

Review

The bacterial spore as a mucosal vaccine delivery system

Anella Saggese¹, Loredana Baccigalupi², Giuliana Donadio³, Ezio Ricca^{1,*} and Rachele Isticato¹

¹ Department of Biology, Federico II University, Naples Italy; (AS) anella.saggese@unina.it; (ER) ericca@unina.it; (RI) isticato@unina.it

² Department of Molecular Medicine and Medical Biotechnology, Federico II University, Naples, Italy; (LB) lorbacci@unina.it

³ Department of Pharmacy, University of Salerno, Salerno, Italy; (GD) gdonadio@unisa.it

* Correspondence: ericca@unina.it; Tel.: +39 081 679036.

Abstract: The development of efficient mucosal vaccines is strongly dependent on the use of appropriate vectors. Various biological systems or synthetic nanoparticles have been proposed to display and deliver antigens to mucosal surfaces. The *Bacillus* spore, a metabolically quiescent and extremely resistant cell, has also been proposed as a mucosal vaccine delivery system and shown able to conjugate the advantages of live and synthetic systems. Several antigens have been displayed on the spore by either recombinant or non-recombinant approaches, and antigen-specific immune responses have been observed in animals immunized by the oral or nasal route. Here we review the use of the bacterial spore as a mucosal vaccine vehicle focusing on the advantages and drawbacks of using the spore and of the recombinant vs. non-recombinant approach to display antigens on the spore surface. An overview of the immune responses induced by antigen-displaying spores so far tested in animals is presented and discussed.

Keywords: spores; mucosal vaccines; vaccine delivery; *Bacillus subtilis*; Spore surface display

1. Introduction

Vaccinations are the most effective strategy to control bacterial and viral infections. The World Health Organization (WHO) considers immunization campaigns as global health success stories, estimating that they prevent millions of deaths yearly (<https://www.who.int/health-topics/vaccines-and-immunization>). By mimicking the pathogen infection, vaccines induce the activation of the immune system and, therefore, the response against the pathogen [1]. After the immunization, the delivered antigen is recognized by the pattern recognition receptors (PRRs) of the innate immune cells (macrophages and dendritic cells), stimulating the production of cytokines and chemokines and leading to an increase in the number of antigen-presenting cells (APCs). These uptake, process, and present the antigen to the T cells that, in turn, induce B cells to produce antigen-specific antibodies [1, 2]. When the pathogen has been eliminated, the adaptive immune system develops the immunological memory, the basis of long-term protection and the final goal of a vaccine, since it leads to the persistence of antibodies and the generation of memory cells able to quickly react upon re-exposure to the same pathogen [1, 2].

Vaccines can be administered through either parenteral or mucosal routes. Most authorized vaccines are administered parenterally, i.e. injected subcutaneously (SC), intradermally (ID) or intramuscularly (IM). All three parenteral routes of vaccine administration have advantages and pitfalls, with the ID route inducing a stronger immune response than IM or SC but also causing more serious adverse reactions at the administration site [3]. In general, parenteral vaccines elicit a strong immune response but weak mucosal protection and do not prevent infection by the pathogen [4]. In addition, they require trained personnel for the administration (injection), making immunization campaigns difficult and expensive, especially in third-world countries [3]. On the other hand, mucosal vaccines induce protective cellular and humoral responses at both mucosal and systemic levels [5, 6]. The

induction of adaptive immunity at mucosal sites, involving secretory antibodies and tissue-resident T cells, prevents pathogen entry into the animal body, thus preventing the infection by the pathogen and its transmission [7]. Since mucosal vaccine do not require an injection, they are not invasive and easy to administrate facilitating mass immunization campaigns. The use of oral vaccinations would be preferable also for animal husbandry and aquaculture, since they reduce the costs of animal management and the stress caused to animals. However, mucosal vaccines also have drawbacks that have so far limited their use. Mucosal vaccines generally have low immunogenicity due to the high levels of tolerance induced by ingested antigens and to the lack of efficient mucosal adjuvants and delivery systems able to prevent antigen degradation at the mucosal sites [7]. Therefore, antigens administered by the mucosal route often cannot efficiently reach the inductive site of the mucosa-associated lymphoid tissues (see below) and trigger a strong immune response. As a consequence, the effectiveness of mucosal vaccines is often hindered and only a few mucosal vaccines, all based on live attenuated or death-inactivated pathogens, are currently approved for human use (Table 1).

This review will discuss the recent advancements in mucosal delivery systems, focusing on the use of bacterial spores as vaccine vehicles able to conjugate the advantages of live cells and synthetic nanoparticles.

Table 1. Licensed mucosal vaccines for human use^a

Pathogen	Trade Name	Composition	Route, dose	Immunological mechanism	Efficacy
Rotavirus	Rotarix; RotaTeq	Live attenuated	Oral, 3 doses	sIgA and systemic neutralizing IgG	Over 70–90%
Poliovirus	Orimune; OPV; Poliomyelitis vaccine	Live attenuated	Oral, 3 doses	sIgA and systemic IgG	Over 90%
<i>Salmonella typhi</i>	Vivotif; Ty21A	Live attenuated	Oral, 3–4 doses	sIgA, systemic IgG and CTL responses	Variable, but more than 50%
<i>Vibrio cholera</i>	Dukoral; ORC-Vax; Shanchol	Inactivated <i>V. cholera</i>	Oral, 2–3 doses	Antibacterial, toxin- specific and LPS-spe- cific IgA	Over 85%
Influenza Virus A	FluMist Quadrivalent®	Antigens incorporated into live attenuated, cold adapted influenza vector	Nasal, 1 dose	Mucosal IgA and systemic IgG	Over 90%
Influenza Virus A and B	Fluenz Tetra®	Antigens into live at- tenuated, cold adapted influenza vector	Nasal, 1 dose	Mucosal IgA, sys- temic IgG and CTL responses	Variable, but more than 50%

^aData from references 2 and 7.

2. Mucosal surfaces and mucosal immune system

The mucosal surfaces of the human body present structural and functional differences at the various body sites, i.e. the gastrointestinal, urogenital, respiratory tracts and oral and ocular cavities. In the gastrointestinal tract (GIT) the mucus is secreted by the Goblet cells and covers columnar epithelial cells. Antigens are transported from the lumen to dendritic cells (DCs) mainly through M cells, allowing antigen presentation to the mucosal immune system and inducing the production of IgG and secretory IgA (sIgA). In other cases, such as the oral and ocular cavities, the mucus layer covers a multilayer squamous epithelium and no M cells and goblet cells are present. In these cases, other tissues (glands) secrete the mucus, the DCs migrate to the adjacent lymph nodes upon antigen recognition, and only an IgG response is induced without the production of sIgA [8] (Figure 1).

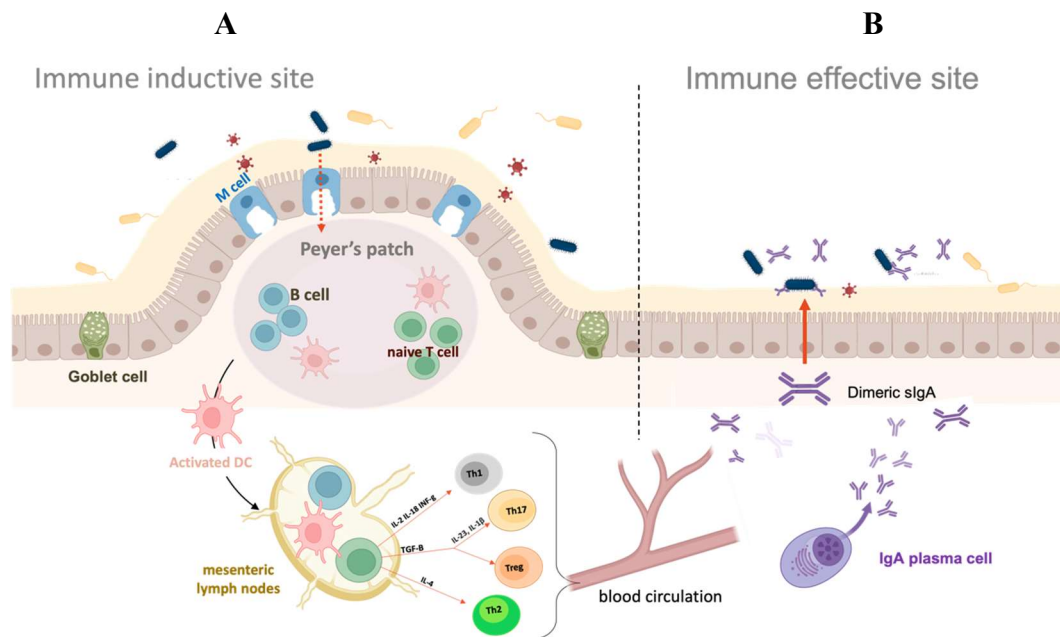


Figure 1. Gut-associated mucosal immune system. (A) Antigens within the intestinal lumen are transferred to the Peyer's patches via M cells and recognized by DCs. In the mesenteric lymph nodes, the activated DCs prime naïve T cells by the secretion of tissue-specific adhesion molecules and cytokines. The T helper cells are responsible for fighting different types of pathogens. Th1s secrete IFN- γ , activating macrophages and cytotoxic T cells against intracellular pathogens. Th2 cells produce IL-4, IL-5, and IL-13, which activate the humoral immune responses against extracellular pathogens by the activation of B cells. Finally, Th17s are responsible for antifungal and antibacterial immunity. T cell-dependent activation of IgA-committed B cells is also induced. (B) Then antigen-specific T cells and IgA-committed B cells migrate to effector sites through blood circulation. IgA-committed B cells differentiate into IgA-producing plasma cells in the presence of cytokines produced by Th2 cells, and they subsequently produce dimeric forms of IgA. Finally, the IgA dimeric forms are secreted and released in the intestinal tract, where play critical roles in mucosal immune responses such as immune exclusion, antigen excretion, and intracellular virus neutralization.

The mucosal surfaces represent contact sites between the body and the external environment and are in direct contact with all microbes, viruses, and molecules present in air, water, and foods. Physical, chemical, and immunological barriers are then essential to protect the mucosal surfaces and maintain homeostasis, avoiding chronic inflammatory responses due to the high antigen load. The dense mucus layer, which prevents adherence to the epithelium, and the tight junctions, connecting epithelial cells and controlling access to the underlying tissues, are the physical barriers protecting the mucosal surfaces. Such physical protection is aided by pH differences and antimicrobial substances (biochemical barriers) and by the action of the innate and adaptive immune systems (immunological barriers) [2]. The innate immune system is naturally present in the organism and is the first

line of defense to respond quickly to bacteria and antigens. It recognizes a limited number of evolutionarily conserved molecules and does not retain a memory of a previous response. On the other hand, the adaptive immune system is acquired during lifetime upon the exposure to pathogens, is highly specific and retains a memory of a previous response. Both systems cooperate to efficiently recognize and eliminate pathogens. The immune system is, in part, located along the mucosal surfaces (mucosal immune system) and has an important role in immune surveillance.

The mucosal immune response is coordinated by the mucosa-associated lymphoid tissue (MALT) diffused in various submucosal sites. The MALT is composed of DC, macrophages, intraepithelial T cells (CD8+), regulatory T cells (Treg), and plasma cells that are organized into inductive sites, where antigens are recognized, and effector sites where the adaptive immune responses are mediated (Figure 1) [9, 10]. The MALT is commonly subdivided into mucosa-specific lymphoid tissues, for example, the gut-associated lymphoid tissue (GALT) or the nasal-associated lymphoid tissue (NALT). In the intestine, a large number of immune cells are found beneath the Peyer's patches (PP) that are formed by groups of M cells (specialized phagocytic cells with high transcytotic activity) [10]. At the level of the PP, antigens are transported from the intestinal lumen across the intestinal barrier and are taken up by the DCs and presented to naïve T cells in the local mesenteric lymph nodes [11]. In this district, the activated DCs promote naïve T cell differentiation into distinct T helper cells (Th1, Th2, or Th17) or T reg cells by secreting different types of pro-inflammatory cytokines (Figure 1a). In a health situation, DCs produce IL-10 and TGF- β inducing tolerance; thus DCs have a dual function: they either boost the immune system or dampen it, leading to tolerance and maintenance of the immune homeostasis. In a stimulus-dependent manner, IL-2, IL-18 and INF- γ induce a Th1 response, IL-4 induces a Th2 response, and IL-6 and IL-23 induce a Th17 response [9]. In turn, the T cells interact with B cells to promote antibody production at multiple mucosal sites. In particular, activated B cells produce antigen-specific IgG and secretory IgA (sIgA) antibodies [9, 10].

Although the mucosal tissues of the body share some common features, each tissue has a peculiar structure (epithelia, mucus, lymphoid structures, and resident immune cells) and a specific commensal microbiota. These specificities affect the nature of the immune inductive sites (GALT and mesenteric lymph nodes in the intestine, NALT and Cervical lymph nodes in the nose), the type of immune response, and its duration. Therefore, all these factors affect the mucosal immune response and are relevant for the design of a mucosal vaccine. In this context, it is essential that, although mucosal immune responses are compartmentalized (GALT, NALT, etc), the various mucosae are connected and, therefore, the immunization at a single site promotes immune responses at distant mucosal sites [12, 13]. A full understanding of the crosstalk between the various mucosal sites is critical to design novel vaccines that can potentially target mucosae distant from the vaccination site.

3. Mucosal adjuvants and delivery systems

The poor immunogenicity of current mucosal vaccines is primarily due to the lack of appropriate adjuvants and delivery systems. Consequently, antigens are exposed to the harsh environmental conditions of the mucosa and are degraded before they can be recognized by mucosal APCs (including DCs, macrophages, B cells) [14]. Furthermore, typical adjuvants used in parenteral vaccinations, for example, aluminum hydroxide (alum), complete Freund's adjuvant, and incomplete Freund's adjuvant, do not successfully stimulate mucosal immune responses and therefore, the future of mucosal vaccines necessarily relies on the use of effective mucosal adjuvant and delivery systems.

3.1. Mucosal adjuvants

A vaccine adjuvant is a molecule or particle able to activate innate immunity by inducing the production of proinflammatory molecules, chemokines, and cytokines by APCs. In recent years, various mucosal adjuvants have been proposed and extensively reviewed [8, 14]. Adjuvants proposed for mucosal vaccinations include modified bacterial toxins, flagellin, and other immunomodulatory molecules of bacterial or cellular origin [14]. The best characterized mucosal adjuvants are non-toxic variants of bacterial enterotoxins, in particular of the cholera toxin (CT) of *Vibrio cholerae* and of the heat-labile toxins (LT) of enterotoxigenic strains of *Escherichia coli* (ETEC). The multiple mutant

cholera toxin (mmCT) and the double mutant heat-labile toxin (dmLT), carrying respectively multiple mutations in the A subunit of the CT or two amino acid replacements (R192G and L211A) in LT, have low or no toxicity but retain the adjuvant activity of the full toxins and have been shown to potentiate the immune responses of various experimental mucosal vaccines against pathogens, such as *Streptococcus pneumoniae*, *Helicobacter pylori*, *V. cholera* and ETEC [14, 15]. Also, flagellin, the main structural component of the bacterial flagella, has been proposed as a mucosal adjuvant. Orally dosed flagellin of *Salmonella typhimurium* induces an inflammatory response by interacting with the Toll-Like receptor 5 (TLR5) and was shown to potentiate the immune response induced by mucosally administered bacterial and viral antigens [14]. In addition to enterotoxins and flagellin, other molecules have been proposed as mucosal adjuvants, including enterocyte binding proteins (for example, InlA of *Listeria monocytogenes* and FnBPA of *Staphylococcus aureus*) [16], cell surface proteins from the protozoan *Giardia lamblia*, M cell-targeting peptides [17], DC-targeting molecules [18], cytokine-derived molecules and the Fc region of antibodies [19]. Immune potentiator molecules, such as cytokines, have also been proposed as mucosal adjuvants, particularly for vaccines based on live-attenuated pathogens and/or live vectors (non-pathogenic microorganisms modified to express heterologous antigens) [20]. Interferon- γ (IFN- γ), IL-1 β , IL-12, IL-6, IL-10 and IL-12 have all been tested as adjuvants of live-attenuated vaccines against a variety of infectious diseases [20].

Quiescent spores of *Bacillus toyonensis*, a widely used animal probiotic, have been shown to act as parenteral [21] and mucosal [22] adjuvant of vaccines against a *Clostridium perfringens* infection and the tetanus toxin, respectively.

3.2. Mucosal delivery systems

Various approaches have been used to deliver antigens to mucosal surfaces, including live cells, bacterial-derived vesicles, viruses, virus-like particles and synthetic materials. All these techniques rely either on the discovery and optimization of proteins/peptides to be used as antigens (subunit vaccines) or on the delivery of DNA/RNA coding for antigens (genetic vaccines) [23]. For subunit vaccines, antigens can be either produced *in vitro* and later formulated in vaccine preparations or their coding genes can be used to obtain antigen synthesis *in vivo* by a live vaccine carrier. Several pathogenic and non-pathogenic bacteria have been tested as live vehicles of heterologous antigens [24, 25]. Initially, epitopes of the hepatitis B virus [26], of the cholera toxin [27], or of the parasite *Plasmodium falciparum* [28] were displayed on the surface of non-virulent strains of the pathogenic bacterium *Salmonella*. Antigen-specific immune responses were observed when the recombinant bacterial cells were used for the mucosal immunization of mice, providing clear evidence that an attenuated bacterium can be effectively used as a vaccine vehicle to deliver heterologous antigens [27, 28]. Later on, also non-pathogenic, commensal bacteria were used to display and deliver antigens to mucosal surfaces to avoid the use of engineered, attenuated pathogens [29].

More recently, bacterial-derived materials have been proposed as mucosal vaccine vehicles. In this context, outer membrane vesicles (OMVs), non-living structures deriving from gram-negative bacteria, stimulated particular interest [30]. OMVs are formed during bacterial growth by either a spontaneous or induced budding of the outer membrane. OMVs obtained from gram-negative pathogens contain periplasmic material and molecules normally exposed on the outer membrane, and a parenteral vaccine against *Neisseria meningitidis* based on OMVs has already been licensed [30]. OMVs can also be obtained from non-pathogenic, gram-negative bacteria genetically engineered to express a bacterial or non-bacterial antigen on the cell surface. Recently, a nasally-delivered vaccine based on OMVs was shown to induce high mucosal and systemic protection against a SARS-CoV-2 infection [31].

A variety of either natural or synthetic molecules have been proposed to deliver and protect antigens at specific mucosal sites [8, 14, 23, 32]. These include micro- or nano-particles made of poly-anhydrides (PHAs), poly(ethylene-glycol), poly(lactic acid), chitin, alginate, and several other polymers that are not cytotoxic, biocompatible, mucoadhesive and that, by different techniques, are used to coat antigens, therefore protecting them and increasing their immunogenicity [14, 32].

The use of liposomes to deliver antigens to the mucosal surfaces has also attracted much interest in the possibility of combining the delivery with the adjuvant effect [33]. Liposomes are micro- or nano-metric lipid bilayers that can contain or expose antigens and be constructed to have an immunomodulatory activity that can induce innate and adaptive responses [33].

Self-assembling protein nanoparticles, such as virus-like particles or nanoparticles displaying antigens, have been shown to facilitate antigen uptake and presentation and are widely applied in developing new vaccines. However, their use has been so far mainly focused on parenteral vaccinations [34, 35].

Various viral vectors, including adenovirus, influenza virus, Newcastle disease virus (NDV), and other paramyxoviruses, have been proposed as vaccine vectors, mainly for parenteral immunizations but more recently also for mucosal ones [36].

In addition, death or quiescent cells have been proposed as mucosal vaccine vectors to live bacteria, bacterial or viral-derived particles, and synthetic materials. An example of dead cells is bacterial ghosts (BGs), empty bacterial envelopes of gram-negative bacteria that do not have nucleic acids. BGs attracted the scientific community's attention for the possibility of using highly immunogenic pathogens, also genetically engineered, to express additional antigens or adjuvants on their surface without the risk associated with the live cells [37]. BGs differ from classical heat- or chemically-inactivated pathogens that have long been used as vaccines for their preparation methods. Classical methods to produce inactivated vaccines were based on formaldehyde or heat treatments that can damage the surface structures (antigens) and reduce the immunogenicity of the vaccine. BGs can be prepared by genetic (expression of lytic enzymes, phage proteins, or antimicrobial peptides) or by chemical (treatments with NaOH, SDS or H₂O₂) approaches, all able to strongly reduce viability without altering the surface structures of the cells [37].

The best-characterized example of quiescent cells used as vaccine delivery systems is the bacterial (endo)spore produced by the gram-positive bacterium *Bacillus subtilis*. A genetically engineered spore expressing the C fragment of the tetanus toxin on its surface [38], induced a protective, mucosal and systemic immune response [39]. Over the years, several other antigens have been displayed on the spore of *B. subtilis* as well as on spores of other *Bacillus* species [25, 40] and this spore-display system will be discussed in detail in the following paragraphs.

4. The *Bacillus* spore as mucosal vaccine vehicles

4.1. The *Bacillus* spore

The *B. subtilis* spore, like that produced by most members of the *Bacillus* and *Clostridium* genera, is a particularly stable and resistant cell formed in the cytoplasm of a vegetative cell when the environmental conditions no longer allow cell growth and/or survival (Figure 2A) [41]. The released spore can survive indefinitely in the absence of water and nutrients and in the presence of toxic chemicals, lytic enzymes, and extremes of temperature and pH [41]. The quiescent spore responds to the renewed presence of nutrients and favourable conditions by germinating and, thus, originating vegetative cells able to grow and eventually sporulate again (Figure 2A) [42].

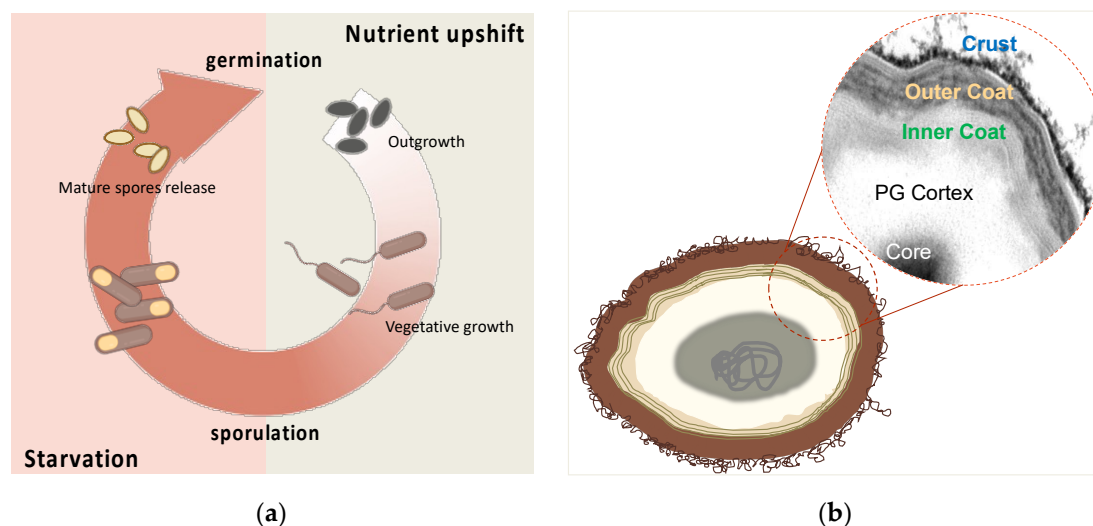


Figure 2. Schematic view of sporulation process (A) and spore structure (B) of *B. subtilis*. (A) In harsh conditions, such as nutrient depletion, the vegetative cells start the alternative cycle of sporulation. Mature released spores can remain dormant for an indefinite period before they germinate and resume the vegetative cycle. Adapted from Mutlu (2018) [42]. (B) Cartoon of a typical *B. subtilis* spore. Spore structural layers of the spore protect the genome contained in the partially dehydrated core. The dotted circle shows a detail of TEM micrograph of a spore stained with ruthenium red.

The spore stability and resistance are in part due to its peculiar structure. A partially dehydrated cytoplasm contains a copy of the chromosome and forms the spore interior (core), which is surrounded and protected by a thick peptidoglycan-like cortex, a multilayered, proteinaceous coat and, in *B. subtilis*, a crust made of proteins and glycoproteins (Figure 2B). Other spore-forming species either do not have an additional layer outside the coat or have an exosporium, a balloon-like structure also made of glycoproteins that loosely surrounds the spore and mediates its interactions with the environment [43]. Proteins and glycoproteins on the outermost spore layers [44, 45] make the spore surface negatively charged and relatively hydrophilic [46]. Some spore surface proteins self-assemble around the spore [47, 48, 49], forming remarkably robust structures [50].

In addition to a survival strategy, producing a spore is also a successful mechanism for the dispersal of these organisms on Earth. Spores are found in almost every environmental niche, including the gut of terrestrial and aquatic animals [51, 52]. Seminal experiments with a murine model have shown that ingested spores of *B. subtilis* safely transit the stomach, germinate and proliferate in the upper part of the intestine [53] and that re-sporulate in the lower part of the intestine [54]. In the GIT, *B. subtilis* spores and germination-derived cells interact with intestinal epithelial and immune cells, contributing to the normal development of the GALT [55, 56] and protecting the host from enteropathogens [57]. Also, based on these properties, spores of several *Bacillus* species are widely commercialized as probiotic preparations for animals and humans [58, 59].

4.2. The spore delivery systems: recombinant approach

The spore structure and stability suggested its use as a platform to display heterologous molecules [38]. In a *proof-of-concept* study, the coat protein CotB of *B. subtilis* was selected as a carrier for the spore display of a model passenger protein, the C fragment of the tetanus toxin (TTFC) of *C. tetani* [38]. DNA coding for TTFC was fused in frame with the *cotB* gene and inserted on the *B. subtilis* chromosome under the transcription and translation signals of *cotB*, thus ensuring genetic stability and proper expression of the chimera [38]. An average of 1.5×10^3 TTFC molecules per spore were displayed [38], and mucosally administered spores were able to induce an antigen-specific immune response in mice [39].

The gene fusion, integrated on the chromosome and expressed under the transcription and translation signals of the anchor protein, produces a chimera in the mother cell. The chimera is then

driven on the spore surface by the spore surface protein used as an anchor thus leading to the release of a mature spore fully decorated with the selected antigen (Figure 3A).

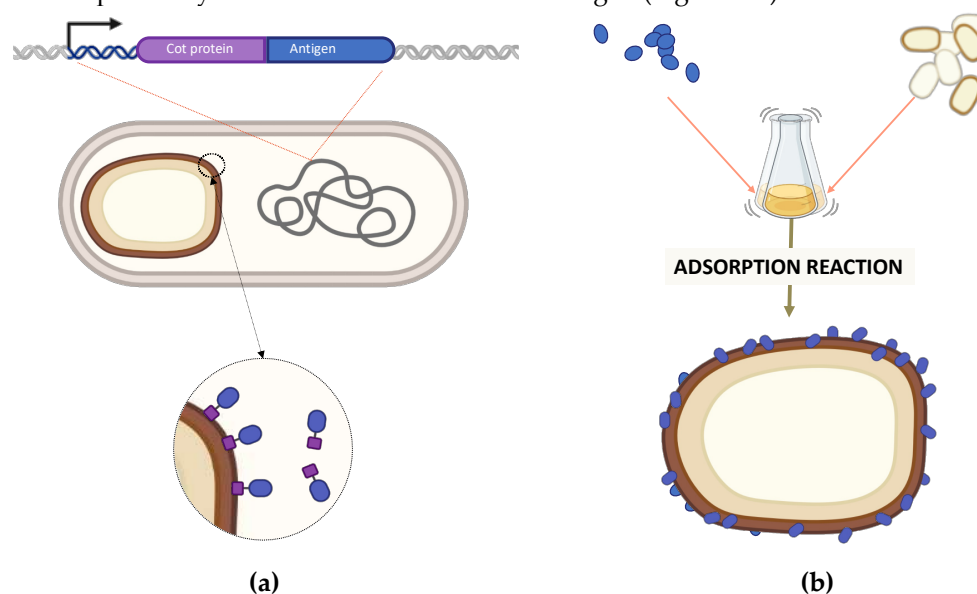


Figure 3. Strategy of recombinant (A) and non-recombinant (B) spore-surface display. (A) A gene fusion is constructed between DNA coding for a coat protein (violet) and for an antigen (blue). The fusion is under the transcriptional and translational signals of the spore surface gene and is integrated into *B. subtilis* chromosome. During sporulation, the recombinant protein is expressed in the mother cell and assembled on the forming spore. (B) Antigens and purified spores are reacted in an acidic reaction buffer. After 1 hour of incubation, antigens are bound on the spore surface.

The same recombinant approach has been used to display a variety of antigens fused to CotB and several other coat proteins as an anchor (Table 2). Sets of either replicative [88] or integrative [89] plasmids have been developed to facilitate the construction of gene fusions. The integrative vectors, allowing the integration of the gene fusions on the *B. subtilis* chromosome, grant a better genetic stability over replicative plasmids.

Table 2. Coat proteins of *B. subtilis* proposed as carriers to display antigens.

Carriers	Antigens	References
CotB	TTFC of <i>Clostridium tetani</i>	38
	LTB of <i>Escherichia coli</i>	60
	FliD of <i>Clostridium difficile</i>	61, 62
	PA of <i>Bacillus anthracis</i>	63
	UreA of <i>Helicobacter acinonychis</i>	64
	TcdA-TcdB of <i>Clostridium difficile</i>	65
	Cpa of <i>Clostridium perfringens</i>	66
	VP28 of White Spot Syndrome Virus	67, 68
	M2 protein of influenza virus	69
	SlpA of <i>Lactobacillus brevis</i>	70
	InvA of <i>Yersinia pseudotuberculosis</i>	70
	MPT64 of <i>Mycobacterium tuberculosis</i>	71
	BclA3 of <i>Clostridium difficile</i>	72
	VP1 capsid protein of Enterovirus 71	73
	HR2P of SARS-CoV-2 spike	74
PCV2 Cap protein of <i>Porcine circovirus</i>	75	
Vp7 of grass carp reovirus	76	

CotC	TTFC of <i>Clostridium tetani</i>	38
	LTB of <i>Escherichia coli</i>	60
	FliD of <i>Clostridium difficile</i>	61, 62
	PA of <i>Bacillus anthracis</i>	63
	UreA of <i>Helicobacter acinonychis</i>	64
	TcdA-TcdB of <i>Clostridium difficile</i>	65
	UreB of <i>Helicobacter pylori</i>	77
	TP22.3 of <i>Clonorchis sinensis</i>	78
	CsSerp3 of <i>Clonorchis sinensis</i>	79
	Pep23 of HIV	80
	GST of <i>Schistosoma japonicum</i>	81
	GP64 of <i>Bombyx mori</i>	82
	Enolase of <i>Clonorchis sinensis</i>	83
	Paramyosin of <i>Clonorchis sinensis</i>	84
	OmpC of <i>Salmonella serovar Pullorum</i>	85
	VP4 of Grass carp reovirus	86
	VP56 of Grass carp reovirus	87
Vp26 of White spot syndrome virus	67, 88	
Vp7 of grass carp reovirus	76	
MCP of Nervous necrosis virus (RGNNV)	89	
CotG	UreA of <i>Helicobacter acinonychis</i>	64
	FliD of <i>Clostridium difficile</i>	61, 62
CotY	OmpK of <i>Vibrio vulnificus</i>	90
	RBD of SARS-CoV-2 spike	91
CotZ	FliD of <i>Clostridium difficile</i>	61, 62
	UreA of <i>Helicobacter acinonychis</i>	64
	RBD of SARS-CoV-2 spike	91
CgeA	CagA of <i>Helicobacter pylori</i>	92

Spores of species other than *B. subtilis* have also been considered for displaying heterologous proteins [93, 94] but so far, not yet tested for expressing antigens.

Recombinant spores have also been proposed as vaccine vehicles for veterinary uses [66, 95], and for aquaculture [86, 87].

Alternative recombinant approaches to display proteins on spores have also been proposed. For example, enzymes [96] and the protective antigen of *B. anthracis* [97] were over-expressed in the mother cell of sporulating cells of *B. subtilis*, and part of the highly concentrated heterologous proteins spontaneously adsorbed around the forming spores, decorating the released spore. This approach is not based on using a spore surface protein as an anchor but rather on the spontaneous attachment of highly concentrated proteins on the spore surface. This concept was previously exploited as a non-recombinant spore display [98, 99] and is discussed below.

More recently, the basement layer of the *B. subtilis* coat has been reconstituted around spherical membranes supported by silica beads. Such artificial spore-like particles (synthetic spore husk-encased lipid bilayers, SSHELs) were covalently bound to small molecules and suggested as a versatile display platform for drugs, antigens and enzymes [100].

4.3. The spore delivery systems: non-recombinant approach

The non-recombinant spore display system (spore-adsorption) has been recently reviewed [99]. It is based on the spontaneous and stable binding of a purified heterologous protein to the spore surface (Fig. 3B). This approach has been used to adsorb on the *B. subtilis* spore various enzymes [99] and model antigens, such as TTFC of *C. tetani*, PA of *B. anthracis*, Cpa of *C. perfringens* [98], B subunit of the heat-labile toxin (LTB) of *E. coli* [12] and BclA2 of *C. difficile* [101].

It is known that at acidic conditions (pH 3.0-4.0) proteins highly concentrated outside the spore spontaneously and tightly bind to the spore surface but the molecular mechanism of spore adsorption is still not fully understood. Experiment using the Red Fluorescent Protein (RFP) of the coral *Disco-soma* sp. as a model protein revealed that adsorbed molecules cross the outermost surface layer of the *B. subtilis* [102] or *B. megaterium* [103] spores. The outermost spore layer of most/all spore former species is characterized by the presence of cysteine-rich proteins that self-assemble into hexameric complexes producing a protein lattice permeated by pores (Fig. 4) [47, 48, 49, 96]. Such pores mediate the spore permeability to germinants [104] and, in *B. subtilis*, are controlled by the spore surface protein CotG [105].

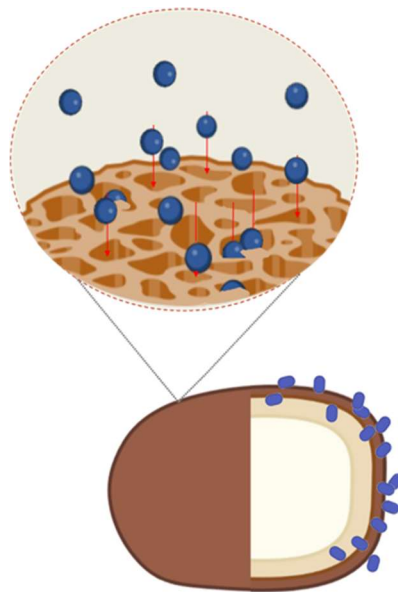


Figure 4. Simplified model of spore-adsorption mechanism. Adsorbed antigens infiltrate through the pores formed by spore surface coat proteins in a size depending-way. The physicochemical properties of the antigens have not been considered in the model.

A model proposed to explain spore adsorption suggests that adsorbed protein infiltrates through the pores present in the outermost layer and localizes between the outermost layer (exosporium or outer coat) and the immediately underneath layer (Fig. 4B) [96].

It has been reported that small proteins often adsorb more efficiently than big proteins, but this trend is not always followed, suggesting that the permeability of the pores depends on the size of the adsorbed protein but also on the physico-chemical properties of both the heterologous protein and the spore [96]. The isoelectric point, the electric charge, and the relative hydrophobicity of the molecule to be adsorbed have been suggested as relevant for efficient adsorption [98; 106]. The same heterologous proteins are adsorbed with different efficiencies by spores of different species [103] or by different strains of the same species [107], or even by spores of the same strain but grown at different conditions [108], suggesting that the different physicochemical properties of the spore surface influence the efficiency of adsorption.

According to the proposed model, the adsorbed proteins would be mainly localized inside the spore and only in a minimal part exposed on the spore surface [96]. The internal localization of the adsorbed proteins would then explain the tight adhesion and the higher stability and resistance to unfavorable conditions of adsorbed over free proteins [96].

In some cases, the standard spore-adsorption procedure (Fig. 3B) has been used in a modified version. Heat-inactivated spores of *B. subtilis* have been successfully used to bind influenza H5N1 virions (NIBRG-14; clade 1) [109] or a *Mycobacterium tuberculosis* antigen [110], introducing the concept of using spores as inert displaying bioparticles.

4.4. Recombinant vs. non-recombinant spore display

Both spore display approaches have advantages and disadvantages and the preference for one or the other system has to be analyzed case-by-case. The recombinant approach has the clear advantage that the antigen does not have to be produced and purified. The recombinant bacterium contains the gene coding for the antigen fused to a spore surface protein; therefore, the sporulating cell produces the antigen to be displayed, reducing the costs and simplifying the production process. On the other end, the non-recombinant system has the obvious advantage of being non-recombinant, thus it does not raise safety concerns related to the use and environmental release of recombinant spores.

In addition to these, the non-recombinant approach is significantly more efficient than the recombinant system. Isticato et al. [12] compared the efficiency of the two display approaches by using the same antigen, LTB of *E. coli*. An average of 9.6×10^{-5} pg per spore of the CotC-LTB fusion protein was displayed by a strain carrying a *cotC::eltB* gene fusion [60], while up to 2.5×10^{-3} pg of LTB per spore were adsorbed to wild type *B. subtilis* spores [12]. This about 25-fold increase of displayed LTB becomes even larger (up to 100-fold increase) by using mutant spores altered in the spore surface [12]. A significantly increased efficiency of display is particularly relevant for a vaccine delivery system, since it allows to reduce either the number of spores/dose or the number of doses needed to deliver a sufficient amount of antigen for the induction of an antigen-specific immune response.

An additional relevant advantage of the non-recombinant system is that multimeric antigens are presented in their mature conformation. LTB of *E. coli* is a pentamer that only in its native form is functional and binds its receptor, the GM1 ganglioside, on the enterocyte surface [111]. When expressed as a fusion protein on *B. subtilis* spores [60] or on the surface of *S. gordonii* [112], LTB is displayed as a monomer, while LTB pentamers are displayed by the non-recombinant system on *B. subtilis* spores [12].

However, the non-recombinant spore display system also has disadvantages: i) displayed antigens are not on the spore surface and can be presented to the immune cells only after that the spore germinates or is destroyed in the animal body; ii) the molecular mechanisms of spore adsorption are still not fully understood. According to the model of Fig. 4B, antigens would accumulate within the spore surface layers in a disordered way, thus impairing the construction of precise structures, with epitopes exposed in the most convenient way for interaction with the immune cells.

5. Mucosal immunizations with recombinant and non-recombinant spores

Spores that do not display any heterologous antigen induce low levels of spore-specific IgG response when are mucosally (orally or nasally) administered to mice [55]. While they are not recognized by TLR2 and TLR4, the two principal Toll-like receptors sensing live bacteria, and seem unable to interact with B cells, spores stimulate the induction of IFN- γ and other mediators of a cellular response [113]. In addition, orally administered spores have been shown to reduce the susceptibility to enteric pathogens in animal models. D'Arienzo et al. [57] used *Citrobacter rodentium*, a mouse pathogen causing epithelial lesions similar to those caused by human enteropathogenic and enterohemorrhagic strains of *E. coli* [114], to show the protective effects of *B. subtilis* spores. In this infection model, a treatment with 1×10^9 spores one day before the infection with *C. rodentium* drastically decreased colon colonization, prevented the enteropathy by reducing crypt length and epithelial damages, and significantly reduced the mortality rate [57]. In other studies, a single oral inoculum of spores suppressed all signs of *Escherichia coli* O78:K80, *Salmonella enterica* or *Clostridium perfringens* infections in chickens [115, 116].

Spores displaying heterologous antigens by the recombinant approach have been shown to induce antigen-specific immune responses in mice orally or nasally immunized. In this context, the best-characterized example is that of spores displaying the C-terminus of the tetanus toxin (TTFC) fused to the spore surface protein CotB of *B. subtilis*. Mice orally immunized with those recombinant spores produced TTFC-specific antibodies (fecal sIgA and serum IgG) with a IgG isotype profile indicating a prevalence of IgG1 and IgG2b and, therefore, a Th2-type immune response [39]. When challenged with the purified tetanus toxin, the immunized animals were fully protected,

demonstrating the potential of spores as a valuable delivery system for mucosal vaccines [39]. The same spores also induced a significant TTFC-specific IgA and IgG response with a prevalence of IgG1 and IgG2b indicative of a Th2 biased immune response when we're used to orally prime mice that were then subcutaneously boosted with soluble TTFC (without adjuvant) [117]. When orally administered, the same spores were also shown to induce a cellular immune response in Balb/C mice with spleen and mesenteric lymph nodes (MLN) cell proliferation as well as production of IFN- γ but not of IL-4 and IL-10 in both districts [118]. When the CotB-TTFC chimera was displayed on the surface of spores of a mutant strain of *B. subtilis* unable to germinate, similar levels of cell proliferation and a similar pattern of cytokine induction were observed with respect to those observed with wild-type spores, indicating that the observed antigen-specific cellular immune response was independent from spore germination in the GIT and was only due to the antigen exposed on the orally ingested spores [118]. The same conclusion was reached also using a different antigen, the MPT64 of *Mycobacterium tuberculosis* [71]. In this case, spores displaying the antigen fused to CotB were heat-inactivated and were still able to induce an immune response reducing the pathogen load in the animal lungs and inducing the secretion of Th1 cytokines [71].

Other examples of antigens displayed on spores as fusion proteins and able to induce a strong immune response when mucosally administered are the LTB of *E. coli* [60], the C-terminus of the alpha toxoid of *Clostridium perfringens* (Cpa) [110] and the BclA₃ of *C. difficile* [72]. In the LTB and Cpa cases, high levels of antigen-specific IgG and sIgA were induced [60, 66], and recombinant spores displaying Cpa fused to CotB protected the orally or nasally immunized mice against a 12 LD₅₀ challenge dose of alpha toxin [66]. Also, spore-displayed BclA3 induced antibody production in mice and attenuated some *C. difficile* infection symptoms after a challenge with the pathogen but was less efficient than the free antigen [72].

The effects of some antigens delivered by spore adsorption have also been characterized. Huang et al. [98] reported that purified Cpa of *C. perfringens* mixed to *B. subtilis* spores induced an immune response indistinguishable from that induced by recombinant spores displaying Cpa fused to CotB [98]. Mice were immunized with either three oral doses of spores mixed with 3.6 μ g of Cpa or a single nasal dose of spores mixed with 0.15 μ g of Cpa. Protection was obtained in nasally dosed mice to a 6 LD₅₀ dose of toxin, while for oral dosing, only one mouse survived, suggesting that with the non-recombinant approach, the nasal route is preferable to the oral one [98]. In the same study, spores were also mixed with TTFC of *C. tetani* and PA of *B. anthracis* and in all cases, a Th1-biased immune response was observed and the immunized animals were protected against the challenge with the purified toxins [98]. In all cases, heat-inactivated spores appeared equally effective as live spores [98]. This remarkable result was further exploited, showing that killed spores adsorbed to inactivated influenza virions (H5N1; NIBRG-14; clade 1) induce both humoral and cell-mediated immune responses when intra-nasally administered to mice [109]. In a challenge experiment, mice nasally dosed two times with spores adsorbed with 20 ng hemagglutinin of inactivated NIBRG-14 were fully protected against challenge with 20 LD₅₀ of H5N2 virus [109]. Humoral and cellular immune responses were also observed in mice nasally immunized with spores adsorbed with LTB of *E. coli* [12]. Production of fecal and serum sIgA, serum IgG and IFN- γ was by both spleen and MLN cells of mice immunized with spore-adsorbed LTB was observed at levels statistically higher than those observed by immunizing mice with purified LTB, an effect that could be related to an increased antigen uptake by competent immune cells or, alternatively, to a reduced antigen degradation [12]. An increased antigen stability was observed when the spore surface protein BclA2 of *C. difficile* was adsorbed to *B. subtilis* spores [101]. In addition, spores adsorbed with BclA2 showed an increased adherence to human intestinal (Caco-2) cells *in vitro* and induced antigen-specific antibody production in nasally immunized mice [101].

Spore-adsorbed TTFC, when nasally administered to mice, was more efficient than the free antigen in inducing fecal sIgA, serum IgG and the cytokines IL-6 and IL-12 [22]. The efficiency of the nasal vaccination was further improved by oral probiotic treatment with *B. toyonensis* [22]. In this case, both the humoral and the cellular immune responses were enhanced by the probiotic treatment

without significantly altering the gut's microbial composition, pointing to the probiotic treatment as an alternative to the use of adjuvants for mucosal vaccinations [22].

6. Future perspectives

Both recombinant and non-recombinant systems of spore display are potentially efficient strategies to deliver heterologous antigens to mucosal surfaces. Each system has advantages and disadvantages leaving the preference to one or the other to a case-by-case analysis. An exciting future perspective is the combined use of both systems. Examples are the use of recombinant spores displaying a protein able either to act as adjuvant [119] or specifically target the spore to a tissue or cell type [120] that can be adsorbed with a different molecule. In this context, *B. subtilis* spores displaying the adjuvant IL-2 or streptavidin as a chimeric fusion were adsorbed with the purified antigen FliD of *C. difficile* [119] or the diterpen paclitaxel, a mitotic inhibitor used in cancer therapy [120]. By this approach spores displaying IL-2 showed an increased immune response [119] and those displaying streptavidin can bind any biotinylated antibody, potentially targeting spores and adsorbed molecules to any potential target cell therapy [120].

Most of the experiments so far performed to display antigens have been carried out with laboratory collection strains. An additional, intriguing future perspective is to display antigens on spores of probiotic strains of *B. subtilis* or other *Bacillus* strains. Several strains of spore formers species are widely used as probiotics for human or animal use [58, 59] and the possibility of using spores of probiotic strains to display antigens would allow to combine the beneficial probiotic effects to the induction of an antigen-specific immune response.

Author Contributions: “Conceptualization, all Authors; writing—original draft preparation, A.S. and E.R.; writing—review and editing, L.B.; G.D.; R.I.; funding acquisition, E.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: All data are reported in the papers indicated as References.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Pashine, A.; Valiante, N.M.; Ulmer, JB. Targeting the innate immune response with improved vaccine adjuvants. *Nat Med* **2005**, *11*:S63–8.
2. Correa, VA.; Portilho, A.I.; De Gaspari, E. Vaccines, adjuvants and key factors for mucosal immune response. *Immunology* **2022**, *167*(2):124–138.
3. Zhang, L.; Wang, W.; Wang, S. Effect of vaccine administration modality on immunogenicity and efficacy. *Expert Rev Vaccines* **2015**, *14*:1509–23.
4. Lycke, N.; Bemark, M. Mucosal adjuvants and long-term memory development with special focus on CTA1-DD and other ADP-ribosylating toxins. *Mucosal Immunol* **2010**, *3*:556–66.
5. Bernasconi, V.; Norling, K.; Bally, M.; Höök, F.; Lycke, NY. Mucosal vaccine development based on liposome technology. *J Immunol Res* **2016**, *2016*:5482087.
6. Criscuolo, E.; Caputo, V.; Diotti, RA.; Sautto, GA.; Kirchenbaum, GA.; Clementi, N. Alternative methods of vaccine delivery: an overview of edible and intradermal vaccines. *J Immunol Res* **2019**, *2019*:8303648.
7. Lavelle, EC.; Ward, RW. Mucosal vaccines - fortifying the frontiers. *Nature Rev Immunol* **2022**, *22*:237–250.
8. Huang, M.; Zhang, M.; Zhu, H.; Du, X.; Wnag, J. Mucosal vaccine delivery: a focus on the breakthrough of specific barriers. *Acta Pharm Sinica B* **2022**, *12*:6453–3474.
9. Traxinger, BR.; Richert-Spuhler, LE.; Lund, JM. Mucosal tissue regulatory T cells are integral in balancing immunity and tolerance at portals of antigen entry. *Mucosal Immunol* **2021**, *15* (3):398–407.
10. Mörbe, UM.; Jørgensen, PB.; Fenton, TM.; Von Burg, N.; Riis, LB.; Spencer, J.; Agace, WW. Human gut-associated lymphoid tissues (GALT); diversity, structure, and function. *Mucosal Immunol* **2021**, *14* (4):793–802.
11. Tordesillas, L.; Berin, MC. Mechanisms of oral tolerance. *Clin Rev Allergy Immunol* **2018**, *55*(2):107–117.

12. Isticato, R.; Sirec, T.; Treppiccione, L.; Maurano, F.; De Felice, M.; Rossi, M.; Ricca, E. Non-recombinant display of the B subunit of the heat labile toxin of *Escherichia coli* on wild type and mutant spores of *Bacillus subtilis*. *Microb Cell Fact* **2013**, *29*:98.
13. Van Splutter, M.; Van Hoffen, E.; Floris-Vollenbroek, EG.; Timmerman, H.; Lucas van de Bos, E.; Meijer, B.; Ulfman, LH.; Witteman, B.; Wells, JM.; Brugman, S.; Savelkoul, HFJ.; Van Neerven, RJJ. Oral cholera vaccination promotes homing of IgA+ memory B cells to the large intestine and the respiratory tract. *Mucosal Immunol* **2018**, *11*:1254-1264.
14. Ou, B.; Yang, Y.; Lin, X.; Zhang, M. Current progress and challenges in the study of adjuvants for oral vaccines. *BioDrugs* **2023**, *37*:143-180.
15. Lycke, N.; Lebrero-Fernandez C. ADP-ribosylating enterotoxins as vaccine adjuvants. *Curr Opin Pharmacol* **2018**, *41*:42-51.
16. Innocentin, S.; Guimares, V.; Miyoshi, A.; Azevedo, V.; Langella, P.; Chatel, J-M. *Lactococcus lactis* expressing either *Staphylococcus aureus* fibronectin-binding protein A or *Listeria monocytogenes* internalin A can efficiently internalize and deliver DNA in human epithelial cells. *Appl Environ Microbiol* **2009**, *75*(14):4870-8.
17. Kim, S-H.; Seo, K-W.; Kim, J.; Lee, K-Y.; Jang, Y-S. The M cell-targeting ligand promotes antigen delivery and induces antigen-specific immune responses in mucosal vaccination. *J Immunol* **2010**, *185*(10):5787-95.
18. Owen, JL.; Sahay, B.; Mohamadzadeh, M. New generation of oral mucosal vaccines targeting dendritic cells. *Curr Opin Chem Biol* **2013**, *17*(6):918-24.
19. Ye, L.; Zeng, R.; Bai, Y.; Roopenian, DC.; Zhu, X. Efficient mucosal vaccination mediated by the neonatal Fc receptor. *Nat Biotechnol* **2011**, *29*(2):158-63.
20. Manohar, MM.; Campbell, BE.; Walduck AK, Moore RJ. Enhancement of live vaccines by co-delivery of immune modulating proteins. *Vaccine* **2022**, *40*:5769-5780.
21. Santos, FDS.; Maubrigades, LR.; Gonsalves, VS.; Franz, HC.; Rodrigues, PRC.; Cunha, RC.; Leite, FPL. *Bacillus toyonensis* BCT-7112 spores as parenteral adjuvant of BoHV-5 vaccine in a murine model. *Probiot Antimicrob Prot* **2021**, *13*:655-663.
22. Santos, FDS.; Mazzoli, A.; Maia, AR.; Saggese, A.; Isticato, R.; Leite, F.; Iossa, S.; Ricca, E.; Baccigalupi, L. A probiotic treatment increases the immune response induced by the nasal delivery of spore-adsorbed TTFC. *Microb Cell Fact* **2020**, *19*:42.
23. Huang, S.; Zhu, Y.; Zhang, L.; Zhang, Z. Recent advances in delivery systems for genetic and other novel vaccines. *Adv Mater* **2022**, *34*:2107946.
24. Benhar, I. 2001. Biotechnological applications of phage and cell display. *Biotechnol Adv* **19**:1-33.
25. Isticato, R.; Ricca, E. Spore surface display. *Microbiol Spect* **2014**, *2*(5): TBS-0011-2012.
26. Wu, JY.; Newton, S.; Judd, A.; Stocker, B.; Robinson, WS. Expression of immunogenic epitopes of hepatitis B surface antigen with hybrid flagellin proteins by a vaccine strain of *Salmonella*. *Proc Natl Acad Sci USA* **1989**, *86*:4726-4730.
27. Newton, SM.; Jacob, CO.; Stocker, BA. Immune response to cholera toxin epitope inserted in *Salmonella* flagellin. *Science* **1989**, *244*:70-72.
28. Schorr, J.; Knapp, B.; Hundt, E.; Kupper, HA.; Amann, E. Surface expression of malarial antigens in *Salmonella typhimurium*: induction of serum antibody response upon oral vaccination of mice. *Vaccine* **1991**, *9*:675-681.
29. Fischetti, VA.; Medaglini, D.; Pozzi, G. Gram-positive commensal bacteria for mucosal vaccine delivery. *Curr Opin Biotechnol* **1996**, *7*:659-666.
30. Lieberman, LA. Outer membrane vesicles: a bacterial-derived vaccination system. *Front Microbiol* **2022**, *13*:1029146.
31. Van der Ley, P. A.; Zariri, A.; Van Riet, E.; Oosterhoff, D.; Kruiswijk, C.P. An intranasal OMV-based vaccine induces high mucosal and systemic protecting immunity against a SARS-CoV-2 infection. *Front Immunol* **2021**, *12*:5303.
32. Haddad, HF.; Roe, EF.; Collier, JH. Expanding opportunities to engineer mucosal vaccination with biomaterials. *Biomater Sci* **2023**, *11*:1625-1647.
33. Bernasconi, V.; Norling, K.; Bally, M.; Hook, F.; Lycke NY. Mucosal vaccine development based on liposome technology. *J Immunol Res* **2016**, 5482087.
34. Wu, F.; Qin, M.; Wang, H.; Sun, X. Nanovaccines to combat virus-related diseases. *WIREs Nanomed Nanobiotechnol* **2023**, *15*: e1857.
35. He, J.; Yu, L.; Lin, X.; Liu, X.; Zhang, Y.; Yang, F.; Deng, W. Virus-like particles as nanocarriers for intracellular delivery of biomolecules and compounds. *Viruses* **2022**, *14*:1905.
36. Garcia-Sastre, A. Mucosal delivery of RNA vaccines by Newcastle disease virus vectors. *Curr Res Immunol* **2022**, *3*:234-238.
37. Chen, H.; Ji, H.; Kong, X.; Lei, P.; Yang, Q.; Wu, W.; Jin, L.; Sun, D. Bacterial ghosts-based vaccine and drug delivery. *Pharmaceutics* **2021**, *13*:1892.
38. Isticato, R.; Cangiano, G.; Tran, T-H.; Ciabattini, A.; Medaglini, D.; Oggioni, MR.; De Felice, M.; Pozzi, G.; Ricca, E. Surface display of recombinant proteins on *Bacillus subtilis* spores. *J Bacteriol* **2001**, *183*:6294-6301.

39. Duc, LH.; Huynh, HA.; Fairweather, N.; Ricca, E.; Cutting, SM. Bacterial Spores as Vaccine Vehicles. *Infect&Immun* **2003**, *71*:2810-2818.
40. Ricca, E.; Baccigalupi, L.; Cangiano, G.; De Felice, M.; Istatico, R. Mucosal vaccine delivery by non-recombinant spores of *Bacillus subtilis*. *Microb Cell Fact* **2014**, *13*(1):115.
41. McKenney, PT.; Driks, A, Eichenberger, P. The *Bacillus subtilis* endospore: assembly and function of the multilayered coat. *Nat Rev Microbiol* **2013**, *11*:33-44.
42. Mutlu, A.; Trauth, S.; Ziesack, M.; Nagler, K.; Bergeest, J. P.; Rohr, K.; Becker, N.; Höfer, T.; Bischofs, I. B. Phenotypic memory in *Bacillus subtilis* links dormancy entry and exit by a spore quantity-quality tradeoff. *Nat Commun* **2018**, *9*(1): 69.
43. Paredes-Sabja, D.; Cid-Rojas, F.; Pizarro-Guajardo, M. Assembly of the exosporium layer in *Clostridioides difficile* spores. *Curr Poin Microbiol* **2022**, *67*:102137.
44. Cangiano, G.; Sirec, T.; Panarella, C.; Istatico, R.; Baccigalupi, L.; De Felice, M.; Ricca, E. The *sps* gene products affect germination, hydrophobicity and protein adsorption of *Bacillus subtilis* spores. *Appl Environ Microbiol* **2014**, *80*:7293–7302.
45. Dubois, T.; Krzewinski, F.; Yamakawa, N.; Lemy, C.; Hamiot, A.; Brunet, L.; Lacoste, AS.; Knirel, Y.; Guerardel, Y.; Faille C. The *sps* genes encode an original legionaminic acid pathway required for crust assembly in *Bacillus subtilis*. *mBio* **2020**, *11*: e01153-20.
46. Pesce, G.; Rusciano, G.; Sirec, T.; Istatico, R.; Sasso, A.; Ricca, E. Surface charge and hydrodynamic coefficient measurements of *Bacillus subtilis* spore by optical tweezers. *Colloids Surf B Biointerfaces* **2014**, *116*:568–575.
47. Tang, J.; Krajcikova, D.; Zhu, R.; Ebner, A.; Cutting, S.; Gruber, HJ.; Barak, I.; Hinterdorfer, P. Atomic force microscopy imaging and single molecule recognition force spectroscopy of coat proteins on the surface of *Bacillus subtilis* spore. *J Mol Recognit* **2007**, *20*:483–489.
48. Ramamurthi, KS.; Losick, R. ATP-driven self-assembly of a morphogenetic protein in *Bacillus subtilis*. *Mol Cell* **2008**, *31*:406–414.
49. Jiang, S.; Wan, Q.; Krajcikova, D.; Tang, J.; Tzokov, SB.; Barak, I.; Bullogh, PA. Diverse supramolecular structures formed by self-assembling proteins of the *Bacillus subtilis* spore coat. *Molec Microbiol* **2015**, *97*:347–359.
50. Janganan, TK.; Mullin, N.; Dafis-Sagarmendi, A.; Brunt, J.; Tzokov, SB.; Stringer, S.; Moir, A.; Chaudhuri, RR.; Fagan, RP.; Hobbs, JK.; Bullogh, PA. Architecture and self-assembly of *Clostridium sporogenes* and *Clostridium botulinum* spore surfaces illustrate a general protective strategy across spore formers. *mSphere* **2020**, *5*: e00424-20.
51. Fakhry, S.; Sorrentini, I.; Ricca, E.; De Felice, M.; Baccigalupi, L. Characterization of spore forming Bacilli isolated from the human gastrointestinal tract. *J Applied Microbiol* **2008**, *105*:2178-2186.
52. Hong, HA.; To, E.; Fakhry, S.; Baccigalupi, L.; Ricca, E.; Cutting, SM. Defining the natural habitat of *Bacillus* spore-formers. *Res Microbiol* **2009**, *160*:375-379.
53. Casula, G.; Cutting, SM. *Bacillus* probiotics: spore germination in the gastrointestinal tract. *Appl Environ Microbiol* **2002**, *68*:2344–2352.
54. Duc, LH.; Hong, AH.; Nguyen, QU.; Cutting, SM. Intracellular fate and immunogenicity of *B. subtilis* spores. *Vaccine* **2004**, *22*:1873–1885.
55. Cutting, SM.; Hong, HA.; Baccigalupi, L.; Ricca, E. Oral vaccine delivery by recombinant spore probiotics. *Intern Rev Immunol* **2009**, *28*:487–505.
56. Rhee, K-J.; Sethupathi, P.; Driks, A.; Lanning, DK.; Knight, KL. Role of commensal bacteria in development of gut-associated lymphoid tissue and preimmune antibody repertoire. *J Immunol* **2004**, *172*:1118–1124.
57. D'Arienzo, R.; Maurano, F.; Mazzarella, G.; Luongo, D.; Stefanile, R.; Ricca, E.; Rossi, M. *Bacillus subtilis* spores reduce susceptibility to *Citrobacter rodentium*-mediated enteropathy in a mouse model. *Res Microbiol* **2006**, *157*, 891–897.
58. Cutting, SM. *Bacillus* probiotics. *Food Microbiol* **2011**, *28*:214–220.
59. Saggese, A.; Baccigalupi, L.; Ricca, E. Spore formers as beneficial microbes for humans and animals. *Appl Microbiol* **2021**, *1*:498-509.
60. Mauriello, EMF.; Duc, LH.; Istatico, R.; Cangiano, G.; Hong, HA.; De Felice, M.; Ricca, E.; Cutting, SM. Display of heterologous antigens on the *Bacillus subtilis* spore coat using CotC as a fusion partner. *Vaccine* **2004**, *22*:1177–1187.
61. Negri, A.; Potocki, W.; Iwanicki, A.; Obuchowski, M.; Hinc, K. Expression and display of *Clostridium difficile* protein FliD on the surface of *Bacillus subtilis* spores. *J Med Microbiol* **2013**, *62*:1379-1385. doi: 10.1099/jmm.0.057372-0.
62. Piekarska, A.; Pełka, P.; Peszyńska-Sularz, G.; Negri, A.; Hinc, K.; Obuchowski, M.; Iwanicki, A. The choice of the anchoring protein influences the interaction of recombinant *Bacillus* spores with the immune system. *Acta Biochim Pol* **2017**, *64*(2):239-244. doi: 10.18388/abp.2016_1315.

63. Duc, L.H.; Hong, H.A.; Atkins, H.S.; Flick-Smith, H.C.; Durrani, Z.; Rijpkema, S.; Titball, R.W.; Cutting SM. Immunization against anthrax using *Bacillus subtilis* spores expressing the anthrax protective antigen. *Vaccine* **2007**, *25*:346–355.
64. Hinc, K.; Isticato, R.; Dembek, M.; Karczewska, J.; Iwanicki, A.; Peszyńska-Sularz, G.; De Felice, M.; Obuchowski, M.; Ricca, E. Expression and display of UreA of *Helicobacter acinonychis* on the surface of *Bacillus subtilis* spores. *Microb Cell Fact* **2010**, *18*:9:2. doi: 10.1186/1475-2859-9-2.
65. Permpoonpattana, P.; Hong, H.A.; Phetcharaburanin, J.; Huang, J.M.; Cook, J.; Fairweather, N.; Cutting, S.M. Immunization with *Bacillus* spores expressing toxin A peptide repeats protects against infection with *Clostridium difficile*. *Infect Immun* **2011**, *79*:2295–2302.
66. Hoang, T.H.; Hong, H.A.; Clark, G.C.; Titball, R.W.; Cutting S.M. Recombinant *Bacillus subtilis* expressing the *Clostridium perfringens* alpha toxoid is a candidate orally delivered vaccine against necrotic enteritis. *Infect Immun* **2008**, *76*:5257–5265.
67. Ning, D.; Leng, X.; Li, Q.; Xu W. Surface-displayed VP28 on *Bacillus subtilis* spores induces protection against white spot syndrome virus in crayfish by oral administration. *J Appl Microbiol* **2011**, *111*:1327–1336.
68. Nguyen, A.T.; Pham, C.K.; Pham, H.T.; Pham, H.L.; Nguyen, A.H.; Dang, L.T.; Huynh, H.A.; Cutting, S.M.; Phan, T.N. *Bacillus subtilis* spores expressing the VP28 antigen: a potential oral treatment to protect *Litopenaeus vannamei* against white spot syndrome. *FEMS Microbiol Lett* **2014**, *358*(2):202-8. doi: 10.1111/1574-6968.12546.
69. Zhao, G.; Miao, Y.; Guo, Y.; Qiu, H.; Sun, S.; Kou, Z.; Yu, H.; Li, J.; Chen, Y.; Jiang, S.; Du, L.; Zhou, Y. Development of a heat-stable and orally delivered recombinant M2e-expressing *B. subtilis* spore-based influenza vaccine. *Hum Vaccin Immunother* **2014**, *10*(12):3649-58. doi: 10.4161/hv.36122.
70. Tavares Batista, M.; Souza, R.D.; Paccez, J.D.; Luiz, W.B.; Ferreira, E.L.; Cavalcante, R.C.; Ferreira, R.C.; Ferreira, L.C. Gut adhesive *Bacillus subtilis* spores as a platform for mucosal delivery of antigens. *Infect Immun* **2014**, *82*(4):1414-23. doi: 10.1128/IAI.01255-13.
71. Sibley, L.; Reljic, R.; Radford, D.S.; Huang, J.M.; Hong, H.A.; Cranenburgh, R.M. Recombinant spores expressing MPT64 evaluated as a vaccine against tuberculosis in the murine model. *FEMS Microbiol Lett* **2014**, *358*(2): 170-179.
72. Maia, A.R.; Reyes-Ramírez, R.; Pizarro-Guajardo, M.; Saggese, A.; Ricca, E.; Baccigalupi, L.; Paredes-Sabja, D. Nasal Immunization with the C-Terminal Domain of BclA3 Induced Specific IgG Production and Attenuated Disease Symptoms in Mice Infected with *Clostridioides difficile* Spores. *Int J Mol Sci* **2020**, *13*:21(18):6696. doi: 10.3390/ijms21186696.
73. Cao, Y.-G.; Li, Z.-H.; Yue, Y.-Y.; Song, N.-N.; Peng, L.; Wang, L.-X. Construction and evaluation of a novel spores-based enterovirus 71 vaccine. *J Appl Biomed* **2013**, 105-113.
74. Hen, C.; Li, Y.L.; Lv, F.L.; Xu, L.D.; Huang, Y.W. Surface Display of Peptides Corresponding to the Heptad Repeat 2 Domain of the Feline Enteric Coronavirus Spike Protein on *Bacillus subtilis* Spores Elicits Protective Immune Responses Against Homologous Infection in a Feline Aminopeptidase-N-Transduced Mouse Model. *Front Immunol* **2022**, *28*:13: 925922. doi: 10.3389/fimmu.2022.925922.
75. Sun, R.; Zhang, M.; Chen, H.; Wei, Y.; Ning, D. Germination-Arrest *Bacillus subtilis* Spores as An Oral Delivery Vehicle of Grass Carp Reovirus (GCRV) Vp7 Antigen Augment Protective Immunity in Grass Carp (*Ctenopharyngodon idella*). *Genes* **2020**, *11*(11):1351. doi: 10.3390/genes11111351.
76. Li, W.; Li, J.; Dai, X.; Liu, M.; Khalique, A.; Wang, Z.; Zeng, Y.; Zhang, D.; Ni, X.; Zeng, D.; Jing, B.; Pan, K. Surface Display of porcine circovirus type 2 antigen protein cap on the spores of *Bacillus subtilis* 168: An effective mucosal vaccine candidate. *Front Immunol* **2022**, *13*:1007202. doi: 10.3389/fimmu.2022.1007202.
77. Zhou, Z.; Dong, H.; Huang, Y.; Yao, S.; Liang, B.; Xie, Y.; Long, Y.; Mai, J.; Gong S. Recombinant *Bacillus subtilis* spores expressing cholera toxin B subunit and *Helicobacter pylori* urease B confer protection against *H. pylori* in mice. *J Med Microbiol* **2017**, *66*(1):83-89. doi: 10.1099/jmm.0.000404.
78. Zhou, Z.; Xia, H.; Hu, X.; Huang, Y.; Li, Y.; Li, L.; Ma, C.; Chen, X.; Hu, F.; Xu, J.; Lu, F.; Wu, Z.; Yu, X. Oral administration of a *Bacillus subtilis* spore-based vaccine expressing *Clonorchis sinensis* tegumental protein 22.3 kDa confers protection against *Clonorchis sinensis*. *Vaccine* **2008**, *28*:26(15):1817-25. doi: 10.1016/j.vaccine.2008.02.015.
79. Lin, Z.; Sun, H.; Ma, Y.; Zhou, X.; Jiang, H.; Wang, X.; Song, J.; Tang, Z.; Bian, Q.; Zhang, Z.; Huang, Y.; Yu, X. Evaluation of immune response to *Bacillus subtilis* spores expressing *Clonorchis sinensis* serpin3. *Parasitology* **2020**, *147*(10):1080-1087. doi: 10.1017/S0031182020000797.
80. D'Apice, L.; Sartorius, R.; Caivano, A.; Mascolo, D.; Del Pozzo, G.; Scotto Di Mase, D.; Ricca, E.; Li Pira, G.; Manca, F.; Malanga, D.; De Palma, R.; De Berardinis, P. Comparative analysis of new innovative vaccine formulations based on the use of procaryotic display systems. *Vaccine* **2007**, *25*:1993–2000.
81. Li, L.; Hu, X.; Wu, Z.; Xiong, S.; Zhou, Z.; Wang, X.; Xu, J.; Lu, F.; Yu, X. Immunogenicity of self-adjunctivity oral vaccine candidate based on use of *Bacillus subtilis* spore displaying *Schistosoma japonicum* 26 KDa GST protein. *Parasitol Res* **2009**, *105*(6):1643-51. doi: 10.1007/s00436-009-1606-7.
82. Li, G.; Tang, Q.; Chen, H.; Yao, Q.; Ning, D.; Chen, K. Display of *Bombyx mori* nucleopolyhedrovirus GP64 on the *Bacillus subtilis* spore coat. *Curr Microbiol* **2011**, *62*(5):1368-73. doi: 10.1007/s00284-011-9867-7.

83. Wang, X.; Chen, W.; Tian, Y.; Mao, Q.; Lv, X.; Shang, M.; Li, X.; Yu, X.; Huang, Y. Surface display of *Clonorchis sinensis* enolase on *Bacillus subtilis* spores potentializes an oral vaccine candidate. *Vaccine* **2014**, *10*;32(12):1338-45. doi: 10.1016/j.vaccine.2014.01.039.
84. Sun, H.; Lin, Z.; Zhao, L.; Chen, T.; Shang, M.; Jiang, H.; Tang, Z.; Zhou, X.; Shi, M.; Zhou, L.; Ren, P.; Qu, H.; Lin, J.; Li, X.; Xu, J.; Huang, Y.; Yu, X. *Bacillus subtilis* spore with surface display of paramyosin from *Clonorchis sinensis* potentializes a promising oral vaccine candidate. *Parasit Vectors* **2018**, *7*;11(1):156. doi: 10.1186/s13071-018-2757-0.
85. Dai, X.; Liu, M.; Pan, K.; Yang, J. Surface display of OmpC of *Salmonella serovar Pullorum* on *Bacillus subtilis* spores. *PLoS One* **2018**, *25*;13(1): e 0191627. doi: 10.1371/journal.pone.0191627.
86. Jiang, H.; Bian, Q.; Zeng, W.; Ren, P.; Sun, H.; Lin, Z.; Tang, Z.; Zhou, X.; Wang, Q.; Wang, Y.; Wang, Y.; Wu, MX.; Li, X.; Yu, X.; Huang, Y. Oral delivery of *Bacillus subtilis* spores expressing grass carp reovirus VP4 protein produces protection against grass carp reovirus infection. *Fish Shellfish Immunol* **2019**, *84*:768-780. doi: 10.1016/j.fsi.2018.10.008.
87. Gao, Y.; Huo, X.; Wang, Z.; Yuan, G.; Liu, X.; Ai, T.; Su, J. Oral Administration of *Bacillus subtilis* Subunit Vaccine Significantly Enhances the Immune Protection of Grass Carp against GCRV-II Infection. *Viruses* **2021**, *14*(1):30. doi: 10.3390/v14010030.
88. Valdez, A.; Yepiz-Plascencia, G.; Ricca, E.; Olmos, J. First *Litopenaeus vannamei* WSSV 100% oral vaccination protection using CotC::Vp26 fusion protein displayed on *Bacillus subtilis* spores surface. *J Appl Microbiol* **2014**, *117*(2):347-57. doi: 10.1111/jam.12550.
89. Weijun, Mai.; Bingjun, Yan.; Jian, Xin. Oral immunizations with *Bacillus subtilis* spores expressing MCP protein provide protection against red-spotted grouper nervous necrosis virus (RGNNV) infection in juvenile grouper, *Epinephelus coioides*. *Aquaculture* **2022**, *552*,738008. <https://doi.org/10.1016/j.aquaculture.2022.738008>.
90. Gonçalves, G.; Santos, RA.; Coutinho, F.; Pedrosa, N.; Curado, M.; Machado, M.; Costas, B.; Bonneville, L.; Serrano, M.; Carvalho, AP.; Díaz-Rosales, P.; Oliva-Teles, A.; Couto, A.; Serra CR. Oral vaccination of fish against vibriosis using spore-display technology. *Front Immunol* **2022**, *13*;13:1012301. doi: 10.3389/fimmu.2022.1012301.
91. Vetráková, A.; Chovanová, RK.; Rechteríková, R.; Krajčíková, D.; Barák, I. *Bacillus subtilis* spores displaying RBD domain of SARS-CoV-2 spike protein. *Comput Struct Biotechnol J* **2023**, *21*:1550-1556. doi: 10.1016/j.csbj.2023.02.007.
92. Iwanicki, A.; Piątek, I.; Stasiłojć, M.; Grela, A.; Łęga, T.; Obuchowski, M.; Hinc, K. A system of vectors for *Bacillus subtilis* spore surface display. *Microb Cell Fact* **2014**, *13*:30.
93. Du, C.; Chan, WC.; McKeithan, W.; Nickerson, KW. Surface display of recombinant proteins on *Bacillus thuringiensis* spores. *Appl Environ Microbiol* **2005**, *71*:3337-3341.
94. Park, TJ.; Choi, S-K.; Jung, H-C.; Lee, SY.; Pan, J-G. Spore display using *Bacillus thuringiensis* exosporium protein InhA. *J Microbiol Biotechnol* **2009**, *19*:495-501.
95. Lee, JE.; Kye, YC.; Park, SM.; Shim, BS.; Yoo, S.; Hwang, E.; Kim, H.; Kim, SJ.; Han, SH.; Park, TS.; Park, B-C.; Yun, C-H. *Bacillus subtilis* spores as adjuvants against avian influenza H9n2 induce antigen-specific antibody and T cell responses in white leghorn chickens. *Vet Res* **2020**, *51*(1):68. doi: 10.1186/s13567-020-00788-8.
96. Pan, JG.; Choim, SK.; Jung, HC.; Kim, EJ. 2014. Display of native proteins on *Bacillus subtilis* spores. *FEMS Microbiol Lett* **2014**, *358*:209-217.
97. Oh, Y.; Kim, JA.; Kim, C-H.; Choi, S-K.; Pan, J-G. *Bacillus subtilis* spore vaccines displaying protective antigen induce functional antibodies and protective potency. *BMC Veter Res* **2020**, *16*:259.
98. Huang, JM.; Hong, HA.; Van, Tong, H.; Hoang, TH.; Brisson, A.; Cutting, SM. Mucosal delivery of antigens using adsorption to bacterial spores. *Vaccine* **2010**, *28*:1021-1030.
99. Ricca, E.; Baccigalupi, L.; Istatico, R. Spore-adsorption: mechanism and applications of a non-recombinant display system. *Biotechnol Adv* **2021**, *47*:107693.
100. Wu, I.L.; Narayan, K.; Castaing, J.-P.; Tian, F.; Subramaniam, S.; Ramamurthi, K.S. A versatile nano display platform from bacterial spore coat proteins. *Nat Commun* **2015**, *6*, 6777.
101. Maia, A.R.; Reyes-Ramírez, R.; Pizarro-Guajardo, M.; Saggese, A.; Castro-Córdova, P.; Istatico, R.; Ricca, E.; Paredes-Sabja, D.; Baccigalupi, L. Induction of a Specific Humoral Immune Response by Nasal Delivery of Bcl2ctd of *Clostridioides difficile*. *Int J Mol Sci* **2020**, *21*, 1277. <https://doi.org/10.3390/ijms21041277>
102. Donadio, G.; Lanzilli, M.; Sirec, T.; Ricca, E.; Istatico, R. Localization of a red fluorescence protein adsorbed on wild type and mutant spores of *Bacillus subtilis*. *Microb Cell Fact* **2016**, *15*, 153. <https://doi.org/10.1186/s12934-016-0551-2>.
103. Lanzilli, M.; Donadio, G.; Addevico, R.; Saggese, A.; Cangiano, G.; Baccigalupi, L.; Christie, G.; Ricca, E.; Istatico, R. The Exosporium of *Bacillus megaterium* QM B1551 Is Permeable to the Red Fluorescence Protein of the Coral *Discosoma* sp. *Front Microbiol* **2016**, *7*:1752. doi: 10.3389/fmicb.2016.01752.
104. Setlow, P. Germination of spores of *Bacillus* species: what we know and do not know. *J Bacteriol* **2014**, *196*(7):1297-305. doi: 10.1128/JB.01455-13.

105. Saggese, A.; Di Gregorio Barletta, G.; Vittoria, M.; Donadio, G.; Istatico, R.; Baccigalupi, L.; Ricca, E. CotG Mediates Spore Surface Permeability in *Bacillus subtilis*. *mbio* **2022**, *13*(6): e0276022. doi: 10.1128/mbio.02760-22.
106. Sirec, T.; Strazzulli, A.; Istatico, R.; De Felice, M.; Moracci, M.; Ricca, E. Adsorption of β -galactosidase of *Alicyclobacillus acidocaldarius* on wild type and mutants spores of *Bacillus subtilis*. *Microb Cell Fact* **2012**, *11*:100. doi: 10.1186/1475-2859-11-100.
107. Sirec, T.; Cangiano, G.; Baccigalupi, L.; Ricca, E.; Istatico, R. The spore surface of intestinal isolates of *Bacillus subtilis*. *FEMS Microbiol Lett* **2014**, *358*(2):194-201. doi: 10.1111/1574-6968.12538.
108. Petrillo, C.; Castaldi, S.; Lanzilli, M.; Saggese, A.; Donadio, G.; Baccigalupi, L.; Ricca, E.; Istatico, R. The temperature of growth and sporulation modulates the efficiency of spore-display in *Bacillus subtilis*. *Microb Cell Fact* **2020**, *19*(1):185. doi: 10.1186/s12934-020-01446-6.
109. Song, M.; Hong, HA.; Huang, JM.; Colenutt, C.; Khang, DD.; Nguyen, TV.; Park, SM.; Shim, BS.; Song, HH.; Cheon, IS.; Jang, JE.; Choi, JA.; Choi, YK.; Stadler, K.; Cutting, SM. Killed *Bacillus subtilis* spores as a mucosal adjuvant for an H5N1 vaccine. *Vaccine* **2012**, *30*(22):3266-77. doi: 10.1016/j.vaccine.2012.03.016.
110. Reljic, R.; Sibley, L.; Huang, JM.; Pepponi, I.; Hoppe, A.; Hong, HA.; Cutting, SM. Mucosal vaccination against tuberculosis using inert bioparticles. *Infect Immun* **2013**, *81*(11):4071-80. doi: 10.1128/IAI.00786-13.
111. Kim, J-M.; Park, S-M.; Kim, J-A.; J-P, M-h Y.; Kim, N-S.; Bae, J-L.; Park, G-S.; Jang, J-S.; Yang, M-S.; Kim, D-H: Functional pentameric formation via coexpression of the *Escherichia coli* heat-labile enterotoxin B subunit and its fusion protein subunit with a Neutralizing Epitope of ApxIIA Exotoxin improves the mucosal immunogenicity and protection against challenge by *Actinobacillus pleuropneumoniae*. *Clin Vaccine Immunol* **2011**, *18*(12):2168-77. doi: 10.1128/CVI.05230-11.
112. Ricci, S.; Medaglini, D.; Rush, CM.; Marcello, A.; Peppoloni, S.; Manganelli, R.; Palú, G.; Pozzi, G. Immunogenicity of the B monomer of *Escherichia coli* heatlabile toxin expressed on the surface of *Streptococcus gordonii*. *Infect Immun* **2000**, *68*:760-766.
113. Huang, J.-M.; La Ragione, R.; Nunez, A.; Cutting, S.M. Immunostimulatory activity of *Bacillus* spores. *FEMS Immunol Med Microbiol* **2008**, *53*: 195-203.
114. Schauer, D.B.; Falkow, S. The *eae* gene of *Citrobacter freundii* biotype 4280 is necessary for colonization in transmissible murine colonic hyperplasia. *Infect Immun* **1993**, *61*: 4654-4661.
115. La Ragione, R.M.; Woodward, M.J. Competitive exclusion by *Bacillus subtilis* spores of *Salmonella enterica* serotype Enteritidis and *Clostridium perfringens* in young chickens. *Vet Microbiol* **2003**, *94*: 245-256.
116. La Ragione, R.M.; Casula, G.; Cutting, S.M.; Woodward, M.J. *Bacillus subtilis* spores competitively exclude *Escherichia coli* O78:K80 in poultry. *Vet Microbiol* **2001**, *79*:113-142.
117. Ciabattini, A.; Parigi, R.; Istatico, R.; Oggioni, M.; Pozzi, G. Oral priming of mice by recombinant spores of *Bacillus subtilis*. *Vaccine* **2004**, *22*: 4139-4143.
118. Mauriello, EM.; Cangiano, G.; Maurano, F.; Saggese, V.; De Felice, M.; Rossi, M.; Ricca, E. Germination-independent induction of cellular immune response by *Bacillus subtilis* spores displaying the C fragment of the tetanus toxin. *Vaccine* **2007**, *25*:788-793.
119. Potocki, W.; Negri, A.; Peszyńska-Sularz, G; Hinc, K.; Obuchowski, M.; Iwanicki, A. The combination of recombinant and non-recombinant *Bacillus subtilis* spore display technology for presentation of antigen and adjuvant on single spore. *Microb Cell Fact* **2017**, *16*, 151. <https://doi.org/10.1186/s12934-017-0765-y>.
120. Nguyen, VA.; Huynh, HA.; Hoang, TV.; Ninh, NT.; Pham, AT.; Nguyen, HA.; Phan, TN.; Cutting, SM. Killed *Bacillus subtilis* spores expressing streptavidin: a novel carrier of drugs to target cancer cells. *J Drug Target* **2013**, *21*:528-541.