60. doi: 10.1042/EBC20170104

89. Mollica MP, Raso GM, Cavaliere G, Trinchese G, Filippo CD, Aceto S, et al. Butyrate regulates liver mitochondrial function, efficiency, and dynamics in insulinresistant obese mice. *Diabetes.* (2017) 66:1405–18. doi: 10.2337/db16-0924

90. Trinchese G, Cavaliere G, Filippo CD, Aceto S, Prisco M, Chun JT, et al. Human milk and donkey milk, compared to cow milk, reduce inflammatory mediators and modulate glucose and lipid metabolism, acting on mitochondrial function and oleylethanolamide levels in rat skeletal muscle. *Front Physiol.* (2018) 9:32. doi: 10.3389/fphys.2018.00032

91. Haigh JL, New LE, Filippi BM. Mitochondrial dynamics in the brain are associated with feeding, glucose homeostasis, and whole-body metabolism. *Front Endocrinol.* (2020) 11:580879. doi: 10.3389/fendo.2020.580879

92. Dai W, Jiang L. Dysregulated mitochondrial dynamics and metabolism in obesity, diabetes, and cancer. *Front Endocrinol.* (2019) 10:570. doi: 10.3389/ fendo.2019.00570

93. Liesa M, Shirihai OS. Mitochondrial dynamics in the regulation of nutrient utilization and energy expenditure. *Cell Metab.* (2013) 17:491–506. doi: 10.1016/j.cmet.2013.03.002

94. Sebastián D, Hernández-Alvarez MI, Segalés J, Sorianello E, Muñoz JP, Sala D, et al. Mitofusin 2 (Mfn2) links mitochondrial and endoplasmic reticulum function with insulin signaling and is essential for normal glucose homeostasis. *Proc Natl Acad Sci.* (2012) 109:5523–8. doi: 10.1073/pnas.1108220109

95. Wang L, Ishihara T, Ibayashi Y, Tatsushima K, Setoyama D, Hanada Y, et al. Disruption of mitochondrial fission in the liver protects mice from diet-induced obesity and metabolic deterioration. *Diabetologia.* (2015) 58:2371–80. doi: 10.1007/s00125-015-3704-7

96. Cervantes-Silva MP, Cox SL, Curtis AM. Alterations in mitochondrial morphology as a key driver of immunity and host defence. *EMBO Rep.* (2021) 22: e53086. doi: 10.15252/embr.202153086

97. Buck MD, O'Sullivan D, Geltink RIK, Curtis JD, Chang C-H, Sanin DE, et al. Mitochondrial dynamics controls T cell fate through metabolic programming. *Cell*. (2016) 166:63–76. doi: 10.1016/j.cell.2016.05.035

98. Zhang B, Alysandratos K-D, Angelidou A, Asadi S, Sismanopoulos N, Delivanis D-A, et al. Human mast cell degranulation and preformed TNF secretion require

mitochondrial translocation to exocytosis sites: Relevance to atopic dermatitis. J Allergy Clin Immunol. (2011) 127:1522–1531.e8. doi: 10.1016/j.jaci.2011.02.005

99. West AP, Shadel GS, Ghosh S. Mitochondria in innate immune responses. *Nat Rev Immunol.* (2011) 11:389–402. doi: 10.1038/nri2975

100. Koshiba T, Yasukawa K, Yanagi Y, Kawabata S. Mitochondrial membrane potential is required for MAVS-mediated antiviral signaling. *Sci Signaling*. (2011) 4:ra7. doi: 10.1126/scisignal.2001147

101. Amini P, Stojkov D, Felser A, Jackson CB, Courage C, Schaller A, et al. Neutrophil extracellular trap formation requires OPA1-dependent glycolytic ATP production. *Nat Commun.* (2018) 9:2958. doi: 10.1038/s41467-018-05387-y

102. Rambold AS, Pearce EL. Mitochondrial dynamics at the interface of immune cell metabolism and function. *Trends Immunol.* (2018) 39:6–18. doi: 10.1016/j.it.2017.08.006

103. Xie J-H, Li Y-Y, Jin J. The essential functions of mitochondrial dynamics in immune cells. *Cell Mol Immunol.* (2020) 17:712–21. doi: 10.1038/s41423-020-0480-1

104. Desdín-Micó G, Soto-Heredero G, Mittelbrunn M. Mitochondrial activity in T cells. *Mitochondrion.* (2018) 41:51–7. doi: 10.1016/j.mito.2017.10.006

105. Ron-Harel N, Santos D, Ghergurovich JM, Sage PT, Reddy A, Lovitch SB, et al. Mitochondrial biogenesis and proteome remodeling promote one-carbon metabolism for T cell activation. *Cell Metab.* (2016) 24:104–17. doi: 10.1016/j.cmet.2016.06.007

106. Sandoval H, Kodali S, Wang J. Regulation of B cell fate, survival, and function by mitochondria and autophagy. *Mitochondrion*. (2018) 41:58–65. doi: 10.1016/j.mito.2017.11.005

107. Westermann B. Mitochondrial fusion and fission in cell life and death. Nat Rev Mol Cell Biol. (2010) 11:872–84. doi: 10.1038/nrm3013

108. Hotamisligil GS. Inflammation, metaflammation and immunometabolic disorders. *Nature*. (2017) 542:177-85. doi: 10.1038/nature21363

109. Krauss M, Schaller S, Borchers S, Findeisen R, Lippert J, Kuepfer L. Integrating cellular metabolism into a multiscale whole-body model. *PloS Comput Biol.* (2012) 8: e1002750. doi: 10.1371/journal.pcbi.1002750

110. Perry RJ, Camporez J-PG, Kursawe R, Titchenell PM, Zhang D, Perry CJ, et al. Hepatic acetyl coA links adipose tissue inflammation to hepatic insulin resistance and type 2 diabetes. *Cell.* (2015) 160:745–58. doi: 10.1016/j.cell.2015.01.012

111. Priest C, Tontonoz P. Inter-organ cross-talk in metabolic syndrome. Nat Metab. (2019) 1:1177-88. doi: 10.1038/s42255-019-0145-5

112. Chu C, Artis D, Chiu IM. Neuro-immune interactions in the tissues. *Immunity*. (2020) 52:464–74. doi: 10.1016/j.immuni.2020.02.017

113. Lercher A, Baazim H, Bergthaler A. Systemic immunometabolism: challenges and opportunities. *Immunity*. (2020) 53:496–509. doi: 10.1016/j.immuni.2020.08.012

114. Cornier M-A, Dabelea D, Hernandez TL, Lindstrom RC, Steig AJ, Stob NR, et al. The metabolic syndrome. *Endocrine Rev.* (2008) 29:777-822. doi: 10.1210/er.2008-0024

115. Marcos A, Nova E, Montero A. Changes in the immune system are conditioned by nutrition. *Eur J Clin Nutr.* (2003) 57:S66–9. doi: 10.1038/sj.ejcn.1601819

116. Matarese G. The link between obesity and autoimmunity. *Science*. (2023) 379:1298–300. doi: 10.1126/science.ade0113

117. Cavaliere G, Cimmino F, Trinchese G, Catapano A, Petrella L, D'Angelo M, et al. From obesity-induced low-grade inflammation to lipotoxicity and mitochondrial dysfunction: altered multi-crosstalk between adipose tissue and metabolically active organs. *Antioxidants.* (2023) 12:1172. doi: 10.3390/antiox12061172

118. Cavaliere G, Catapano A, Trinchese G, Cimmino F, Menale C, Petrella L, et al. Crosstalk between adipose tissue and hepatic mitochondria in the development of the inflammation and liver injury during ageing in high-fat diet fed rats. *IJMS*. (2023) 24:2967. doi: 10.3390/ijms24032967

119. Lionetti L, Mollica MP, Lombardi A, Cavaliere G, Gifuni G, Barletta A. From chronic overnutrition to insulin resistance: The role of fat-storing capacity and inflammation. *Nutrition Metab Cardiovasc Dis.* (2009) 19:146–52. doi: 10.1016/j.numecd.2008.10.010

120. Roberts SB. Energy regulation and aging: recent findings and their implications. *Nutr Rev.* (2009) 58:91–7. doi: 10.1111/j.1753-4887.2000.tb07538.x

121. Schipper HS, Prakken B, Kalkhoven E, Boes M. Adipose tissue-resident immune cells: key players in immunometabolism. *Trends Endocrinol Metab.* (2012) 23:407–15. doi: 10.1016/j.tem.2012.05.011

122. Jaitin DA, Adlung L, Thaiss CA, Weiner A, Li B, Descamps H, et al. Lipidassociated macrophages control metabolic homeostasis in a trem2-dependent manner. *Cell.* (2019) 178:686–698.e14. doi: 10.1016/j.cell.2019.05.054

123. Aouadi M, Tencerova M, Vangala P, Yawe JC, Nicoloro SM, Amano SU, et al. Gene silencing in adipose tissue macrophages regulates whole-body metabolism in obese mice. *Proc Natl Acad Sci.* (2013) 110:8278–83. doi: 10.1073/pnas.1300492110

124. Cinti S, Mitchell G, Barbatelli G, Murano I, Ceresi E, Faloia E, et al. Adipocyte death defines macrophage localization and function in adipose tissue of obese mice and humans. *J Lipid Res.* (2005) 46:2347–55. doi: 10.1194/jlr.M500294-JLR200

125. Kunz HE, Hart CR, Gries KJ, Parvizi M, Laurenti M, Man CD, et al. Adipose tissue macrophage populations and inflammation are associated with systemic inflammation and insulin resistance in obesity. *Am J Physiology-Endocrinol Metab.* (2021) 321:E105–21. doi: 10.1152/ajpendo.00070.2021

126. Guo H, Callaway JB, Ting JP-Y. Inflammasomes: mechanism of action, role in disease, and therapeutics. *Nat Med.* (2015) 21:677–87. doi: 10.1038/nm.3893

127. Chen GY, Nuñez G. Sterile inflammation: sensing and reacting to damage. Nat Rev Immunol. (2010) 10:826–37. doi: 10.1038/nri2873

128. van de Vyver M. Immunology of chronic low-grade inflammation: relationship with metabolic function. *J Endocrinol.* (2023) 257:e220271. doi: 10.1530/JOE-22-0271

129. Muscogiuri G, Pugliese G, Laudisio D, Castellucci B, Barrea L, Savastano S, et al. The impact of obesity on immune response to infection: Plausible mechanisms and outcomes. *Obes Rev.* (2021) 22:e13216. doi: 10.1111/obr.13216

130. Kawai T, Autieri MV, Scalia R. Adipose tissue inflammation and metabolic dysfunction in obesity. *Am J Physiology-Cell Physiol.* (2021) 320:C375–91. doi: 10.1152/ajpcell.00379.2020

131. Liu R, Nikolajczyk BS. Tissue immune cells fuel obesity-associated inflammation in adipose tissue and beyond. *Front Immunol.* (2019) 10:1587. doi: 10.3389/fimmu.2019.01587

132. Wang Q, Wu H. T cells in adipose tissue: critical players in immunometabolism. Front Immunol. (2018) 9:2509. doi: 10.3389/fimmu.2018.02509

133. Bredella MA, Torriani M, Ghomi RH, Thomas BJ, Brick DJ, Gerweck AV, et al. Vertebral bone marrow fat is positively associated with visceral fat and inversely associated with IGF-1 in obese women. *Obesity.* (2011) 19:49–53. doi: 10.1038/ oby.2010.106

134. Chan ME, Adler BJ, Green DE, Rubin CT. Bone structure and B-cell populations, crippled by obesity, are partially rescued by brief daily exposure to low-magnitude mechanical signals. *FASEB J*. (2012) 26:4855–63. doi: 10.1096/fj.12-209841

135. Karlsson EA, Sheridan PA, Beck MA. Diet-induced obesity in mice reduces the maintenance of influenza-specific CD8+ Memory T cells. *J Nutr.* (2010) 140:1691–7. doi: 10.3945/jn.110.123653

136. Adler BJ, Kaushansky K, Rubin CT. Obesity-driven disruption of haematopoiesis and the bone marrow niche. *Nat Rev Endocrinol.* (2014) 10:737–48. doi: 10.1038/nrendo.2014.169

137. Olivares-Villagómez D, Kaer LV. Intestinal intraepithelial lymphocytes: sentinels of the mucosal barrier. *Trends Immunol.* (2018) 39:264–75. doi: 10.1016/j.it.2017.11.003

138. Kim M, Qie Y, Park J, Kim CH. Gut microbial metabolites fuel host antibody responses. *Cell Host Microbe*. (2016) 20:202-14. doi: 10.1016/j.chom.2016.07.001

139. Zigmond E, Bernshtein B, Friedlander G, Walker CR, Yona S, Kim KW, et al. Macrophage-restricted interleukin-10 receptor deficiency, but not IL-10 deficiency, causes severe spontaneous colitis. *Immunity*. (2014) 44:720–33. doi: 10.1016/j.immuni.2014.03.012

140. Hoogerland JA, Staels B, Dombrowicz D. Immune-metabolic interactions in homeostasis and the progression to NASH. *Trends Endocrinol Metab.* (2022) 33:690–709. doi: 10.1016/j.tem.2022.07.001

141. Bogdanos DP, Gao B, Gershwin ME. Liver Immunology. In: Terjung R, editor. *Comprehensive Physiology*. Hoboken, New Jersey: Wiley (2013). p. 567–98. doi: 10.1002/cphy.c120011

142. Knolle PA, Wohlleber D. Immunological functions of liver sinusoidal endothelial cells. *Cell Mol Immunol.* (2016) 13:347-53. doi: 10.1038/cmi.2016.5

143. Thomson AW, Knolle PA. Antigen-presenting cell function in the tolerogenic liver environment. *Nat Rev Immunol.* (2010) 10:753–66. doi: 10.1038/nri2858

144. Warren A, Couteur DGL, Fraser R, Bowen DG, McCaughan GW, Bertolino P. T lymphocytes interact with hepatocytes through fenestrations in murine liver sinusoidal endothelial cells. *Hepatology*. (2006) 44:1182–90. doi: 10.1002/hep.21378

145. Böttcher JP, Knolle PA, Stabenow D. Mechanisms balancing tolerance and immunity in the liver. *Diges Dis.* (2011) 29:384–90. doi: 10.1159/000329801

146. Fritsch SD, Weichhart T. Effects of interferons and viruses on metabolism. *Front Immunol.* (2016) 7:630. doi: 10.3389/fimmu.2016.00630

147. McNab F, Mayer-Barber K, Sher A, Wack A, O'Garra A. Type I interferons in infectious disease. *Nat Rev Immunol.* (2015) 15:87–103. doi: 10.1038/nri3787

148. Schoggins JW, Wilson SJ, Panis M, Murphy MY, Jones CT, Bieniasz P, et al. A diverse range of gene products are effectors of the type I interferon antiviral response. *Nature*. (2011) 472:481–5. doi: 10.1038/nature09907

149. Bhattacharya A, Hegazy AN, Deigendesch N, Kosack L, Cupovic J, Kandasamy RK, et al. Superoxide dismutase 1 protects hepatocytes from type I interferon-driven oxidative damage. *Immunity*. (2015) 43:974–86. doi: 10.1016/j.immuni.2015.10.013

150. Lercher A, Bhattacharya A, Popa AM, Caldera M, Schlapansky MF, Baazim H, et al. Type I interferon signaling disrupts the hepatic urea cycle and alters systemic metabolism to suppress T cell function. *Immunity*. (2019) 51:1074–1087.e9. doi: 10.1016/j.immuni.2019.10.014

151. Schoggins JW, Rice CM. Interferon-stimulated genes and their antiviral effector functions. *Curr Opin Virol.* (2011) 1:519–25. doi: 10.1016/j.coviro.2011.10.008

152. Robinson MW, Harmon C, O'Farrelly C. Liver immunology and its role in inflammation and homeostasis. *Cell Mol Immunol.* (2016) 13:267–76. doi: 10.1038/cmi.2016.3

153. Konrad D, Wueest S. The gut-adipose-liver axis in the metabolic syndrome. *Physiology.* (2014) 29:304–13. doi: 10.1152/physiol.00014.2014

154. Okin D, Medzhitov R. The effect of sustained inflammation on hepatic mevalonate pathway results in hyperglycemia. *Cell.* (2016) 165:343–56. doi: 10.1016/j.cell.2016.02.023

155. Zeng Y, David J, Rémond D, Dardevet D, Savary-Auzeloux I, Polakof S. Peripheral blood mononuclear cell metabolism acutely adapted to postprandial transition and mainly reflected metabolic adipose tissue adaptations to a high-fat diet in minipigs. *Nutrients.* (2018) 10:1816. doi: 10.3390/nu10111816

156. Caimari A, Oliver P, Keijer J, Palou A. Peripheral blood mononuclear cells as a model to study the response of energy homeostasis-related genes to acute changes in feeding conditions. *OMICS: A J Integr Biol.* (2010) 14:129–41. doi: 10.1089/omi.2009.0092

157. Altintas MM, DiBartolo S, Tadros L, Samelko B, Wasse H. Metabolic changes in peripheral blood mononuclear cells isolated from patients with end stage renal disease. *Front Endocrinol.* (2021) 12:629239. doi: 10.3389/fendo.2021.629239

158. Díaz-Rúa R, Keijer J, Caimari A, van Schothorst EM, Palou A, Oliver P. Peripheral blood mononuclear cells as a source to detect markers of homeostatic alterations caused by the intake of diets with an unbalanced macronutrient composition. *J Nutr Biochem.* (2015) 26:398–407. doi: 10.1016/j.jnutbio.2014.11.013

159. Caimari A, Oliver P, Rodenburg W, Keijer J, Palou A. Feeding conditions control the expression of genes involved in sterol metabolism in peripheral blood

mononuclear cells of normoweight and diet-induced (cafeteria) obese rats. J Nutr Biochem. (2010) 21:1127-33. doi: 10.1016/j.jnutbio.2009.10.001

160. Reynés B, Hazebroek MK, García-Ruiz E, Keijer J, Oliver P, Palou A. Specific features of the hypothalamic leptin signaling response to cold exposure are reflected in peripheral blood mononuclear cells in rats and ferrets. *Front Physiol.* (2017) 8:581. doi: 10.3389/fphys.2017.00581

161. Janssen JJE, Lagerwaard B, Bunschoten A, Savelkoul HFJ, van Neerven RJJ, Keijer J, et al. Novel standardized method for extracellular flux analysis of oxidative and glycolytic metabolism in peripheral blood mononuclear cells. *Sci Rep.* (2021) 11:1662. doi: 10.1038/s41598-021-81217-4

162. Newton R, Priyadharshini B, Turka LA. Immunometabolism of regulatory T cells. *Nat Immunol.* (2016) 17:618–25. doi: 10.1038/ni.3466

163. Shekhova E. Mitochondrial reactive oxygen species as major effectors of antimicrobial immunity. *PloS Pathog.* (2020) 16:e1008470. doi: 10.1371/journal.ppat.1008470

164. Tavassolifar MJ, Vodjgani M, Salehi Z, Izad M. The influence of reactive oxygen species in the immune system and pathogenesis of multiple sclerosis. *Autoimmune Dis.* (2020) 2020:1–14. doi: 10.1155/2020/5793817